

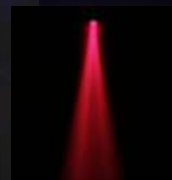


22nd - 27th May, 2017
Zakopane, Poland



LII Zakopane School of Physics

breaking frontiers:
submicron structures
in physics and biology



Laser-Matter Interaction (LMI) Group

Nanoimaging using soft X-ray and EUV sources

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wachulak@gmail.com



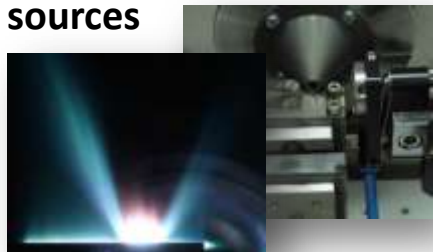
<http://www.ztl.wat.edu.pl/zoplzm>

LASER-MATTER INTERACTION LABORATORY

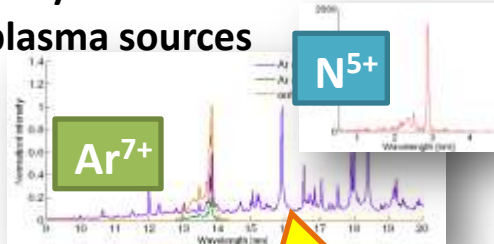
Institute of Optoelectronics,
Military University of Technology



Efficient generation of the
SXR/EUV high intensity
radiation, laser-plasma
sources



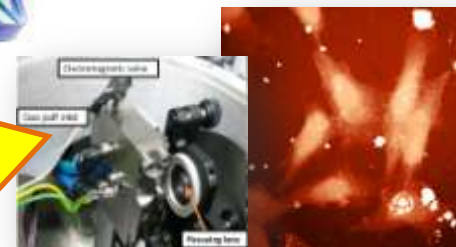
Generation of a monochromatic
EUV/SXR radiation from laser-
plasma sources



Radiobiology



Contact
microscopy

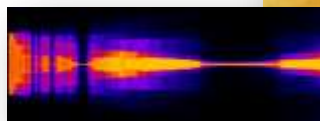


Laser radiation

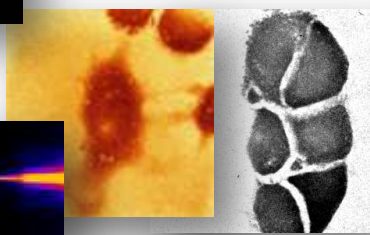
EUV light interactions
with gasses



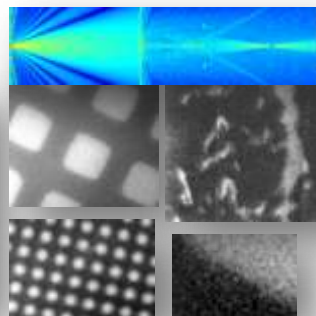
EUV photo-
ionization



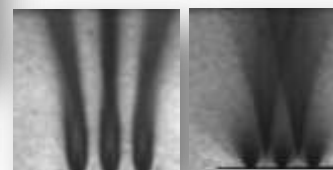
Water window
microscopy



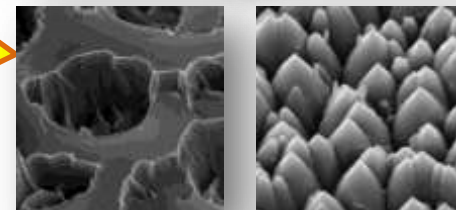
EUV high resolu-
tion imaging



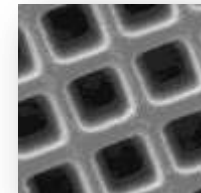
EUV/SXR
radiography
and tomography



Polymer surface modifi-
cation by EUV light



Polymer surface
processing using EUV

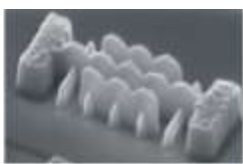


Outline

- Nanotechnology and nanoimaging
- EUV and SXR radiation, motivation and sources
- EUV holography (Gabor, Fourier)
- Reconstruction of CG holograms
- Talbot imaging
- Coherent diffraction (lens-less) imaging
- Ptychography
- Zone plates for various wavelengths
- EUV microscopy using FZP
- Scanning EUV microscopy
- SXR and „water-window“ microscopy
- EUV, SXR HXR tomography
- Contact microscopy



Quantum devices



Transistor gates



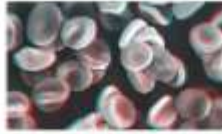
MEMS devices



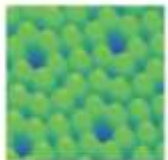
Proteins



Visible light



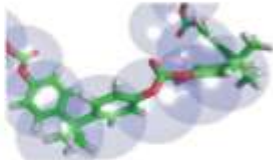
Cells



Atoms



Molecules



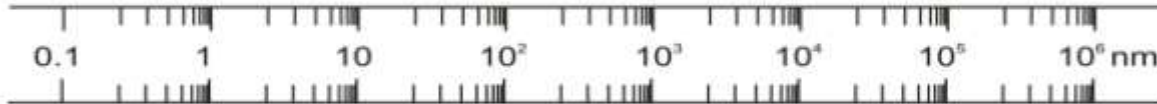
Polymers



Bacteria



Liquid drops



Nanotechnology:

Manipulation on objects like atoms and molecules and the observation of the results of such manipulation in the nanometer scale

„NANO”

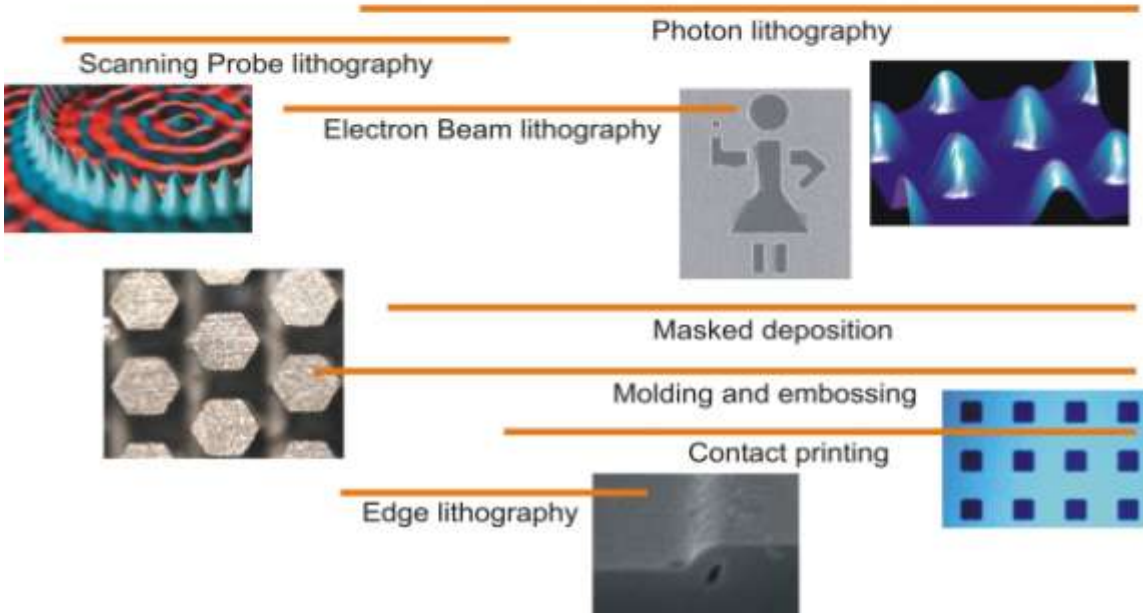
1nm =0.000 000 001m=
10 H atoms

Diameter of a human hair: 50 000nm

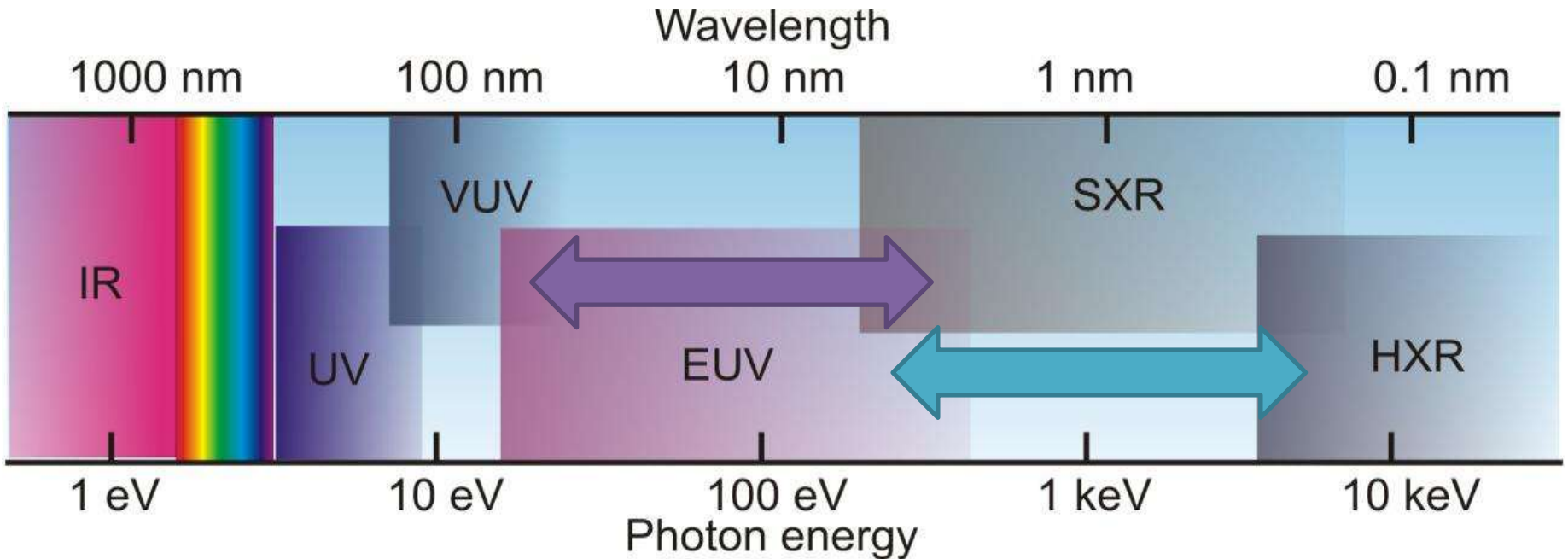
Smallest feature seen by a human eye: 10 000 nm

Nanoimaging:

Imaging in the nanometer scale: <1um or <100nm



Electromagnetic Spectrum



Adequate wavelengths of light:

Extreme Ultraviolet (EUV)

$\lambda \sim 10-120\text{nm}$

$E_{\text{ph}} \sim 10-120\text{eV}$

Soft X-rays (SXR)

$\lambda \sim 1\text{\AA}-10\text{nm}$

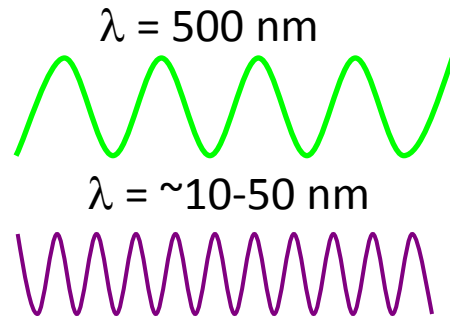
$E_{\text{ph}} \sim 120 - 10\text{keV}$

Motivation for EUV and SXR imaging

EUV ($\lambda=10-120\text{nm}$)

- shorter wavelength λ – better spatial resolution

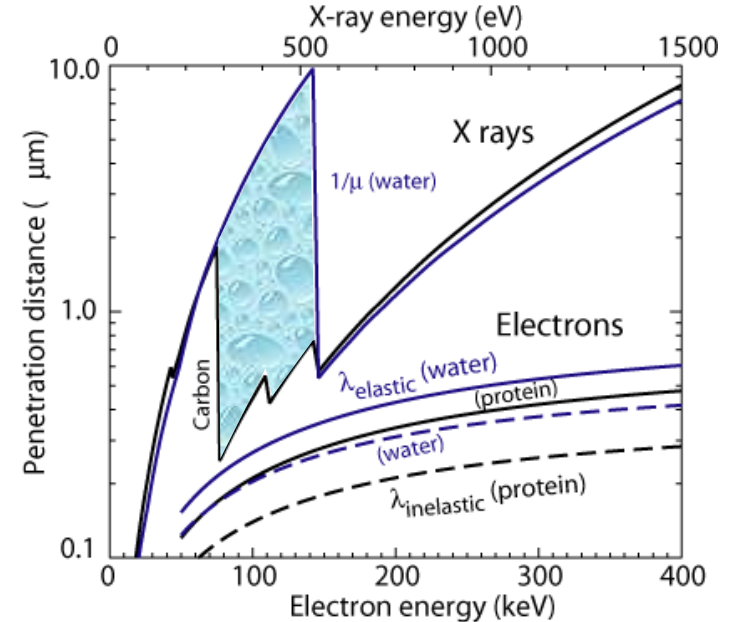
$$\delta = \frac{k \cdot \lambda}{NA}$$



EUV/SXR radiation „see” 10-100x smaller features

- strong optical contrast in the EUV; **very high absorption of EUV light in very thin layers**, practically almost all materials are opaque, including gasses, which allows for (**direct – based on light absorption - observation of gasses**).
- EUV light is used in lithography
- short EUV pulses allow to study transient processes

SXR ($\lambda=0.1-10\text{nm}$)

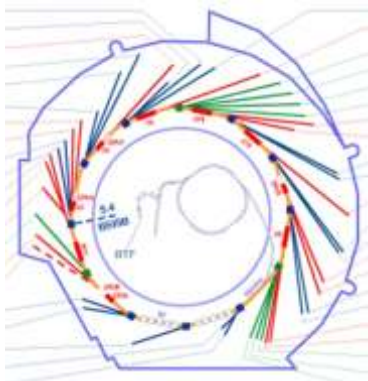


xray1.physics.sunysb.edu

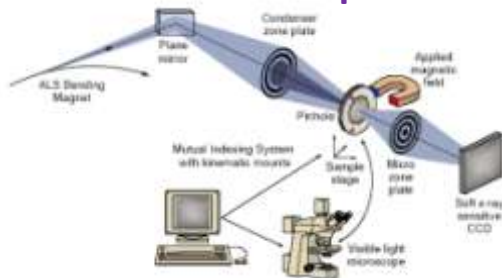
Water window: 284 - 543 eV, $\lambda = 2.3 - 4.4 \text{ nm}$

- short wavelength \rightarrow possible **high spatial resolution** : $\delta = k\lambda/NA$.
- very high **optical contrast between organic material (carbon based) and water (oxygen)**
- **radiation with relatively large penetration depth** tens of microns for biological samples.

Synchrotrons and FEL used for nanoimaging



XM-1 microscope

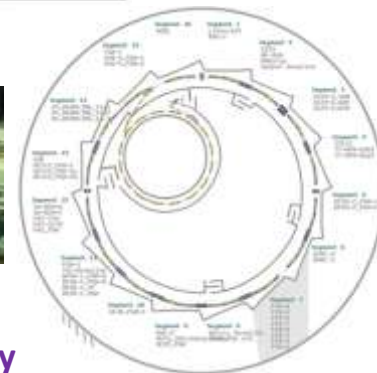


<http://www-als.lbl.gov/>



<http://www.lightsources.org/facility/hzb-helmholtz-zentrum-berlin>

BESSY II - Helmholtz-Zentrum Berlin (HZB)



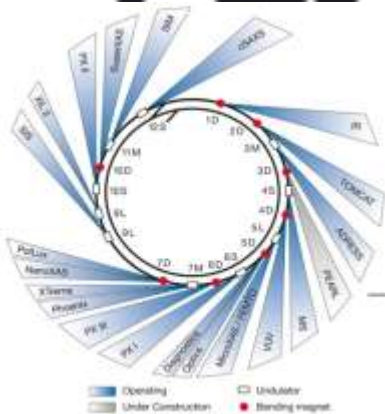
XM, X-ray Microscopy **U41-TXM**
X-ray-Microscopy, X-ray-Tomography



TOMCAT - X02DA: Tomography
A beamline for TOMographic
Microscopy and **C**oherent
radiology experimen**T**s

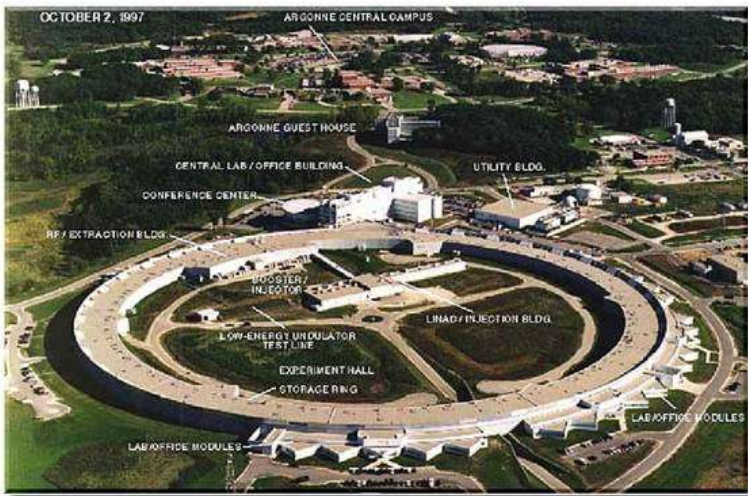
SIM - X11MA: Surfaces /
Interfaces: Microscopy

<http://www.psi.ch/sls/sim/sim>



Transmission X-ray
Microscopy
beamline
MISTRAL

<https://www.cells.es/Beamlines/XM/>



Advanced Photon Source, Argonne National Lab, USA

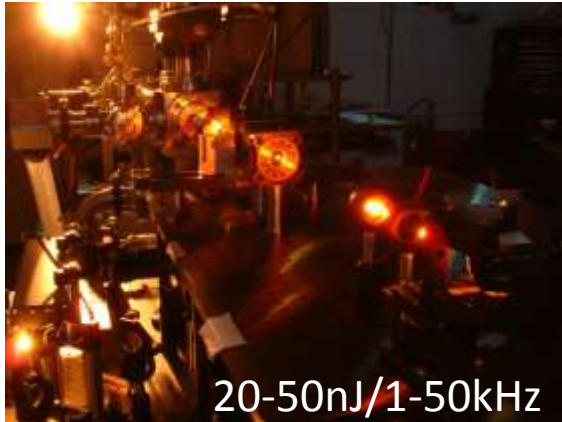


Stanford Linear Accelerator Center – SLAC and Linac Coherent Light Source - LCLS (USA)



Free Electron Laser (FEL), Hamburg, Germany

Compact sources



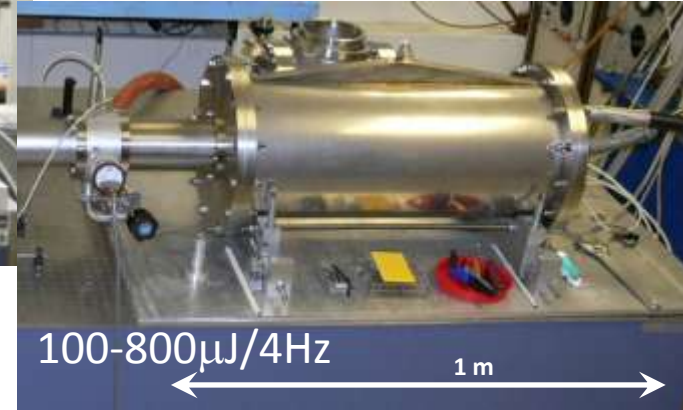
20-50nJ/1-50kHz

High Harmonic Generation (HHG) system,
James R., Macdonald Laboratory, Atomic,
Molecular & Optical Physics Research Facility,
Kansas State University, KS, USA



12 μ J/10Hz

Capillary discharge type EUV laser
(fast electric discharge in Ar filled
capillary), EUV ERC, Prof. Jorge Rocca,
Fort Collins, USA



100-800 μ J/4Hz

1 m



1.3 μ J/10Hz @13.8nm

2.3 μ J/10Hz @3.4nm

EUV/SXR lamp for metrology and
microscopy, Prof. Henryk Fiedorowicz,
IOE WAT, Warsaw, Poland

Compact sources:

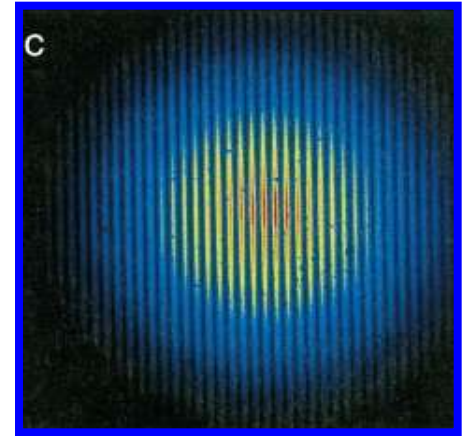
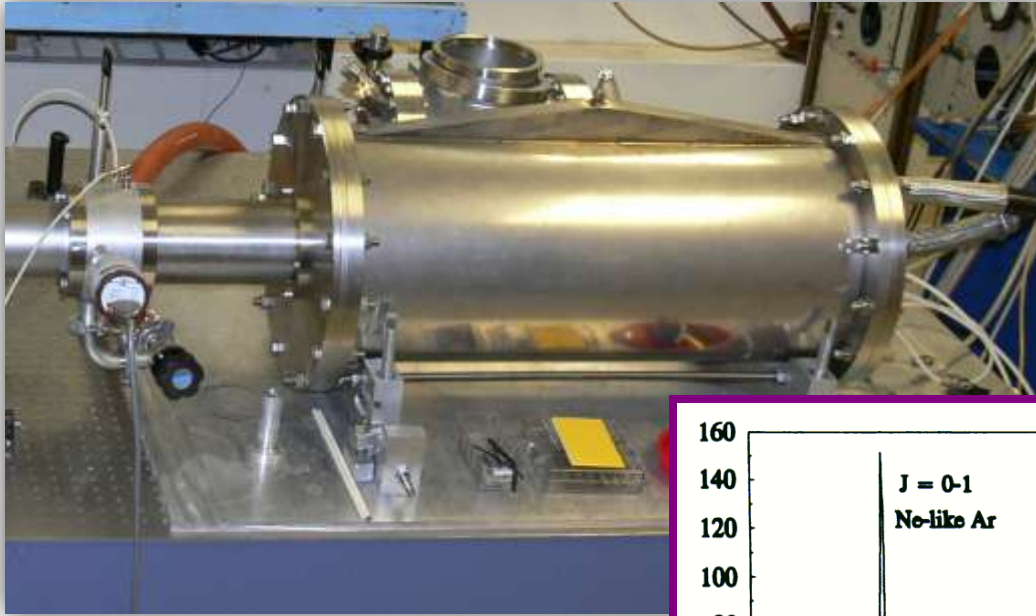
- possibility to perform imaging experiments **in the laboratory environment**
- higher chances for **future commercialization**
- **easy, low cost operation, easy access to the user**

Gabor Holography: Coherent EUV source – Capillary discharge laser $\lambda=46.9\text{nm}$

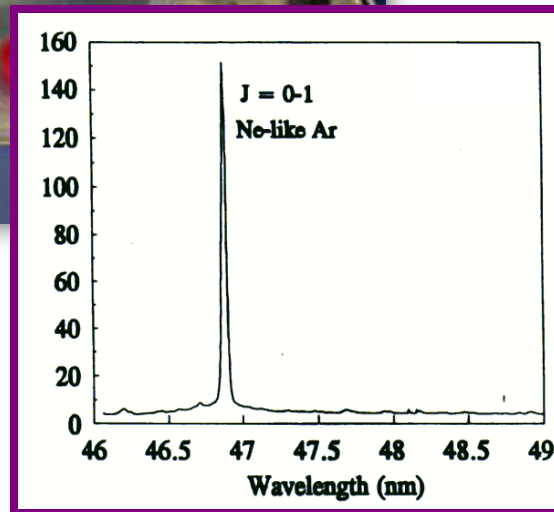
- High fluence
mW average power

- High monochromaticity

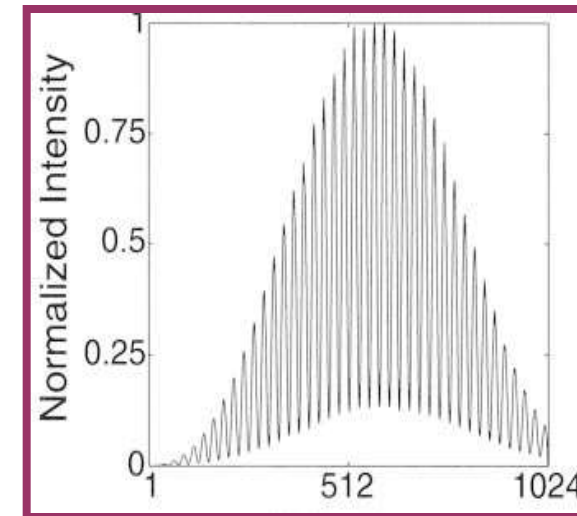
- High spatial coherence



- Repetition rate: 4 Hz
- High energy per pulse max - 0.8 mJ
- Average power ~ 3 mW
 - High monochromaticity: $\Delta\lambda/\lambda < 10^{-4}$
- Coherence radius: $R_c = 550 \mu\text{m}$
at 0.157m from 36cm capillary
- Very compact



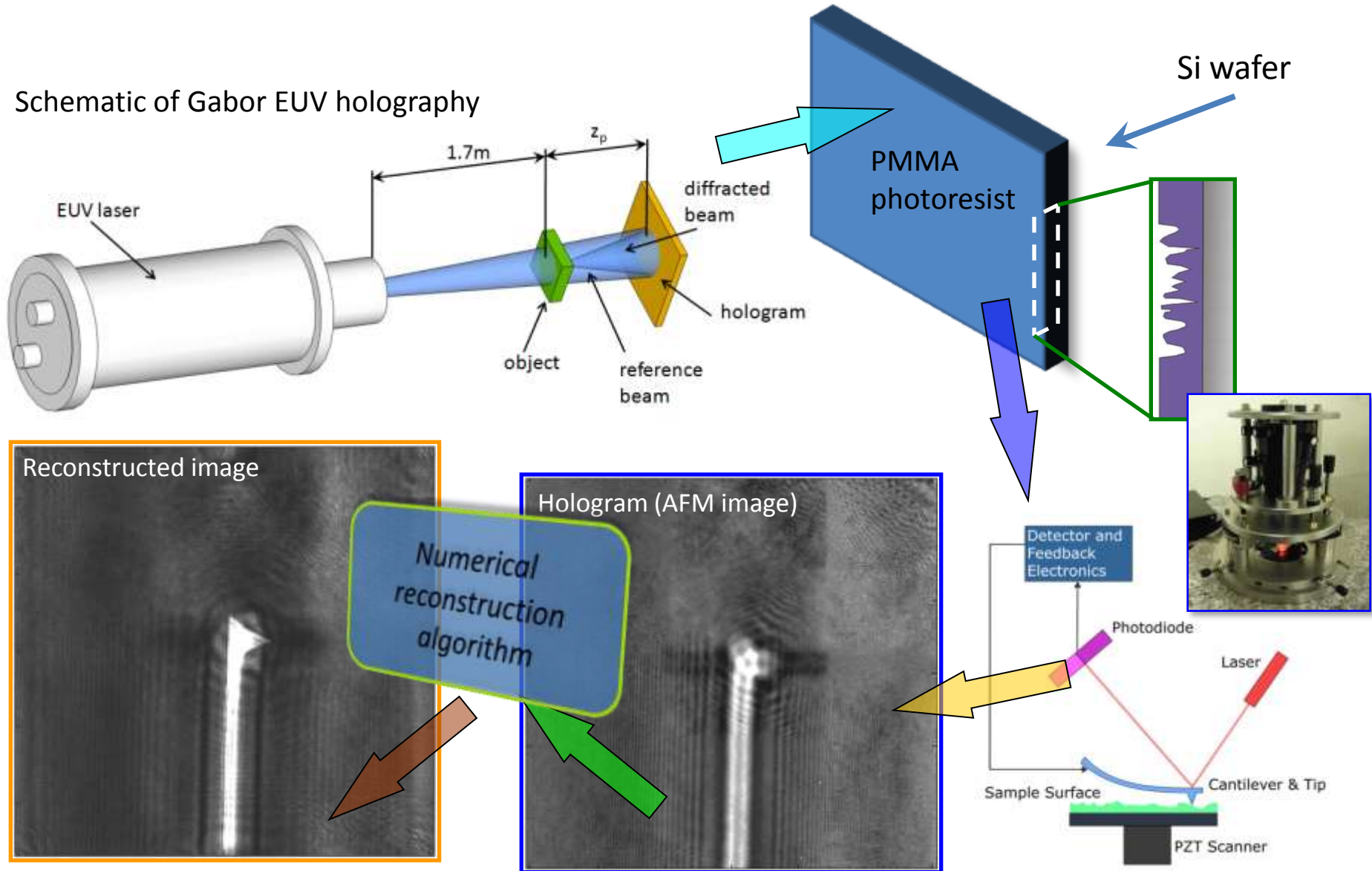
J.J. Rocca, et al. Phys. Rev. Lett. 73, 2192 (1994).



Y. Liu, et al. Phys. Rev. A, 63, 033802 (2001).

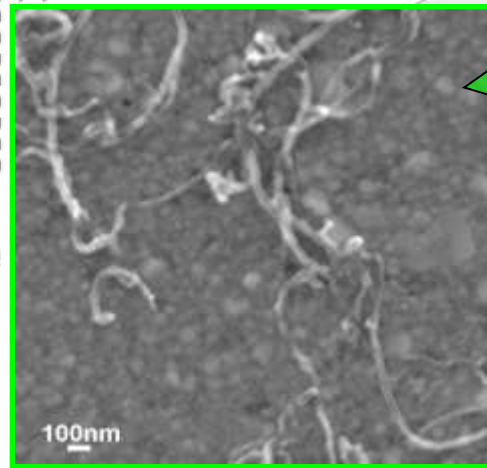
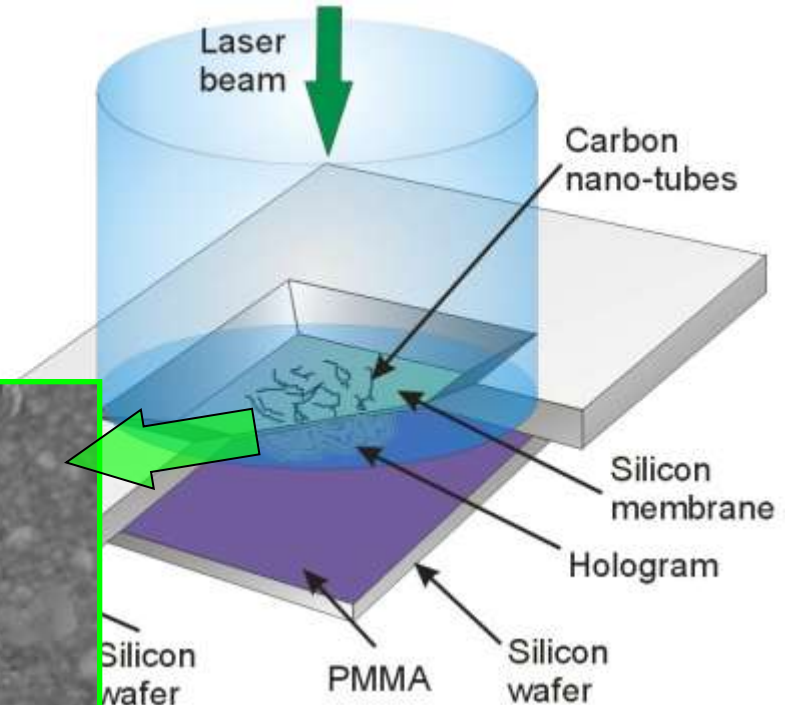
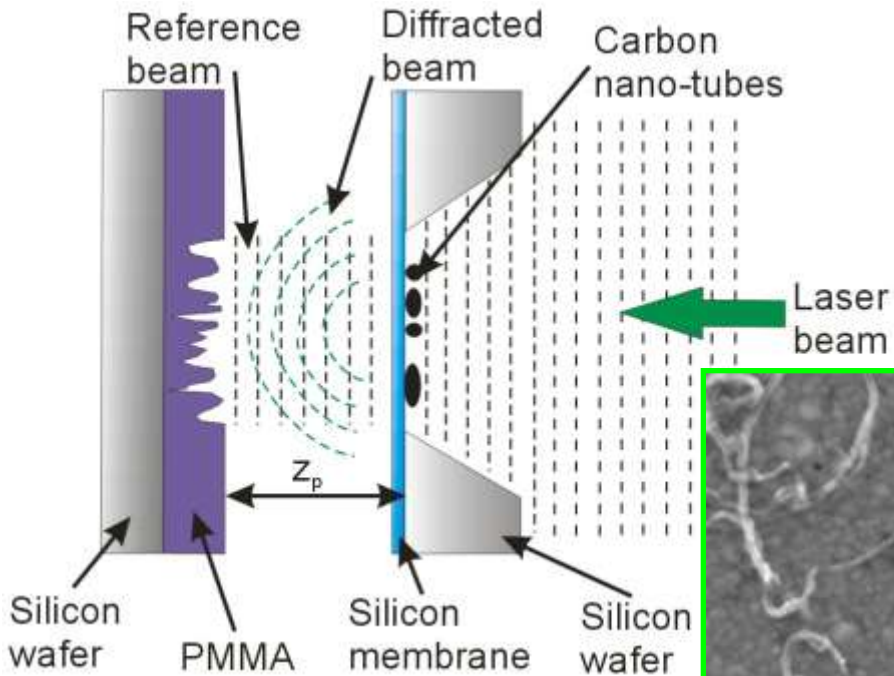
Gabor holography - recording and reconstruction

Schematic of Gabor EUV holography



Wavelength resolution holographic scheme

Experimental arrangement



SEM image of carbon nanotubes

Object characteristics:

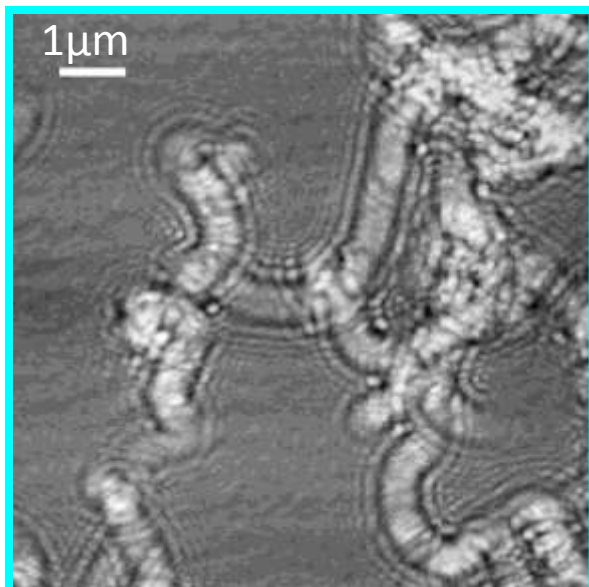
Multi-Wall Carbon Nanotubes

- Outer Diameter 50-80nm
- Length 10-20 μ m

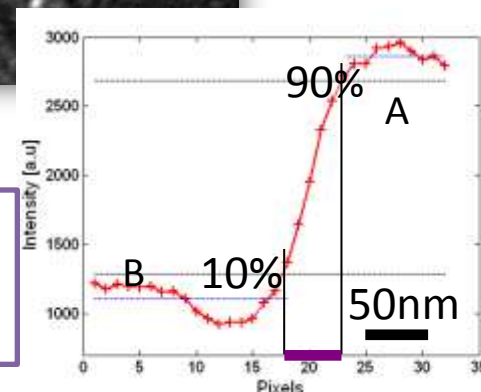
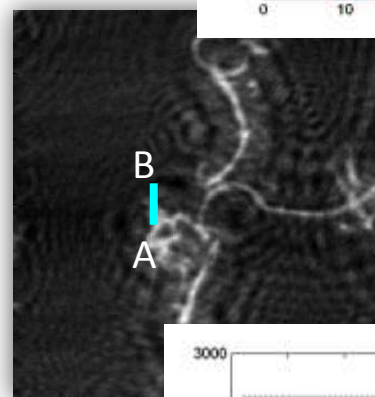
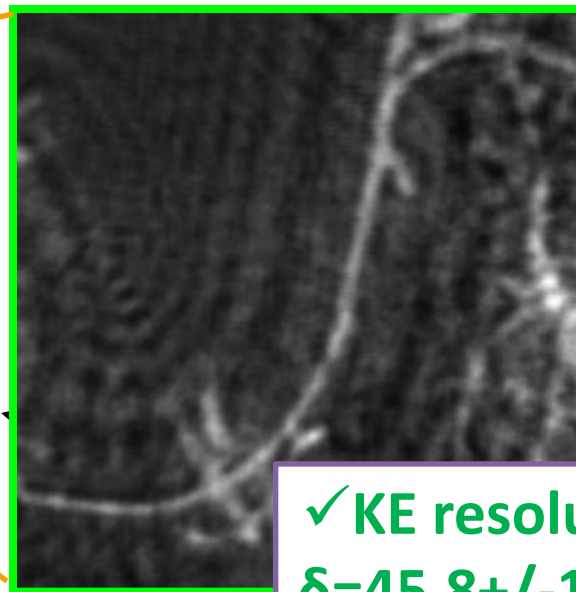
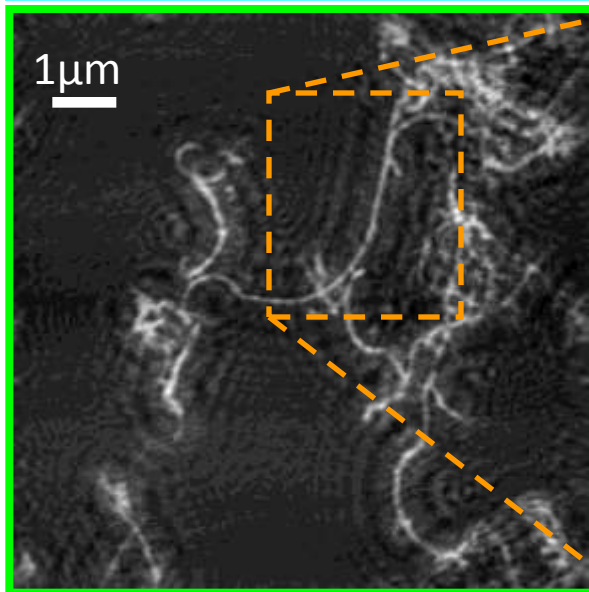
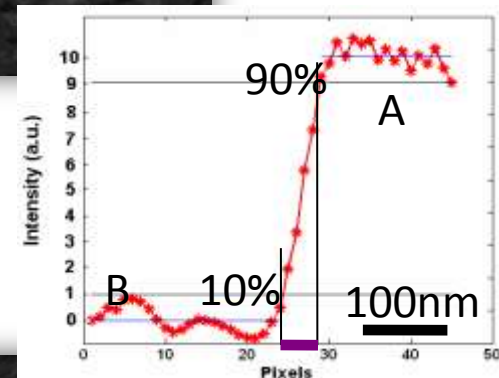
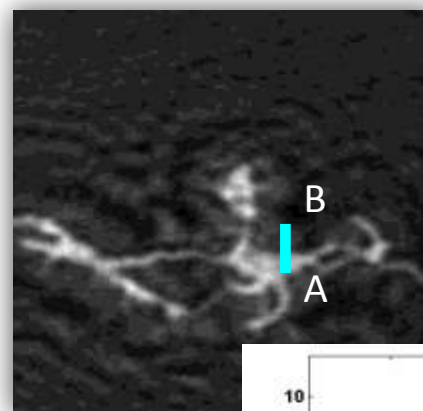
Experimental details:

- Illumination wavelength $\lambda = 46.9\text{nm}$
- Object-photoresist distance $z_p \sim 2.5 - 4\mu\text{m}$
- Laser-object distance $z_s = 75\text{cm}$
- Required number of laser shots = 150, \Rightarrow Dose = 53mJ/cm²
- Membrane transmission $T \sim 15-20\%$

2-D holograms and reconstructions of CNT

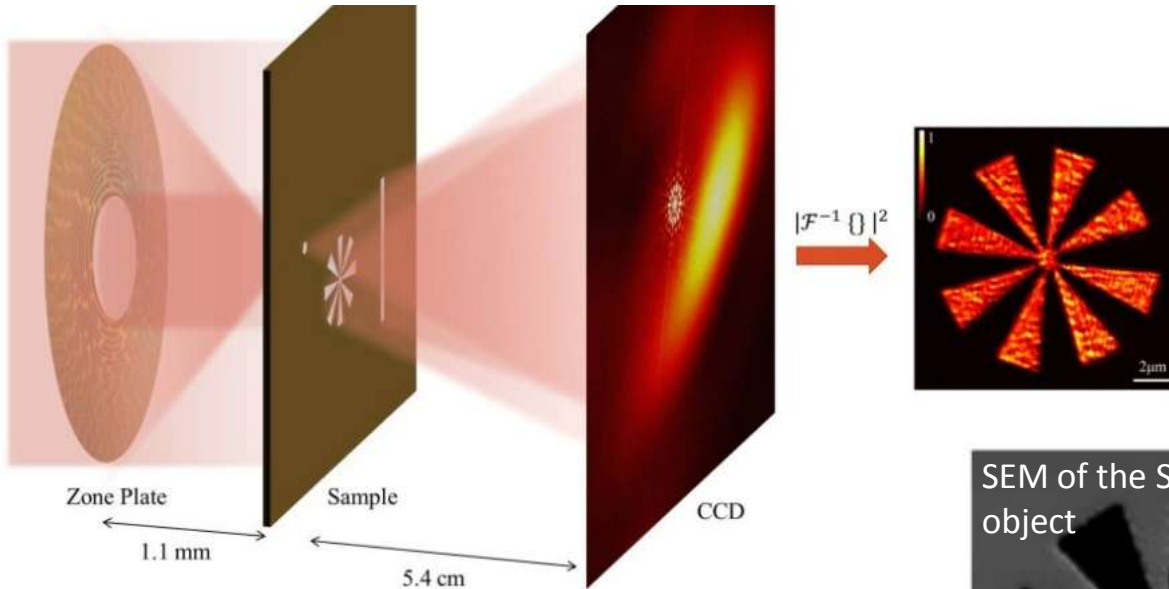


- hologram
- Reconstruction parameters:
- $z_p = 2.66 \mu\text{m}$
 - pixel size = 9.7nm
 - scan size typically $10 \times 10 \mu\text{m}^2$
 - max scan size $42 \times 42 \mu\text{m}^2$



✓ KE resolution:
 $\delta = 45.8 \pm 1.9 \text{ nm}$

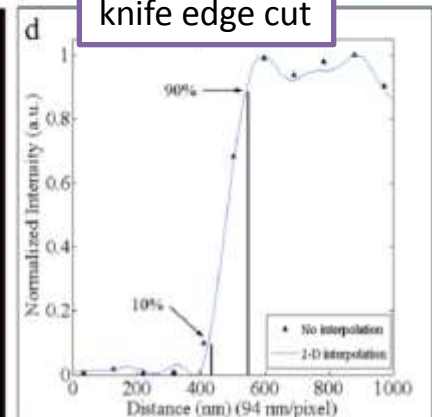
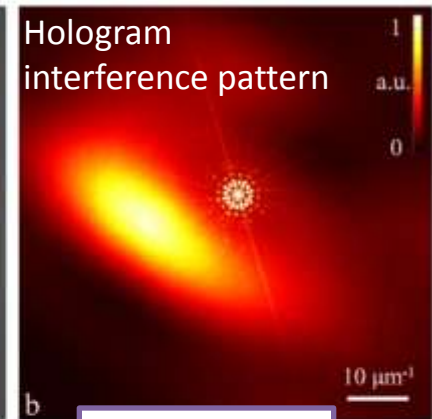
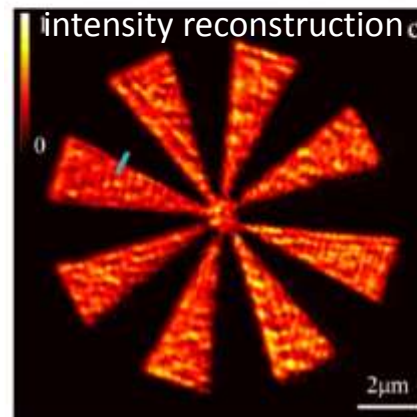
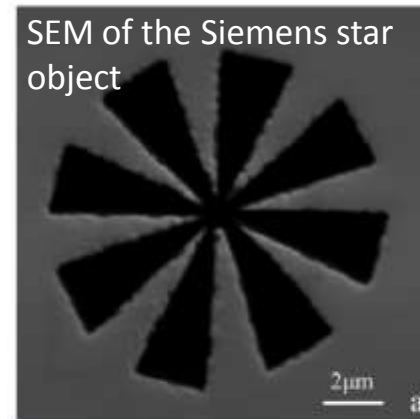
Single-shot EUV Fourier transform holography of an extended object



Schematic of the Fourier transform holography setup

- EUV beam is incident on the zone plate
- The central opening in the ZP passed the 0-order beam to illuminate the object
- 1st order focus is the reference wave.
- The interference between the two beams was collected on a CCD and numerically reconstructed to obtain the final image

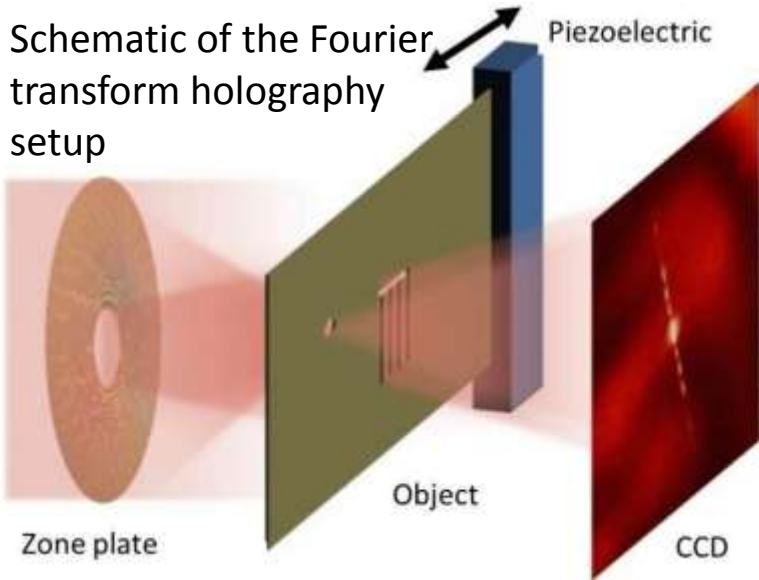
KE spatial resolution **120nm** (multishot exposure, **167nm** for a single shot exposure)



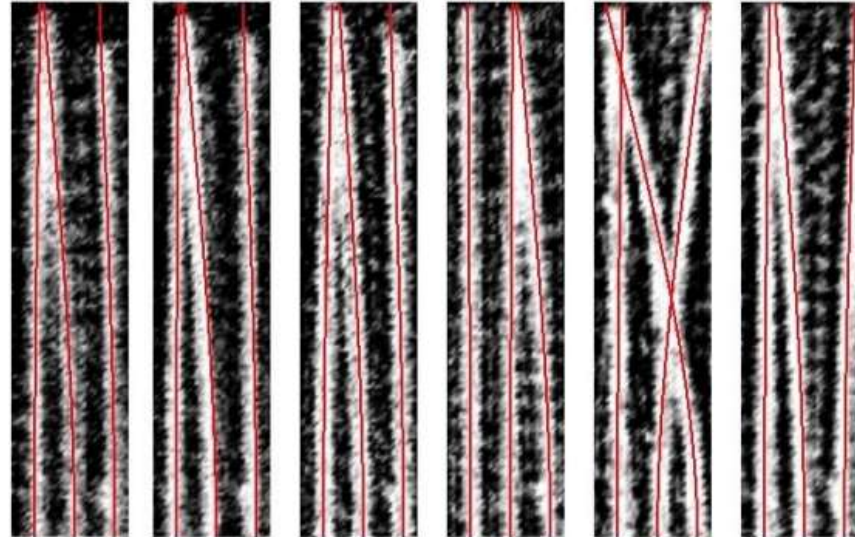
Recording oscillations of sub-micron size cantilevers by EUV Fourier transform holography



Schematic of the Fourier transform holography setup

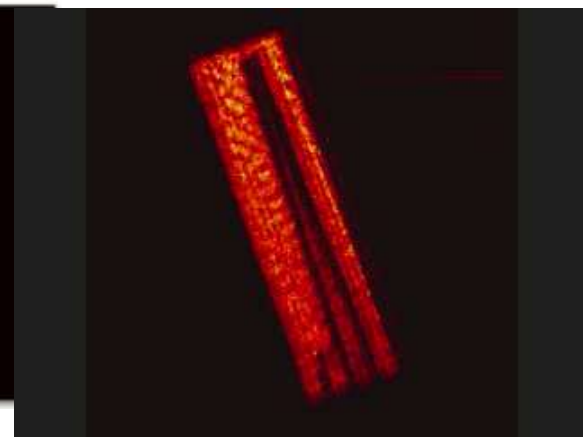
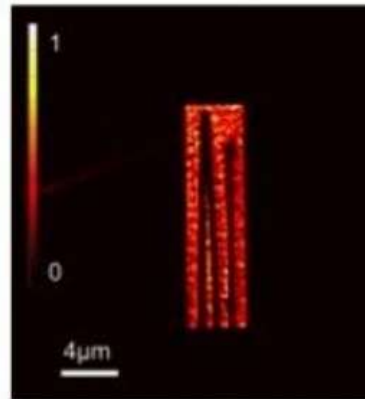
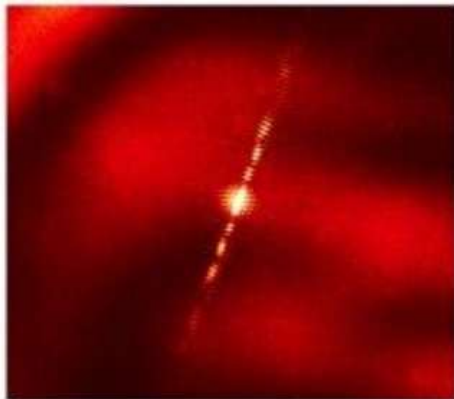
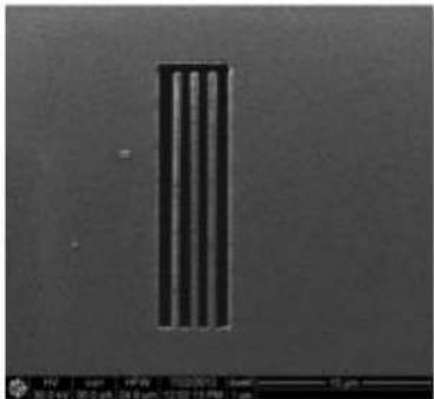


Examples of reconstructed frames of the oscillating pillars



N. Monserud, ... P. Wachulak et al.,
Optics Express 22,4,4161 (2014)

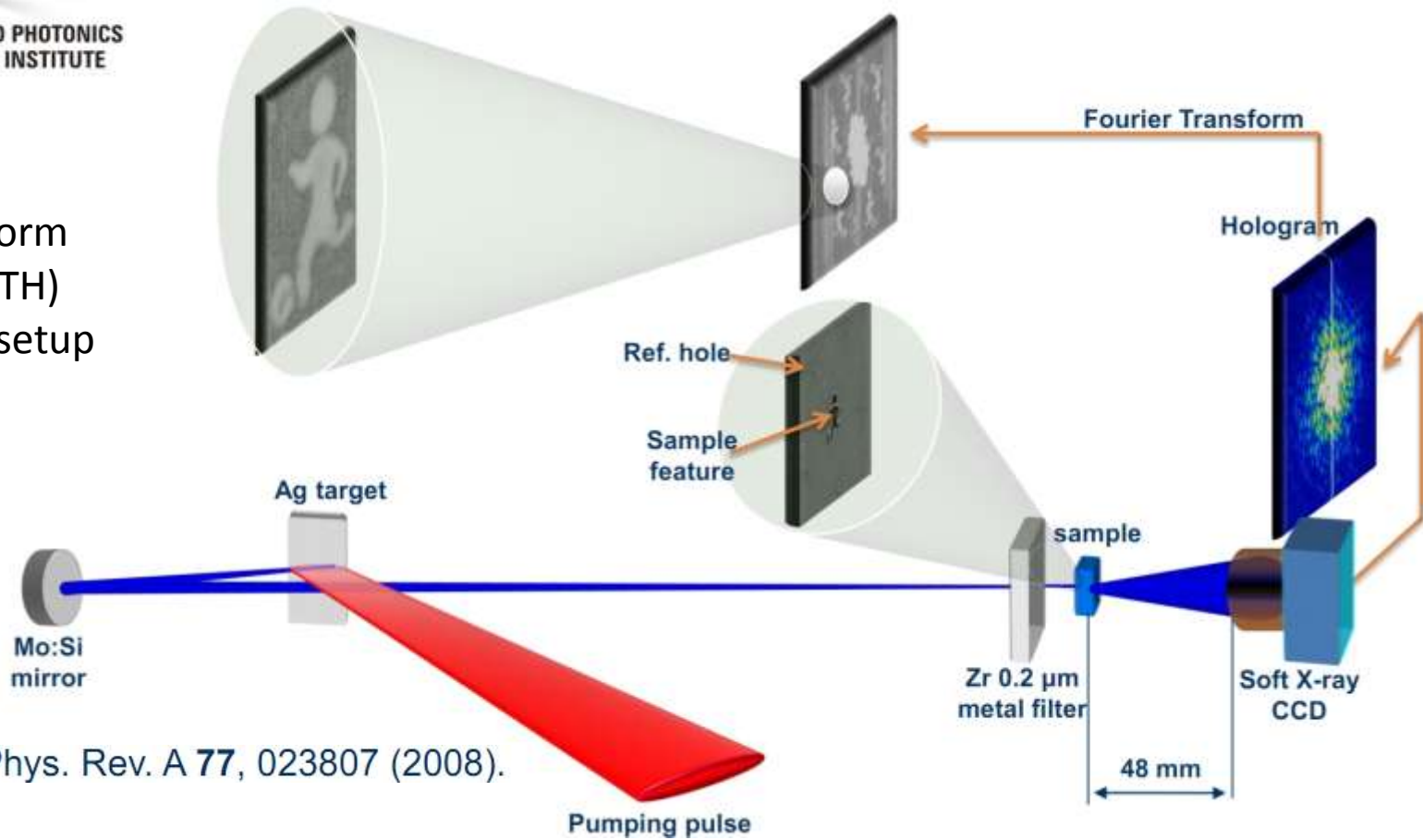
SEM of three nanopillars Central portion of the hologram reconstruction of the pillars



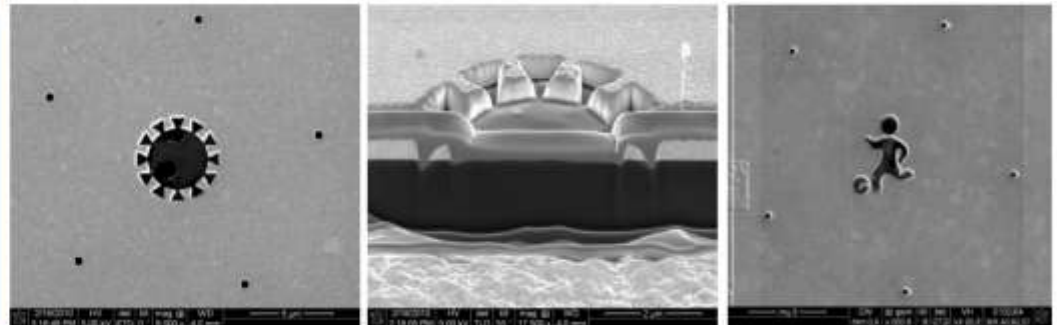
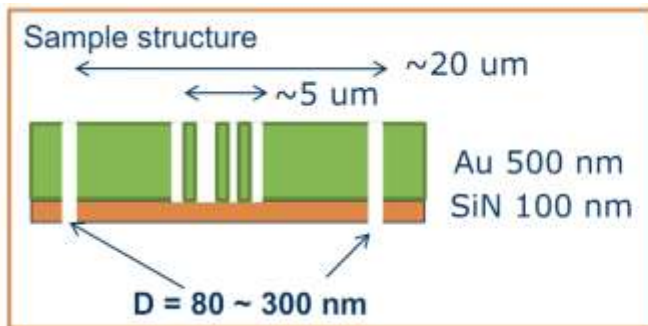
Single shot Fourier Hologram using Ni-Like Ag X-ray laser



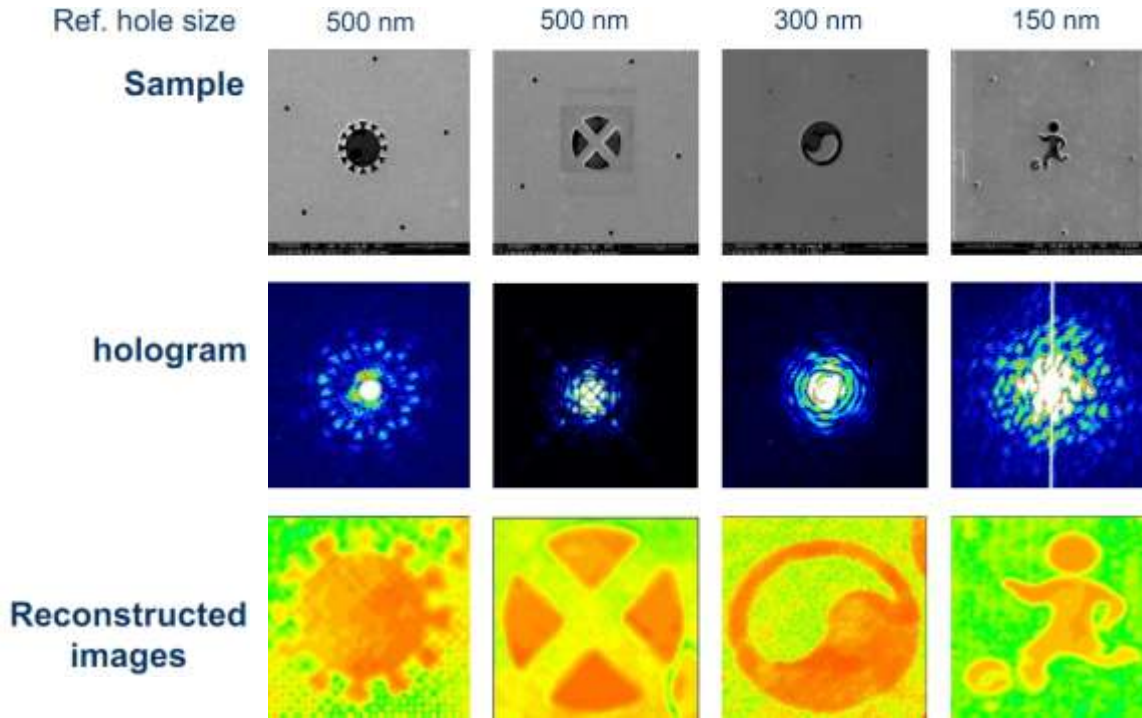
Fourier Transform Holography (FTH) experimental setup



H. T. Kim et al., Phys. Rev. A 77, 023807 (2008).



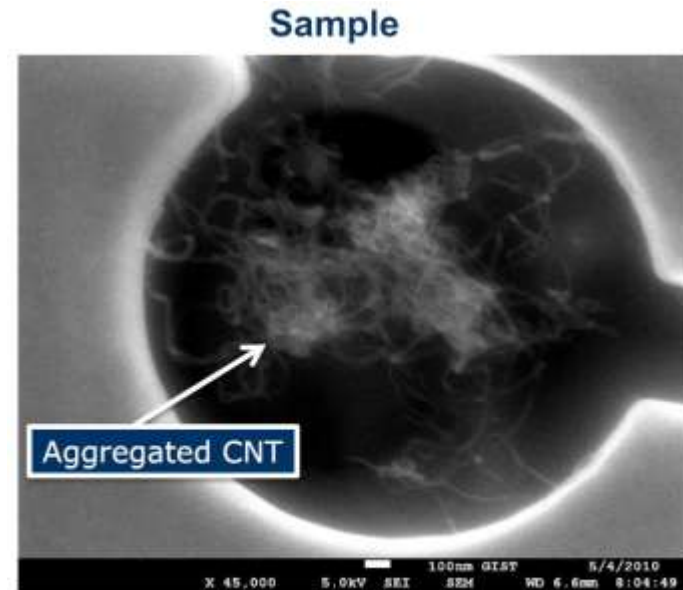
Single shot Fourier Hologram using Ni-Like Ar X-ray laser



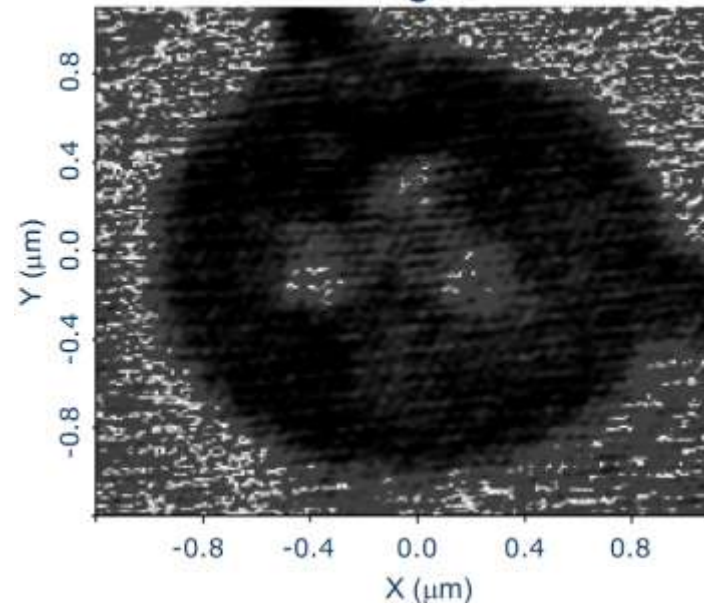
Various holograms and reconstructed images

Resolution of 94nm,
single shot, $\lambda=13.9\text{nm}$

Single shot
hologram and
reconstruction
of carbon
nanotubes



Carbon Nano Tube on the SiN membrane
(width : 20 – 30 nm)
Image



EUV reconstruction of computer generated holograms (CGH)

Experimental details:

Wavelength $\lambda=46.9\text{nm}$

Pixel Size= 140nm

Line Width= $1.54\mu\text{m}$

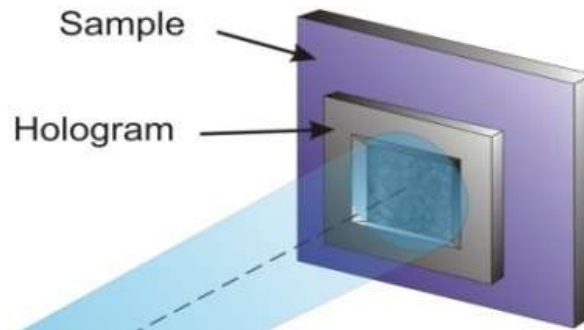
Reconstruction EUV laser

distance $Z=500\mu\text{m}$

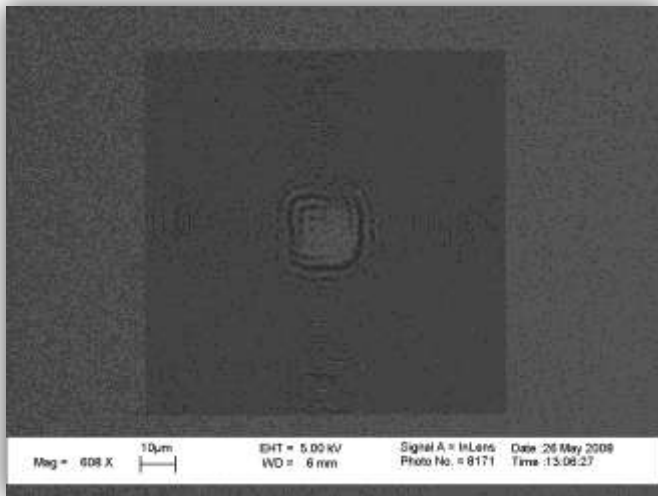
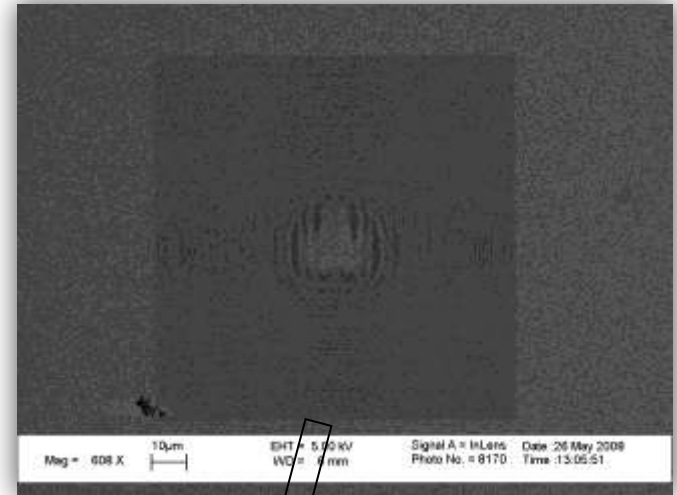
Field= $102.9 \times 102.9\mu\text{m}^2$

NA= 0.102 , res. $\sim 230\text{nm}$

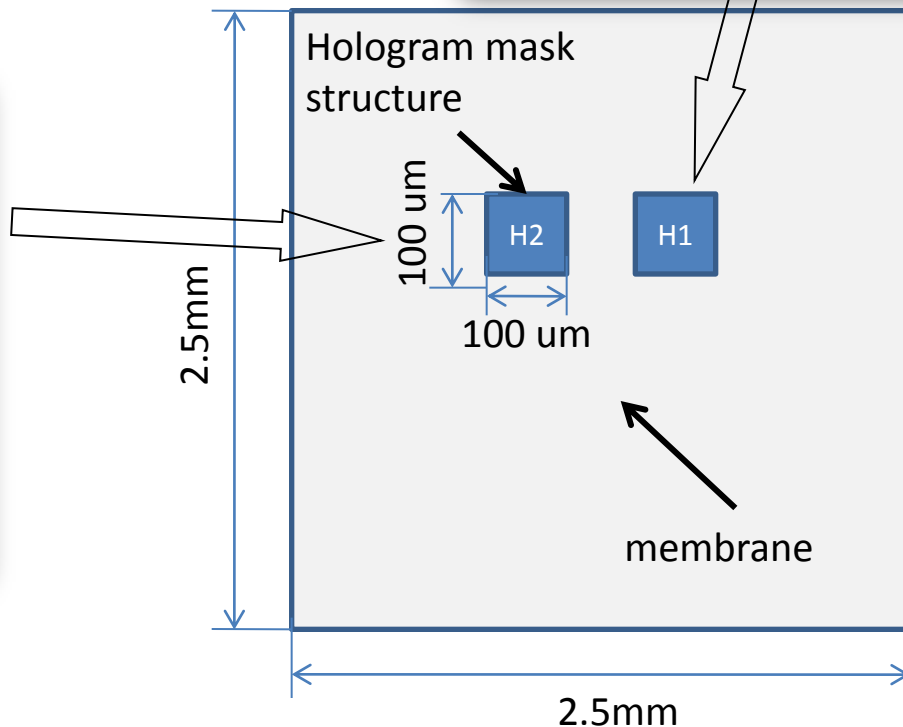
DOF= $4.5\mu\text{m}$



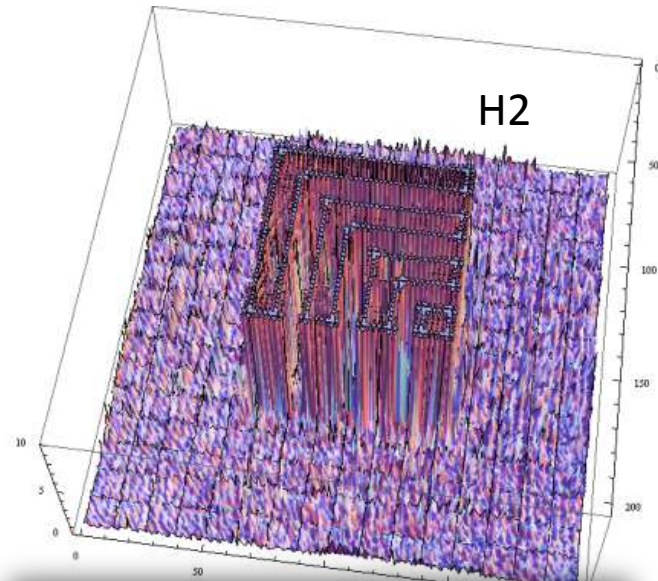
SEM images of Hologram mask: H1



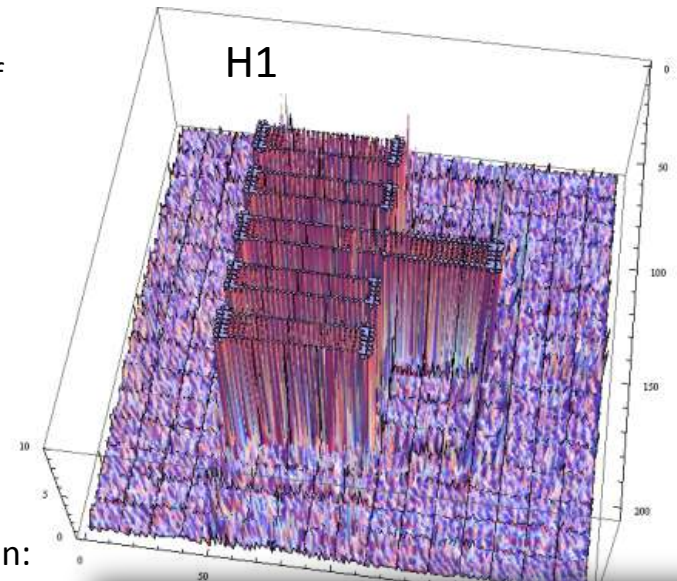
SEM images of Hologram mask: H2



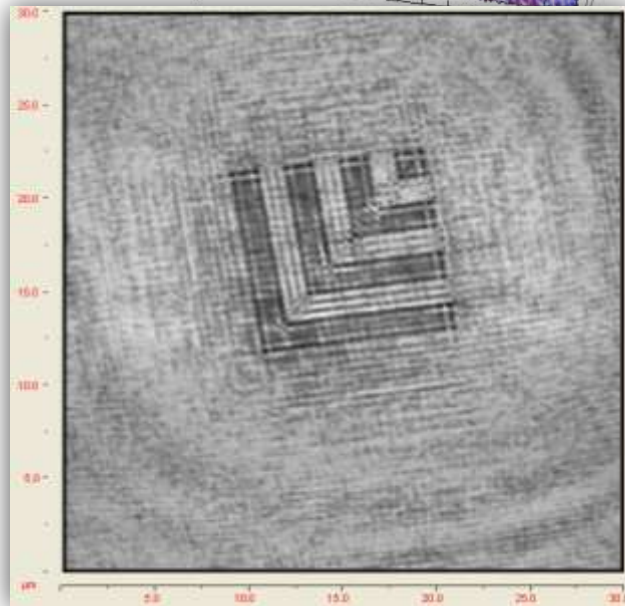
CGH EUV reconstruction results



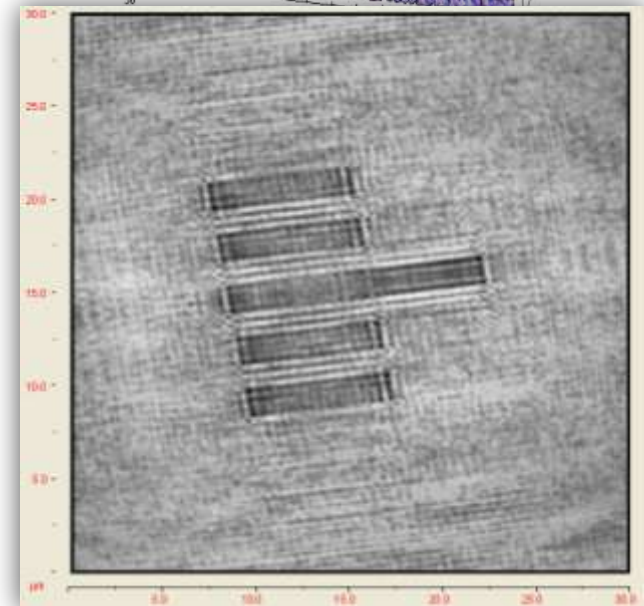
Simulated images of
reconstructed
holograms



Hologram reconstruction:
AFM images
@ 500um gap (PMMA)

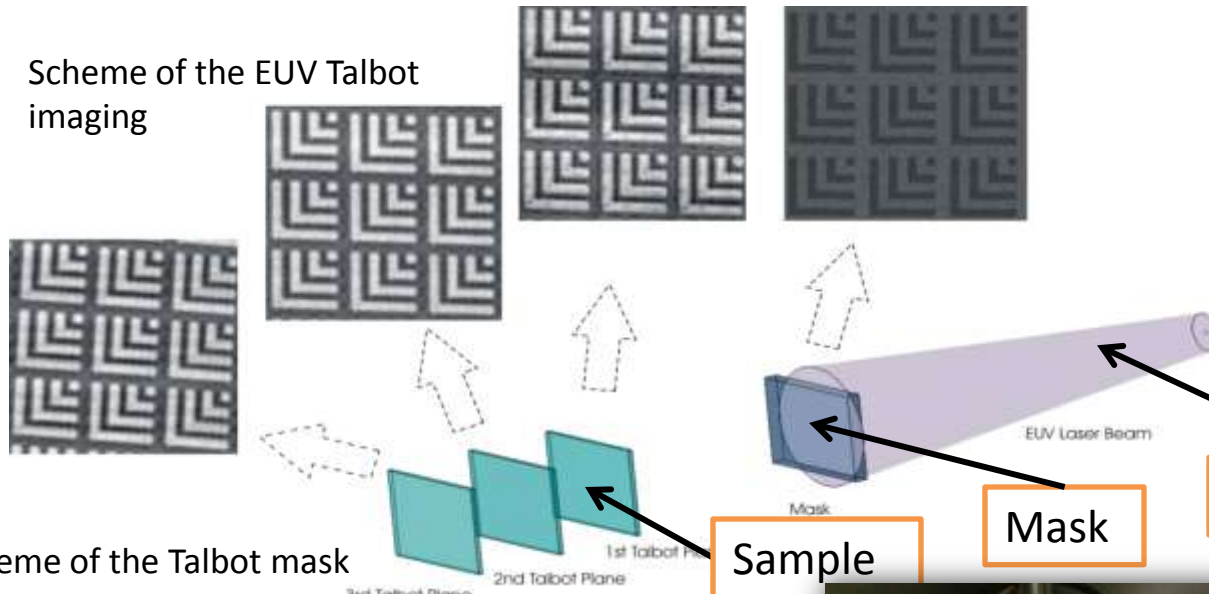


Advantage:
- your object does
not need to exist
- Hologram might be
partially damaged,
your object will be
reconstructed anyway.



Talbot imaging at EUV wavelengths

Scheme of the EUV Talbot imaging



Experimental details:

Wavelength $\lambda=46.9\text{nm}$

Smallest features= 140nm

Cell size $p=4.842\ \mu\text{m}$ (124×124)

Rec. distance $z_T=1000\ \mu\text{m}$

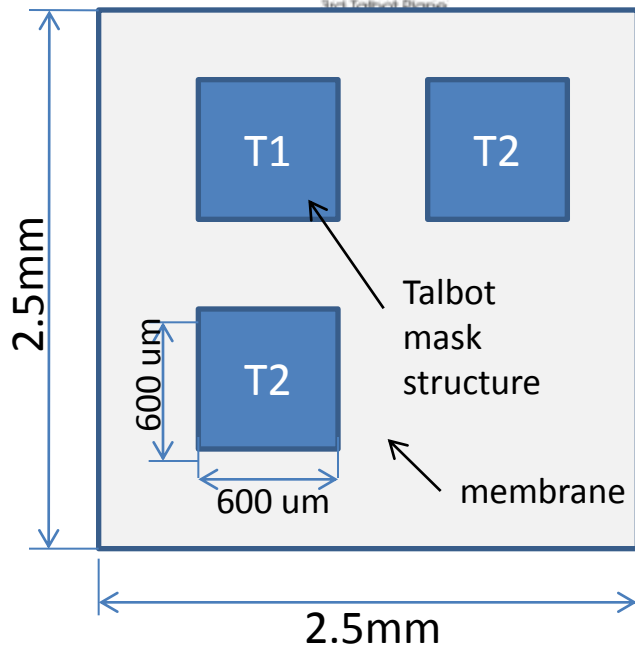
Field= $600\times 600\ \mu\text{m}^2$

NA= 0.282 , res.= 83nm

DOF= $0.6\ \mu\text{m}$

$$z_T(n) = \frac{2p^2}{\lambda} n$$

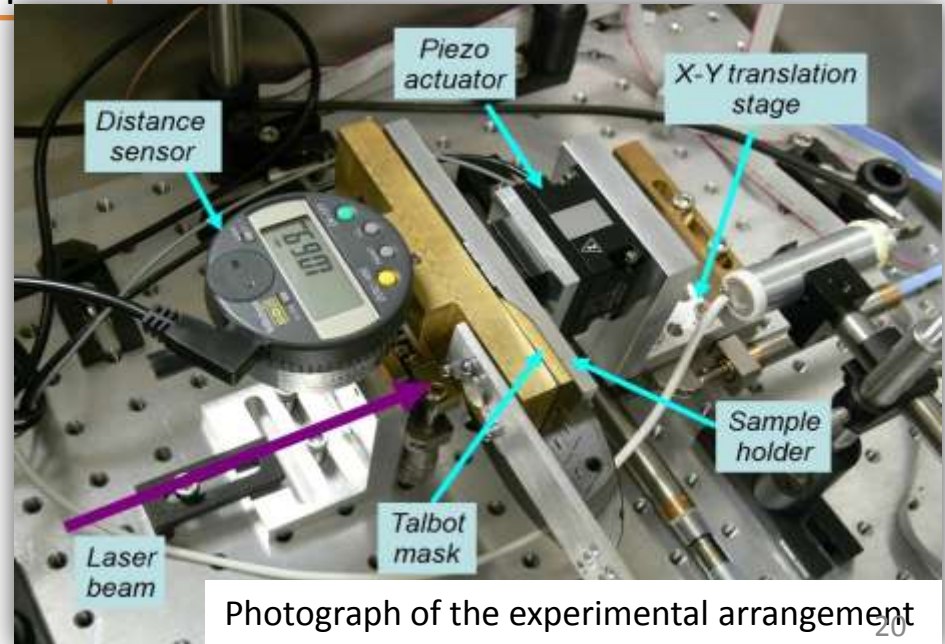
Scheme of the Talbot mask



Sample

Mask

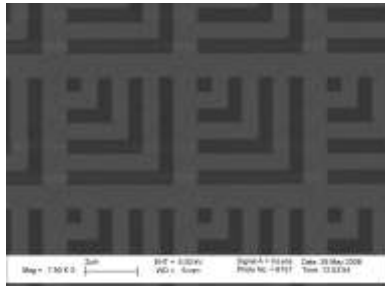
EUV beam



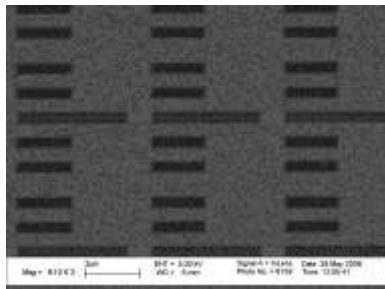
Photograph of the experimental arrangement

Talbot imaging results

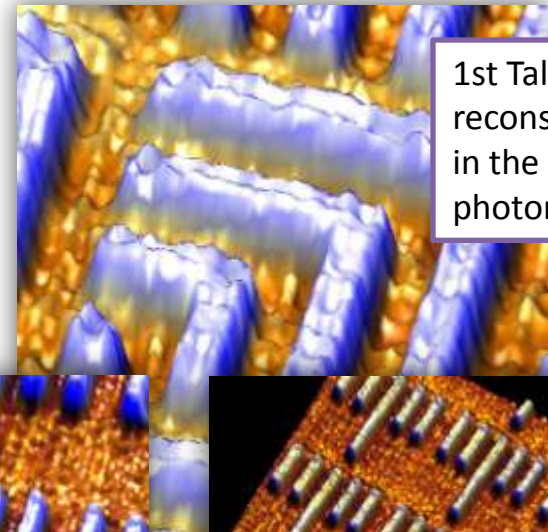
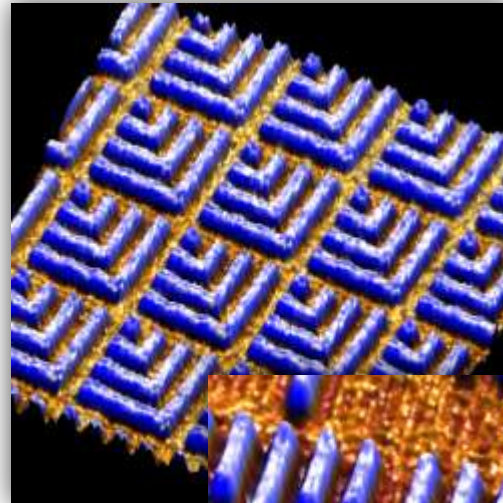
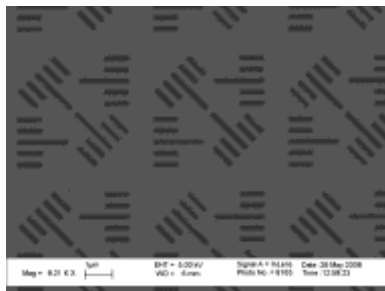
SEM images of Talbot self-imaging mask: T1



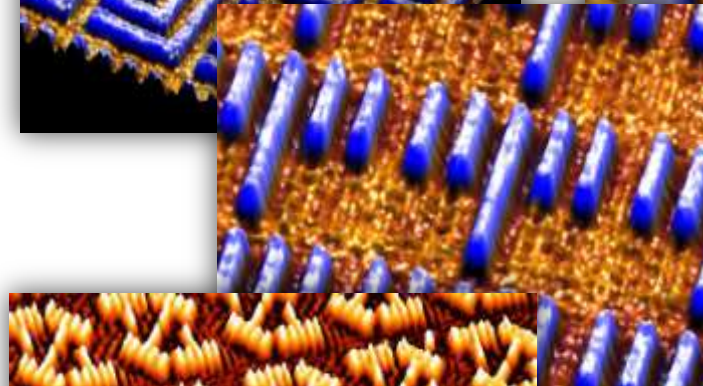
mask: T2



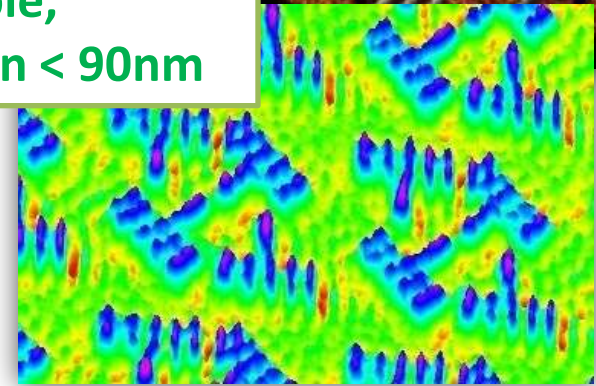
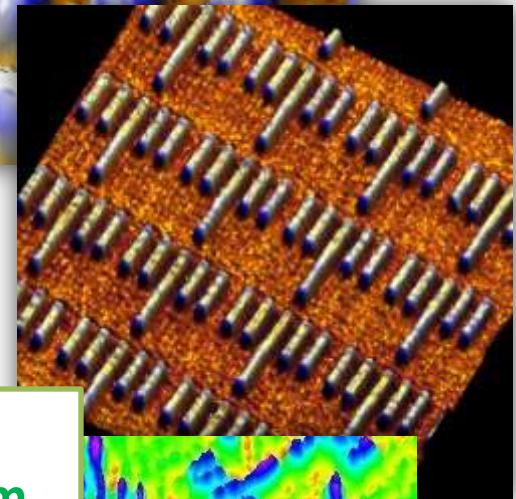
mask: T3



1st Talbot planes reconstructions in the PMMA photoresist

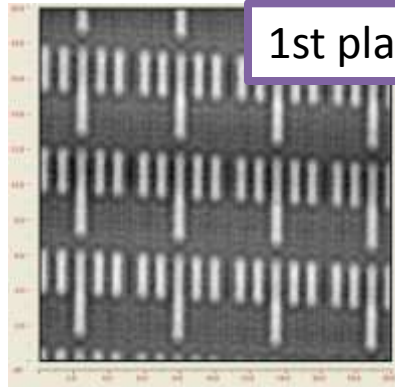


✓ 140nm features visible, theoretical resolution < 90nm

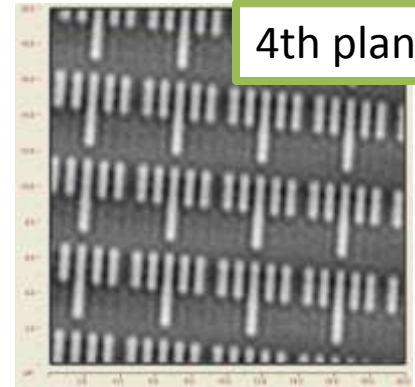
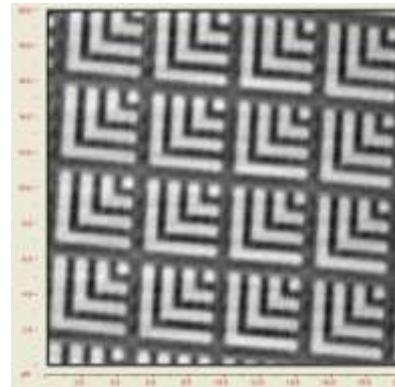


Different Talbot planes

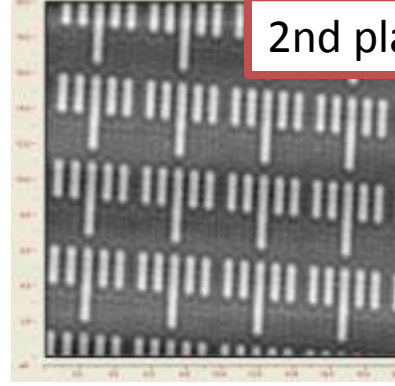
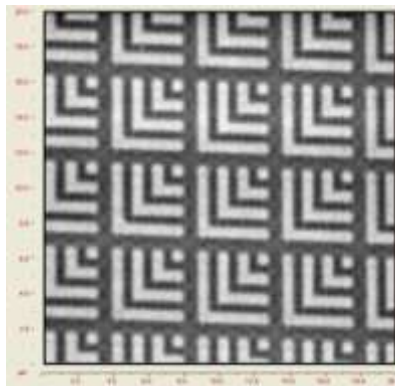
Talbot image plane AFM images $20 \times 20 \mu\text{m}^2$, $z_T = 1 \text{mm}$



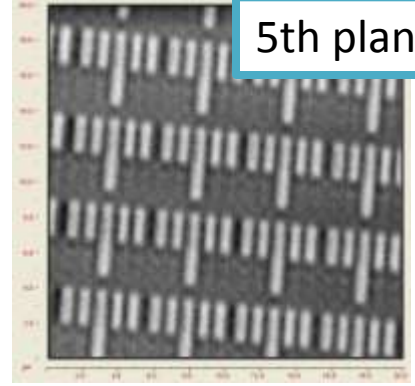
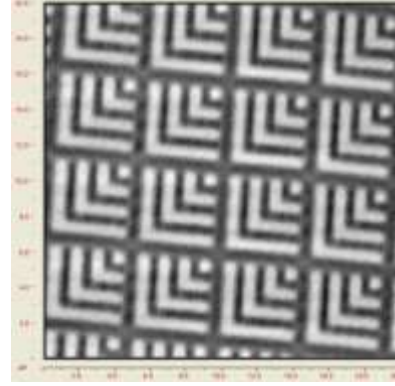
1st plane



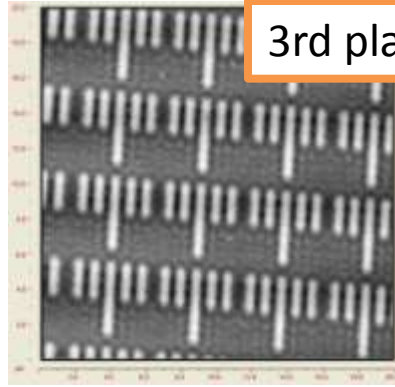
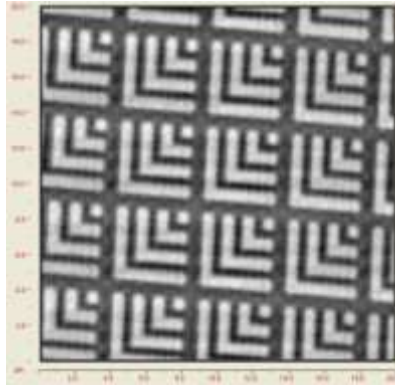
4th plane



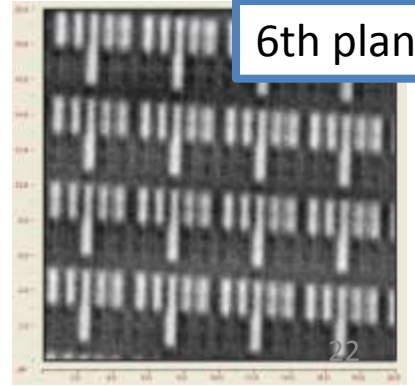
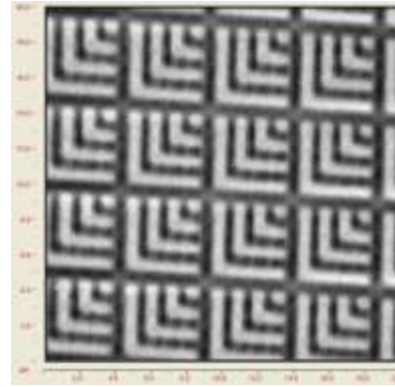
2nd plane



5th plane



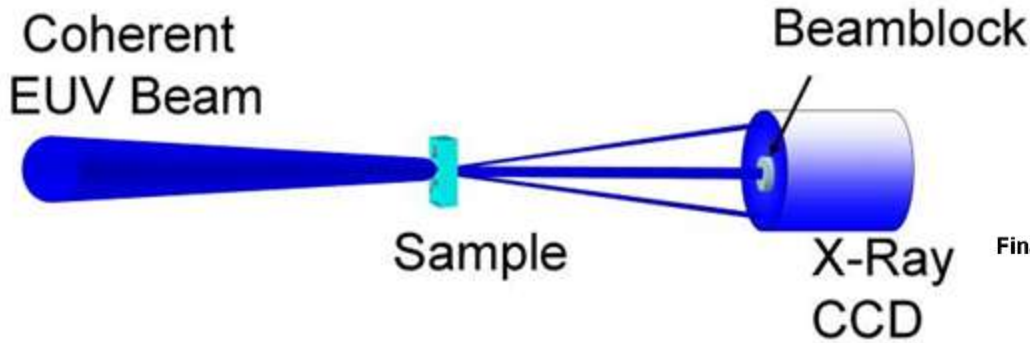
3rd plane



6th plane

Coherent Diffractive (Lens-less) Imaging

No lens— Fourier relationship of diffraction



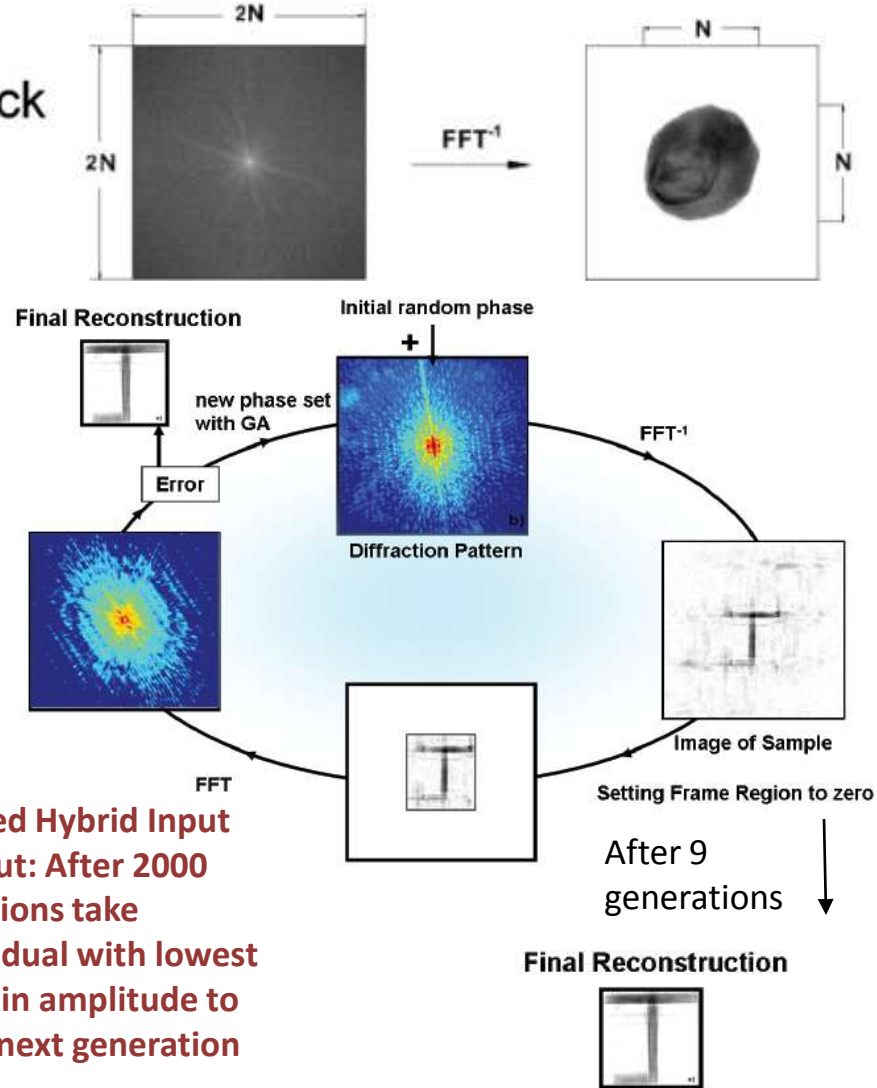
$$E(x, y, z = d) \propto \int E(x, y, z = 0) e^{-\frac{ik}{d}(xx' + yy')} dx' dy'$$

Far Field

$$z \gg \frac{D^2}{\lambda}$$

Far field diffraction is two dimensional Fourier transform of exit wave

Reconstruction



Guided Hybrid Input Output: After 2000 iterations take individual with lowest error in amplitude to seed next generation

Peatross and Ware, *Physics of Light and Optics* (2008)
 D. Sayre, *Acta Cryst* 5, 843 (1952)
 J. Miao et al., *Nature* 400, 342 (1999)

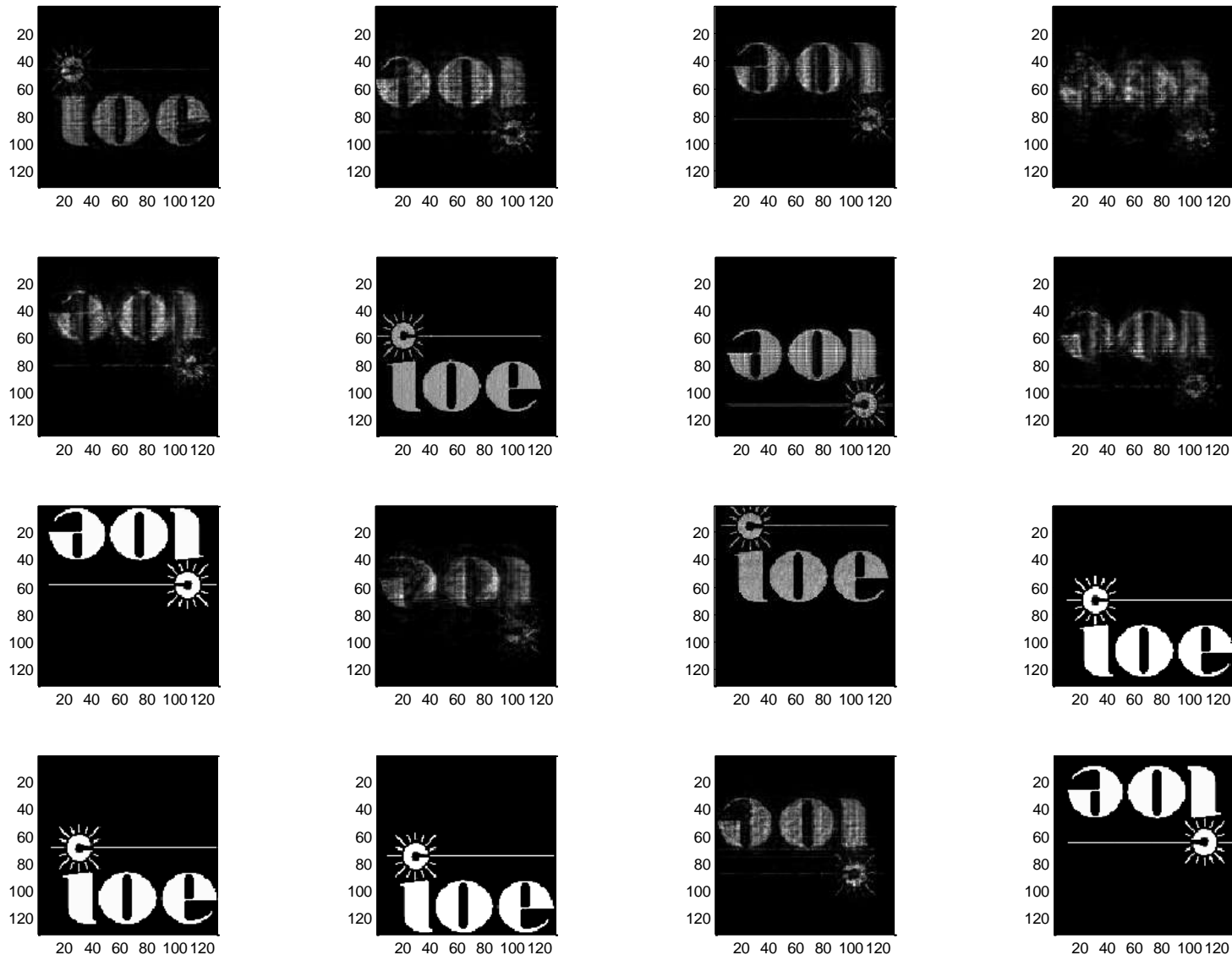
Miao and Sayre, *Act Cryst A56*, 596 (2000)
 C. Song et al., *Phys. Rev. B* 75, 012102 (2007)

Coherent Diffractive (Lens-less) Imaging code

O=4
512x512 pix

500 iteration
per seed

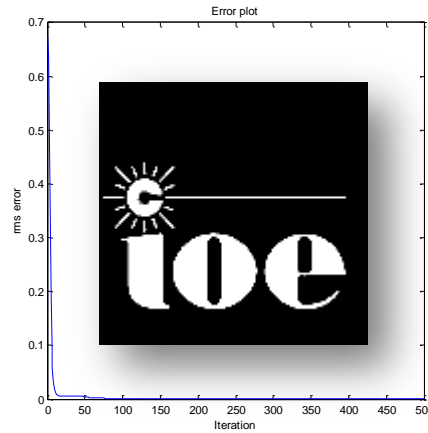
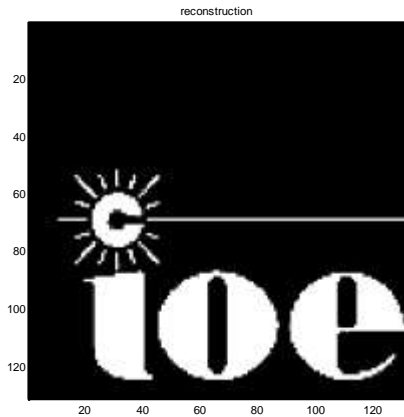
$$|\mathfrak{F}\{f(-x)\}| = |\mathfrak{F}\{f(x)\}|$$



Initial, random phase seed is very important: 16 reconstructions with random phase start

Coherent Diffractive (Lens-less) Imaging code

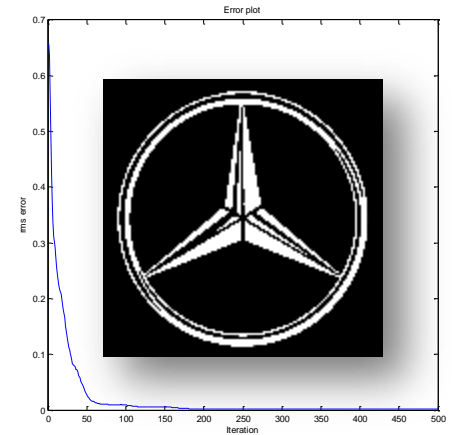
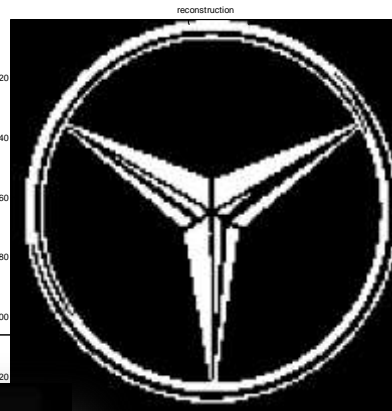
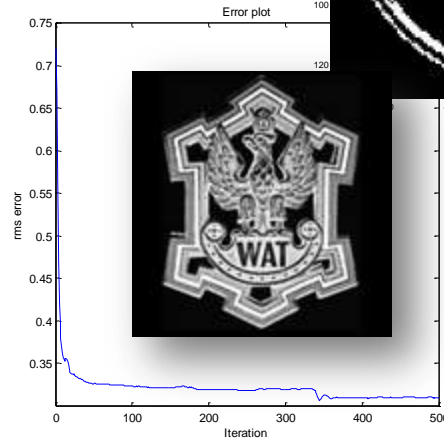
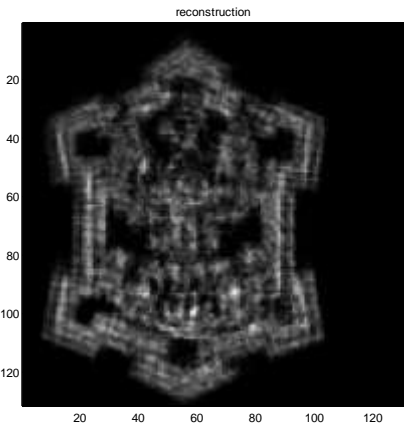
Best reconstruction according to the rms error plot



Fastest code convergence

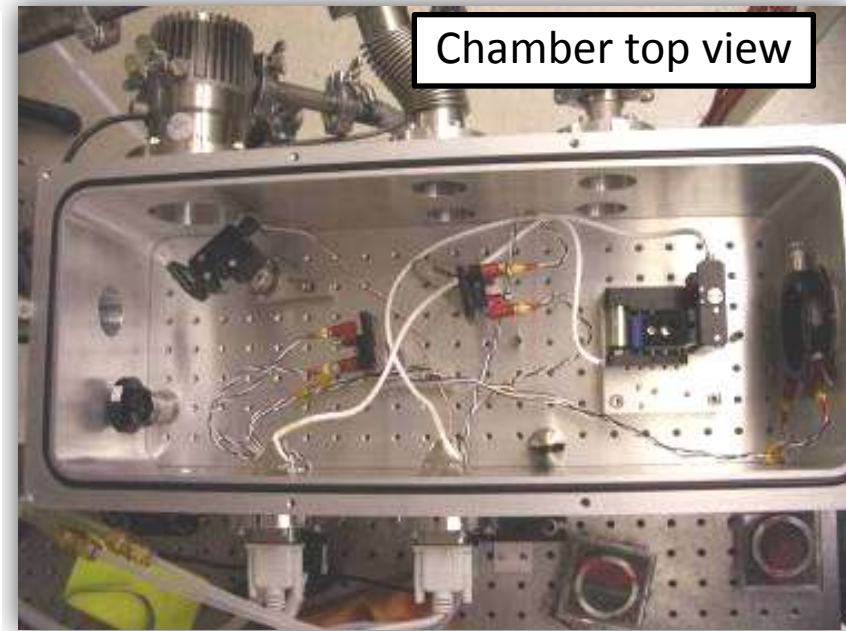
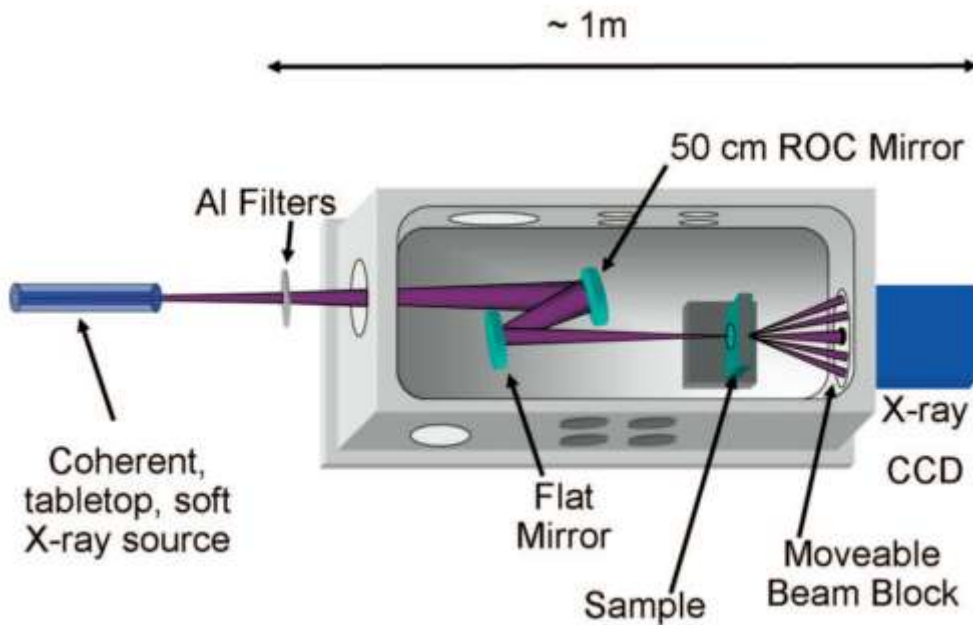
$$Err = \frac{\int_{-\infty}^{\infty} [|G_k(u)| - |F(u)|]^2 du}{\int_{-\infty}^{\infty} |F(u)|^2 du}$$

Smaller Err -> images are more similar



Gray-scaled images need more iterations, harder for the algorithm to converge

Lens-less, Diffraction Imaging - Experimental Setup



Experimental details:

Wavelength $\lambda=46.9\text{nm}$, energy per pulse = 0.2mJ , divergence half angle $\sim 4.5\text{mrad}$, $\lambda/\Delta\lambda=10^4$

Spatially filtered by 1.5mm diameter pinhole at 1.5m from the laser to improve the spatial coherence

100nm Al filter to block a visible light, two Sc/Si multilayer mirrors $R\sim 40\%$ at $\lambda = 46.9\text{nm}$

CCD camera: Andor 2048×2048 pix, $p=13.5\mu\text{m}$ pixel size

Object: $D= 7\mu\text{m}$ tall „stick-figure” etched in 100nm thick SiN membrane

Sample-CCD distance $z= 17\text{mm}$, $O=8.4$

$\text{NA}=0.63$, $\text{res.}=\sim 40\text{nm}$

$$O = \frac{z\lambda}{pD}$$

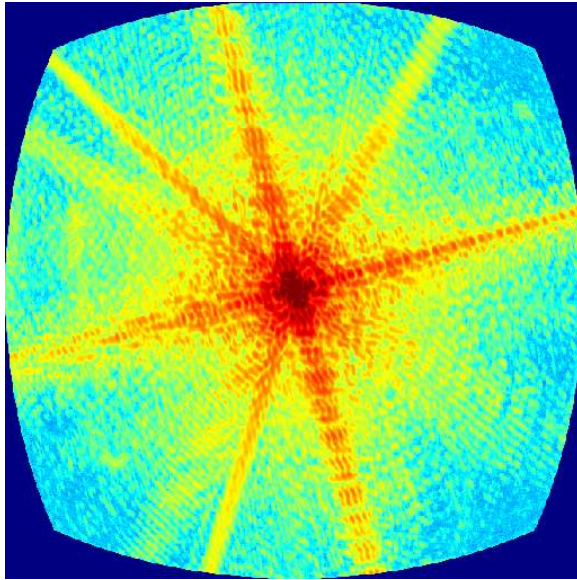


EUV tabletop lens-less imaging at 72 nm resolution ($\lambda=47$ nm)

K-E resolution measurements



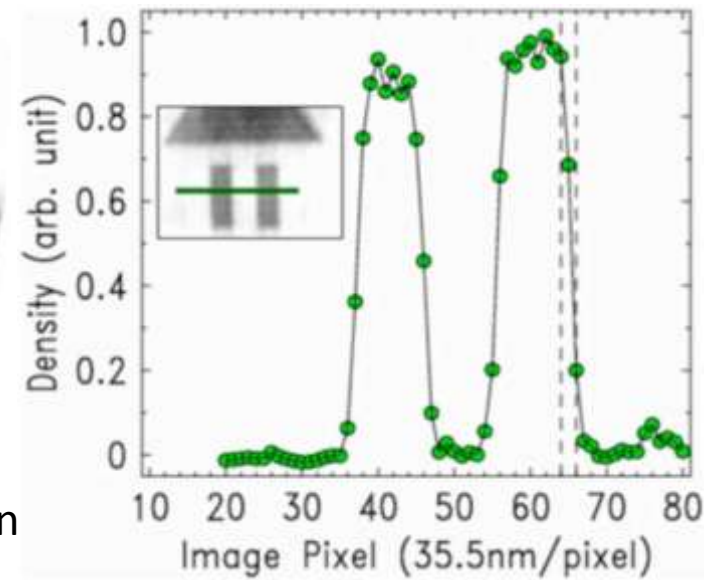
SEM Image



Curvature
Diffraction

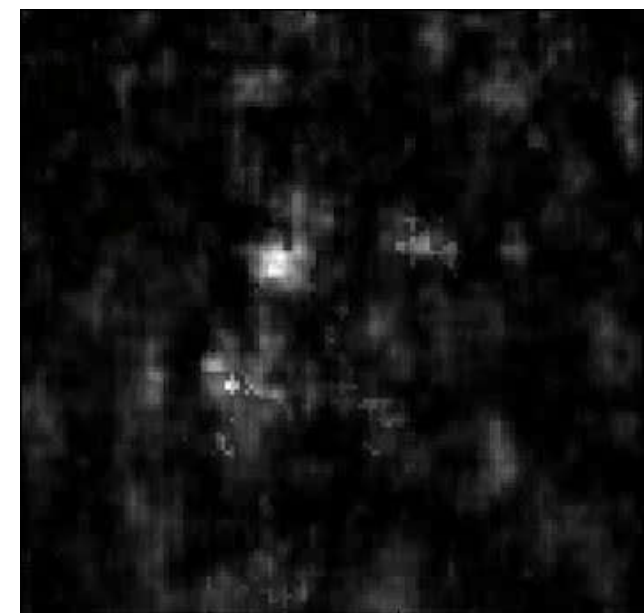
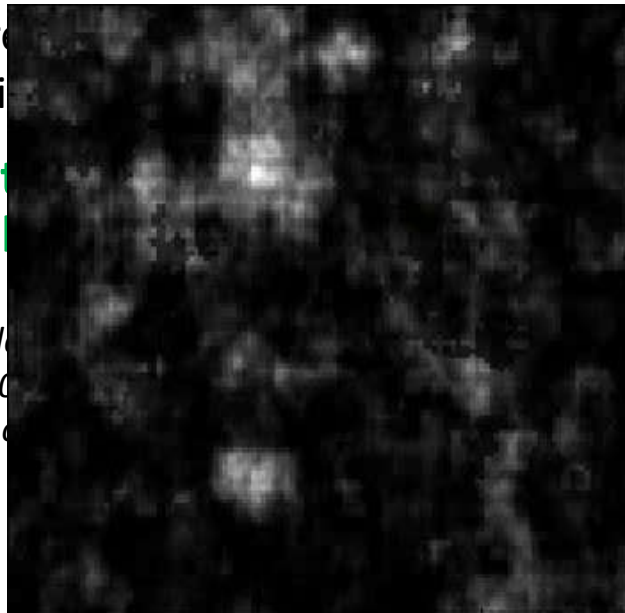


Reconstruction



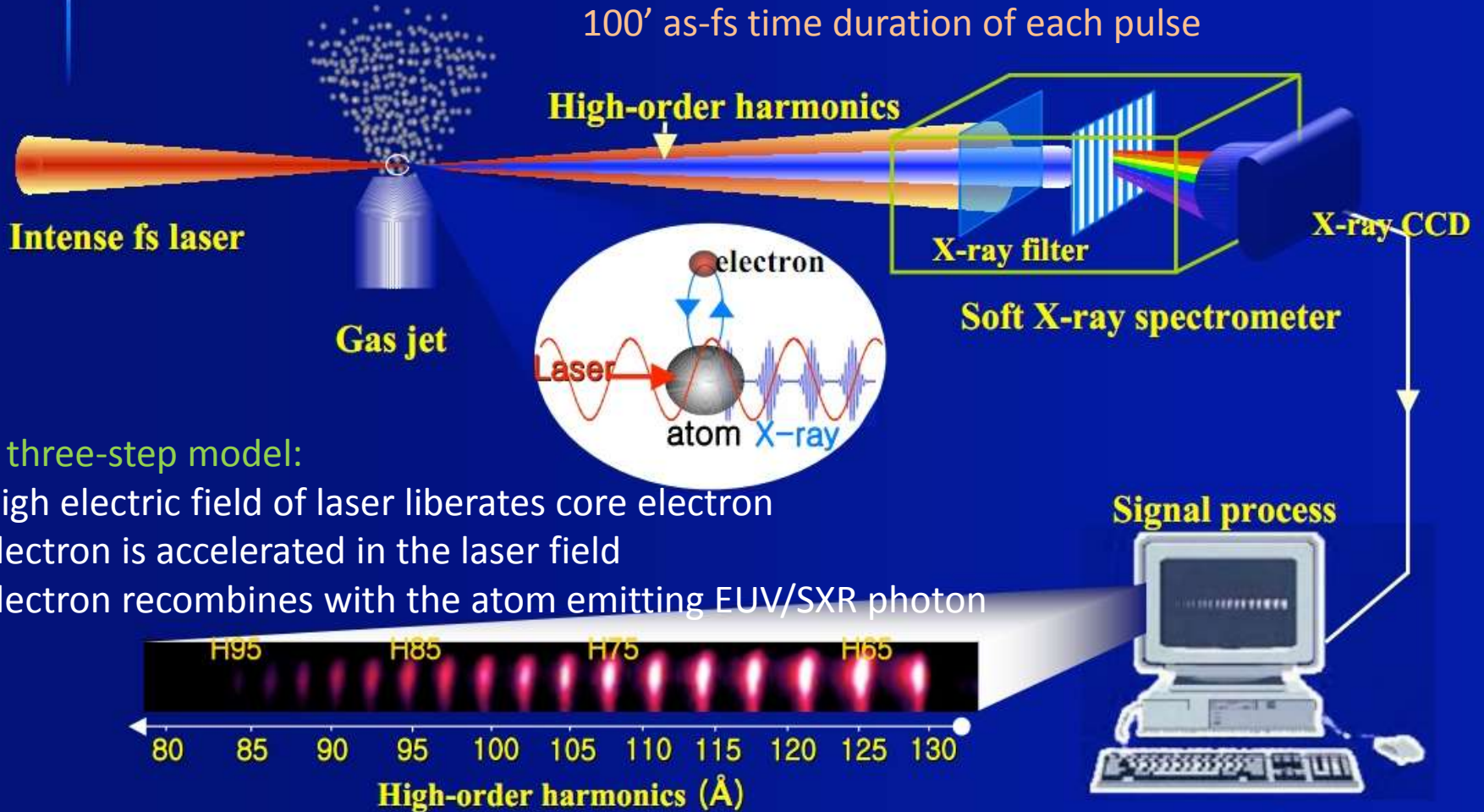
✓ High numerical aperture achieves 72 nm resolution

R.L. Sandberg, C. Song, P.W. Wachulka, *Proceedings of the National Academy of Science*, 108, 10800-10805 (2011) and Research Highlights, *Nature Photonics*

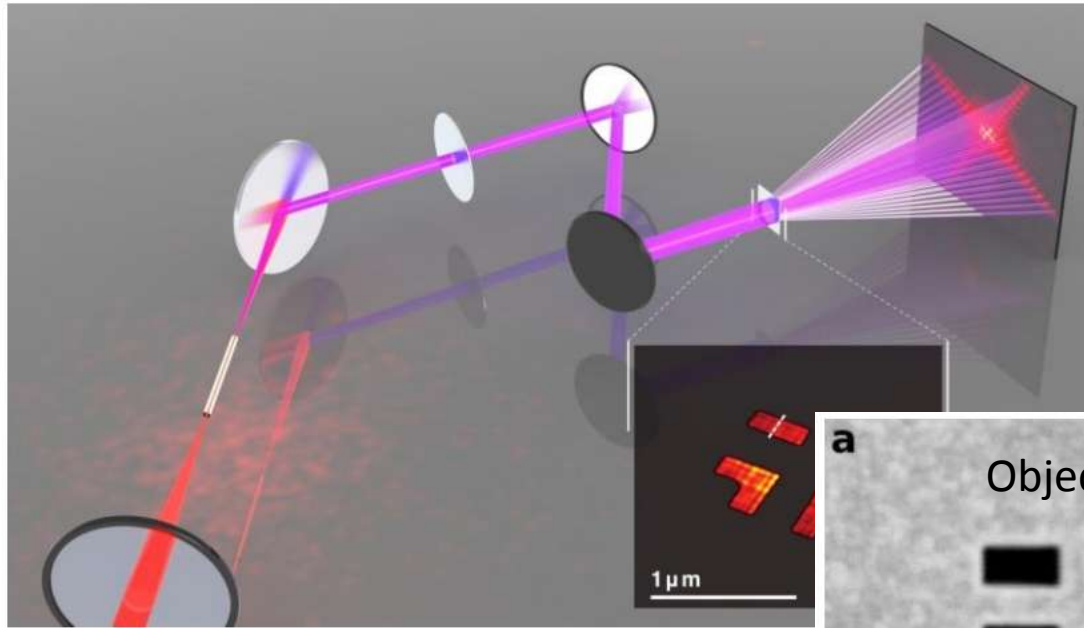


High-order Harmonic Generation (HHG)

typically nJ/pulse, kHz rep. rate, EUV/SXR photons
100' as-fs time duration of each pulse



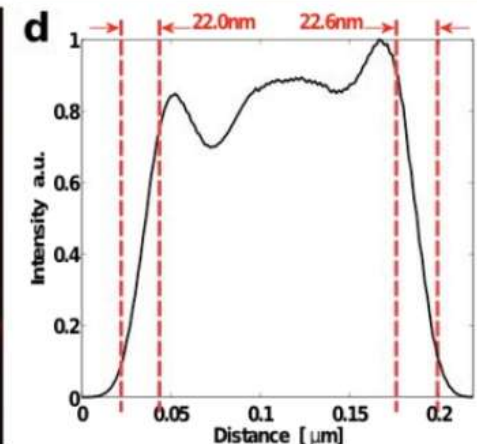
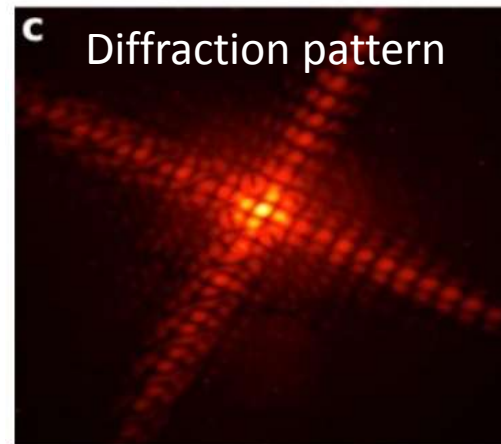
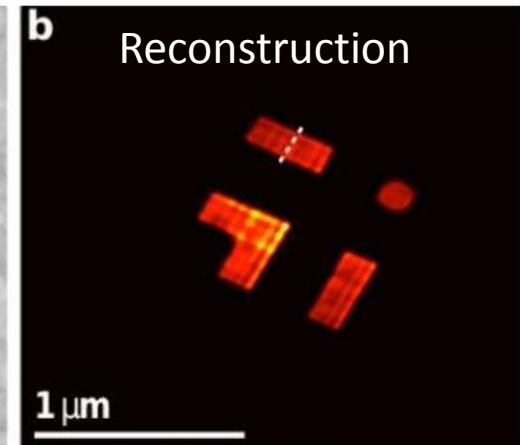
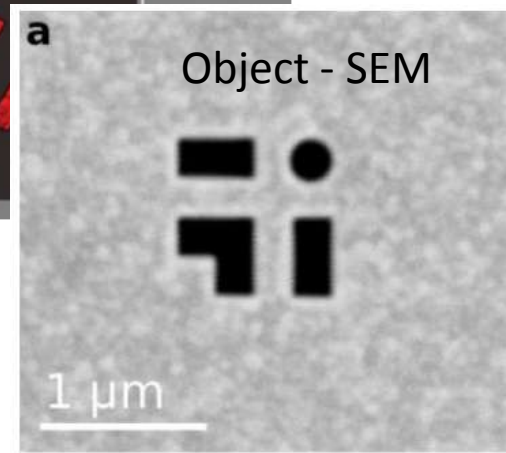
22nm resolution diffraction imaging with HHG



Rayleigh resolution = **22nm** @
 $\lambda=13\text{nm}$

- A femtosecond laser is focused into a gas-filled waveguide.
- Bright, coherent 13 nm HHG beam is produced and focused into the sample.
- the diffraction pattern is captured on a CCD camera
- the image is retrieved using an iterative phase retrieval algorithm.

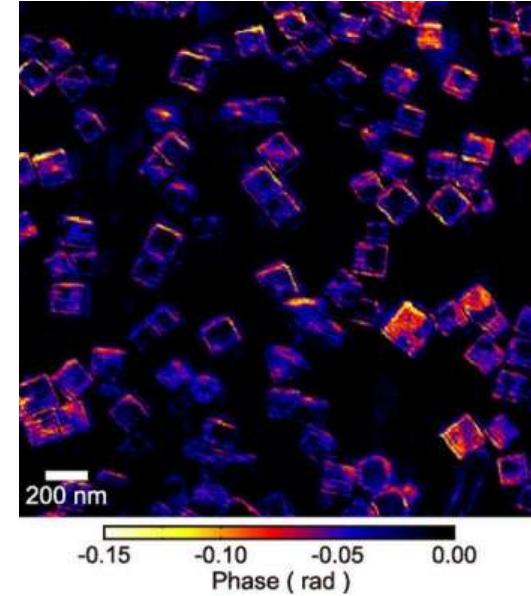
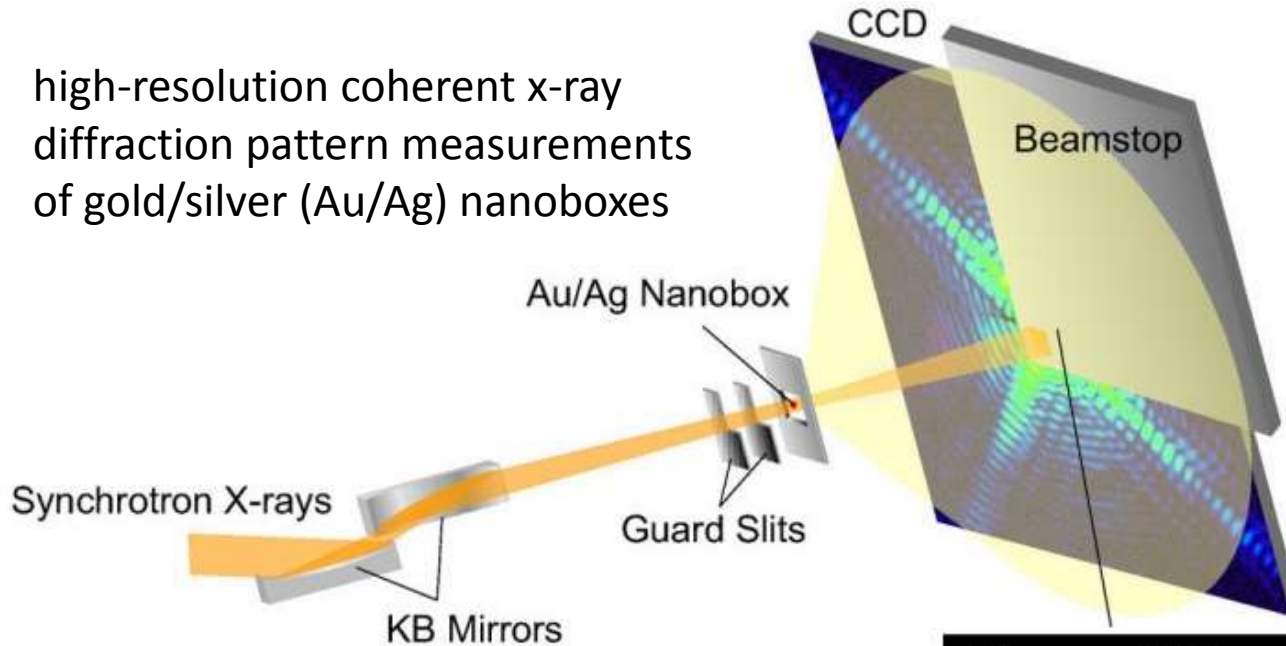
M. D. Seaberg Optics Express, 19,
23, 22470



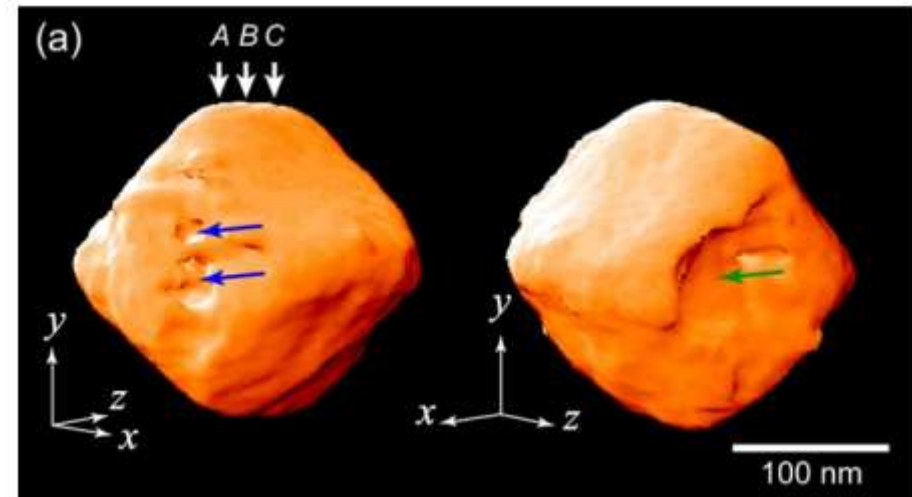
Coherent diffraction imaging with focused hard x-rays achieves sub-10nm resolution



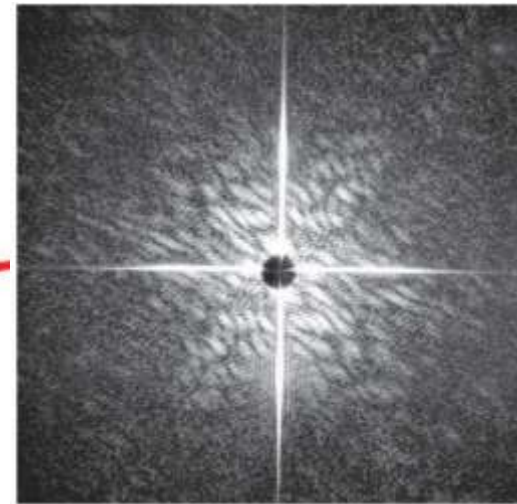
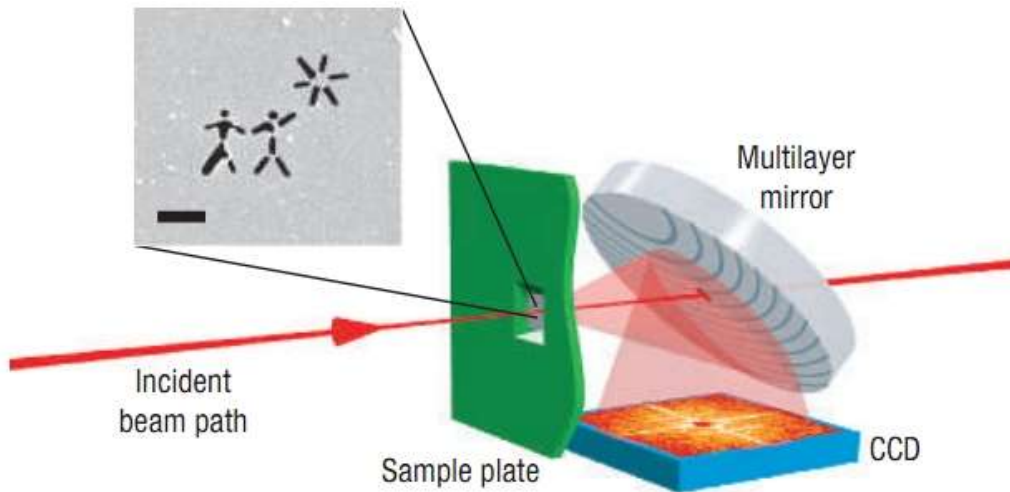
high-resolution coherent x-ray
diffraction pattern measurements
of gold/silver (Au/Ag) nanoboxes



Isosurface rendering of a 3D
reconstruction of a single
Au/Ag nanobox viewed from two
directions, showing small pits
(blue arrows) and a depression
(green arrow)

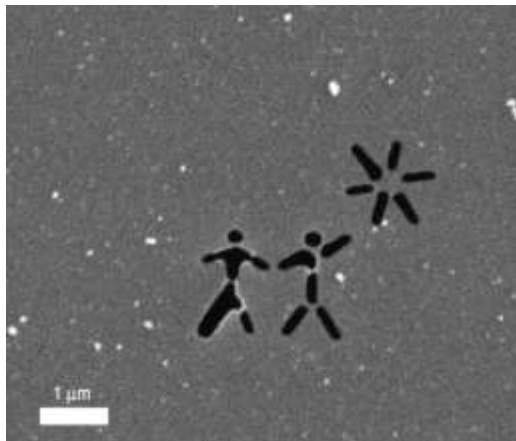


Femtosecond diffractive imaging with a soft-X-ray free-electron laser



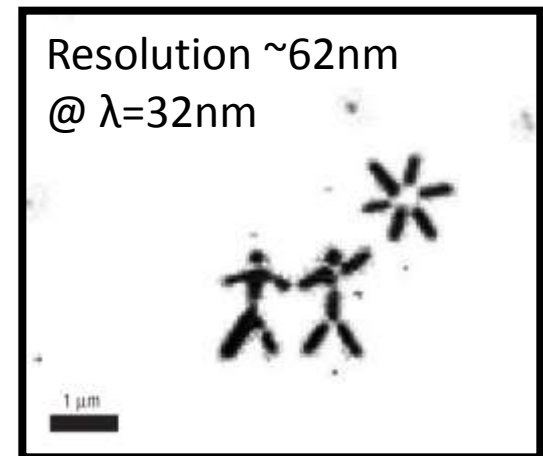
Schematic diagram of the experimental apparatus

Coherent diffraction pattern recorded for a single $(4\pm 2)\times 10^{14}$ Wcm^{-2} , 25 ± 5 fs pulse



SEM image of the sample before exposure to the FEL beam

Image reconstructed, from the ultrafast coherent diffraction pattern



Resolution $\sim 62\text{nm}$
@ $\lambda=32\text{nm}$

Ptychography – High resolution Scanning Diffraction Microscopy

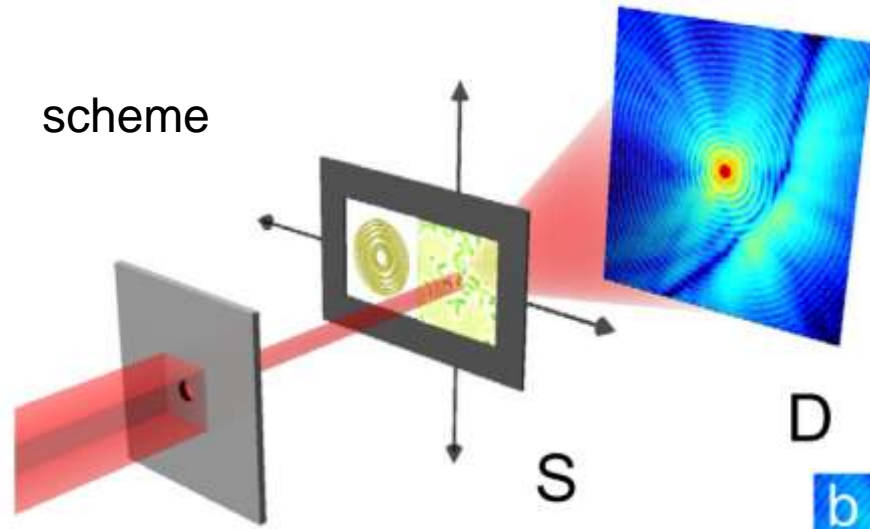
ptychographic imaging method bridges the gap between CDI and STXM by measuring a set of diffraction patterns at each point of a STXM scan.

Advantages:

- Very high resolution (for HXR ~10-20nm) for test samples. Resolution limited by geometry,
- lack of optics allows for such high resolution
- contrast for phase samples

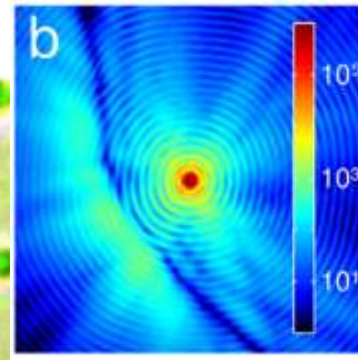
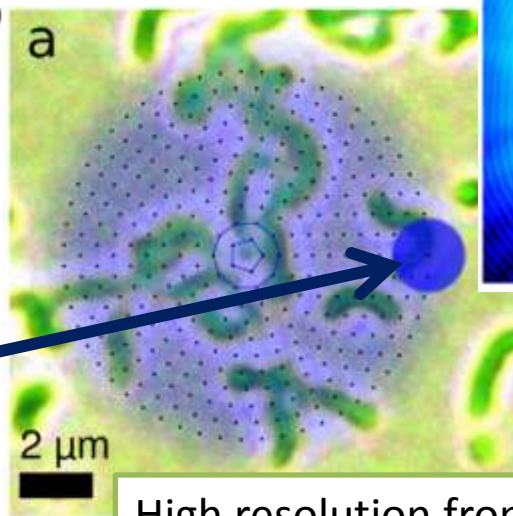
Disadvantages:

- Challenging to reconstruct

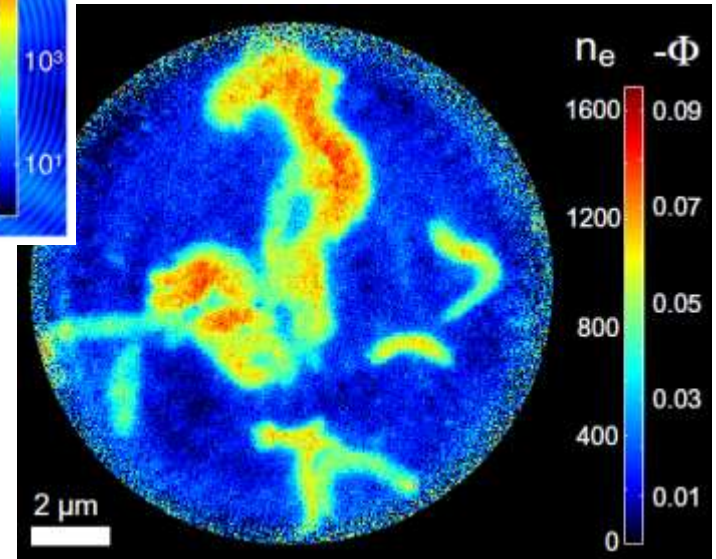


Typical scanning positions

X-ray spot much larger than resolution

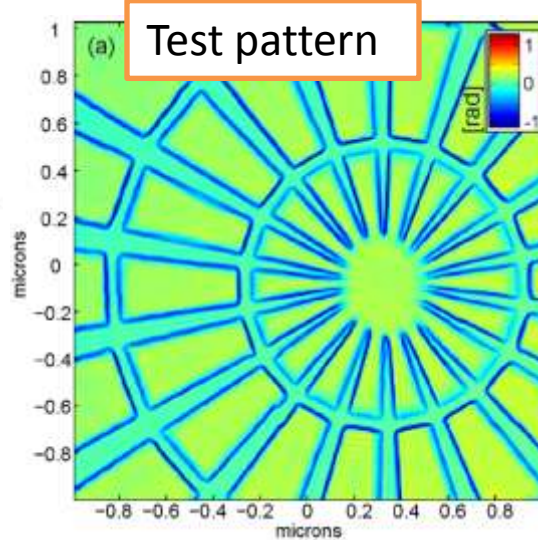
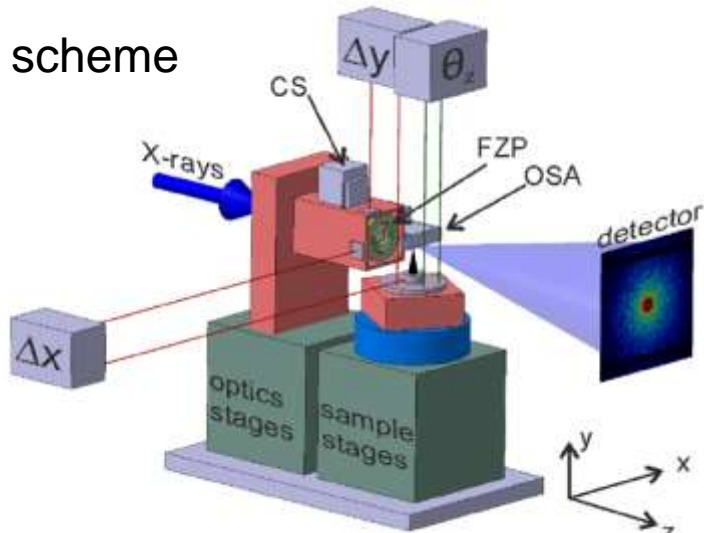


High resolution from heavily dversampled set of data

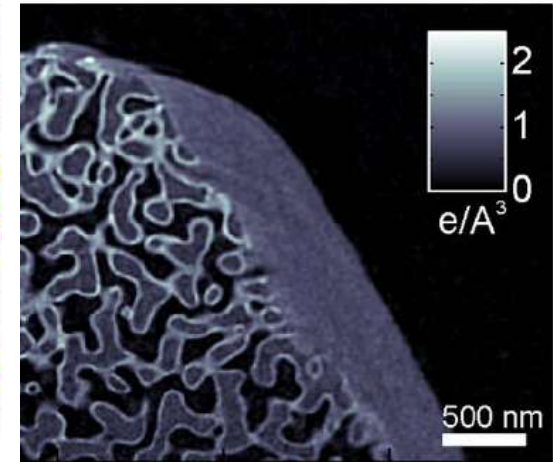


Reconstruction results

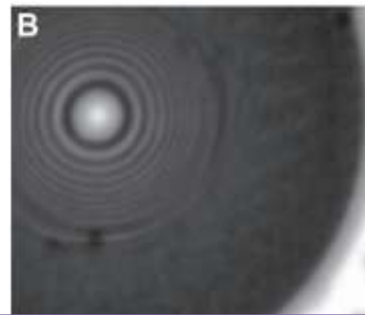
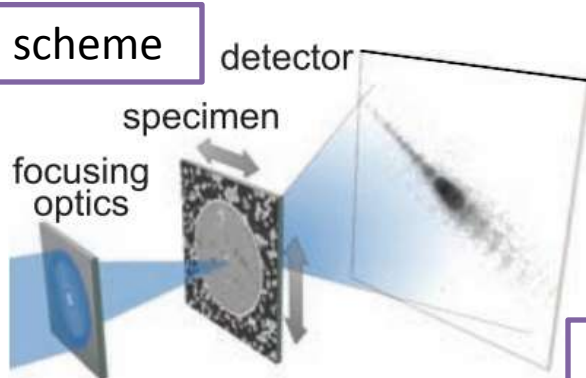
X-ray ptychographic computed tomography at 16 nm isotropic 3D resolution



Part of 3-D reconstr.

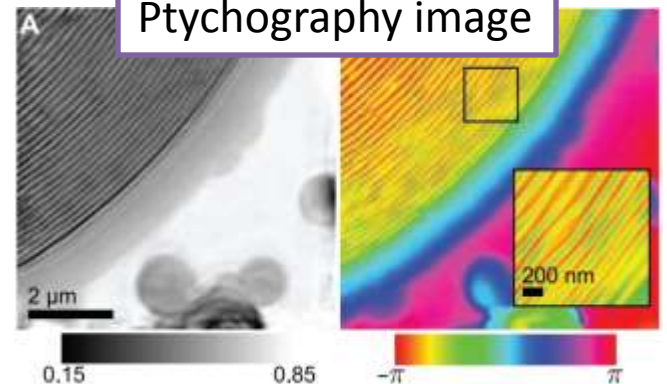


Holler et al., Scientific Reports, 4, 3857 (2014)



Transmission image

FZP
dr=70nm



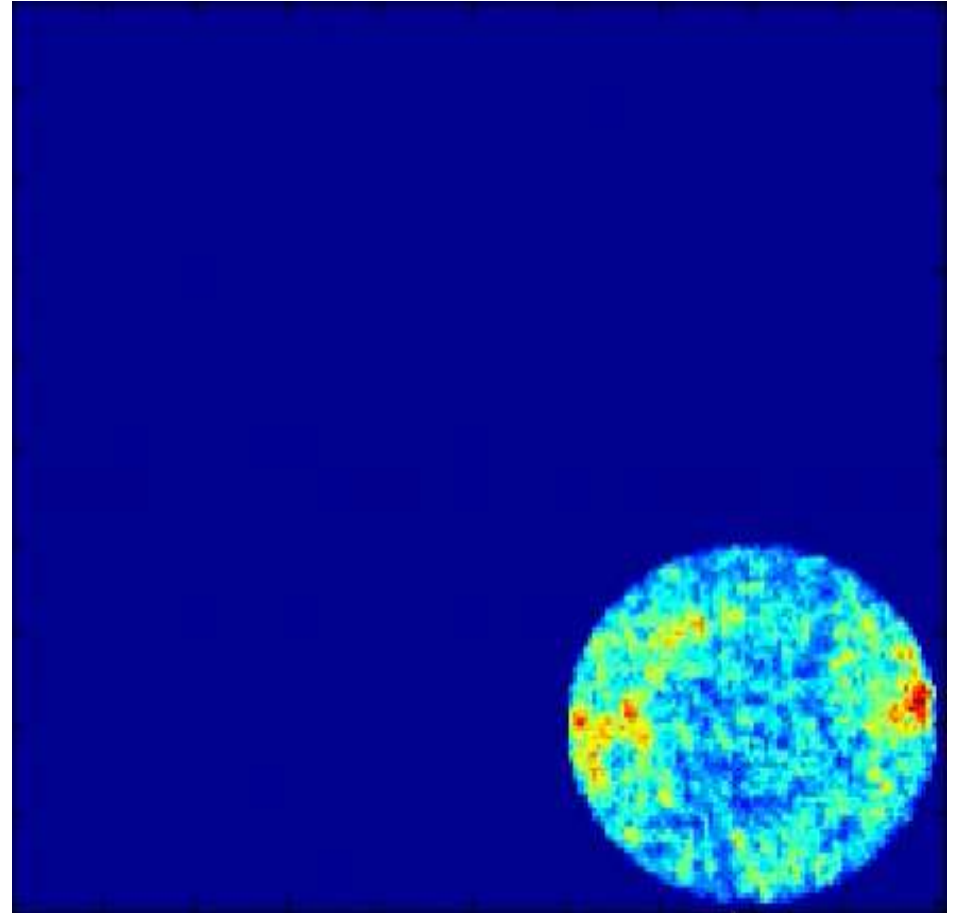
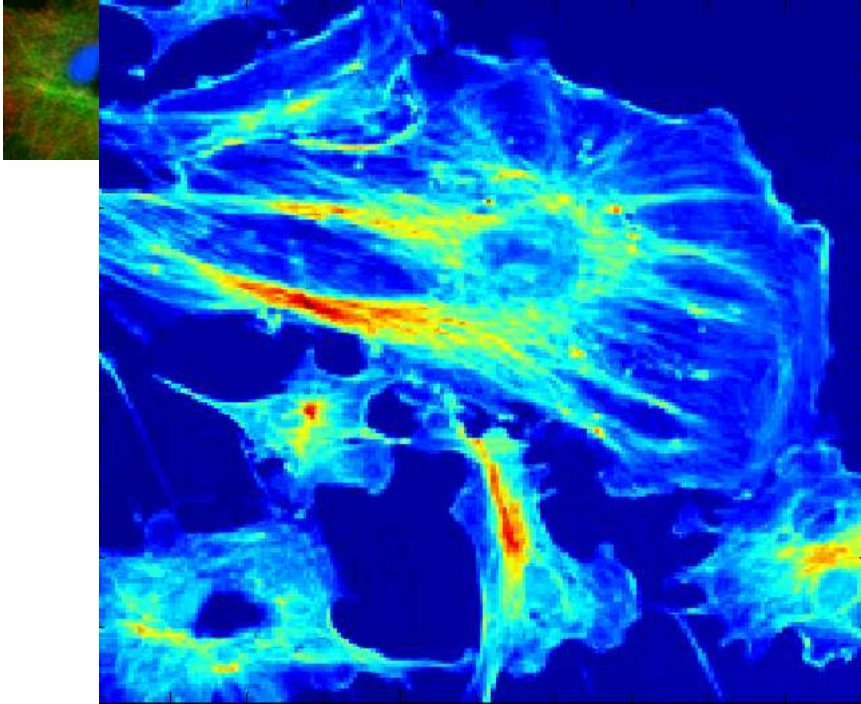
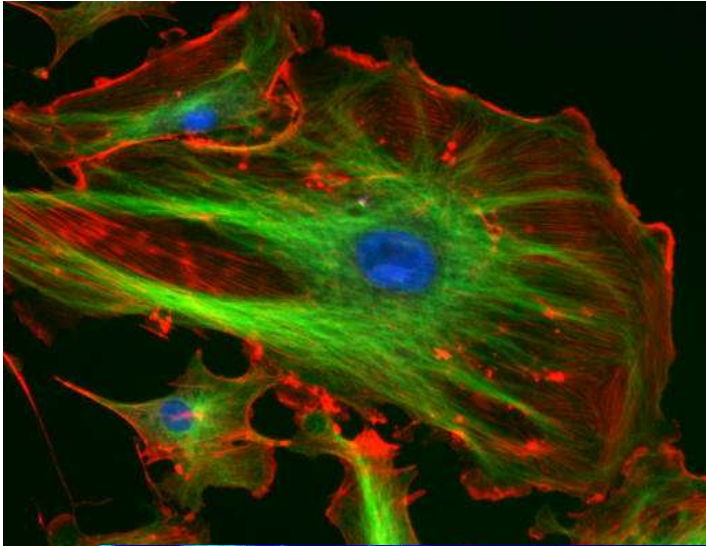
Thibault et al., Science **321**, 5887, 379-382 (2008)

Algorithms:

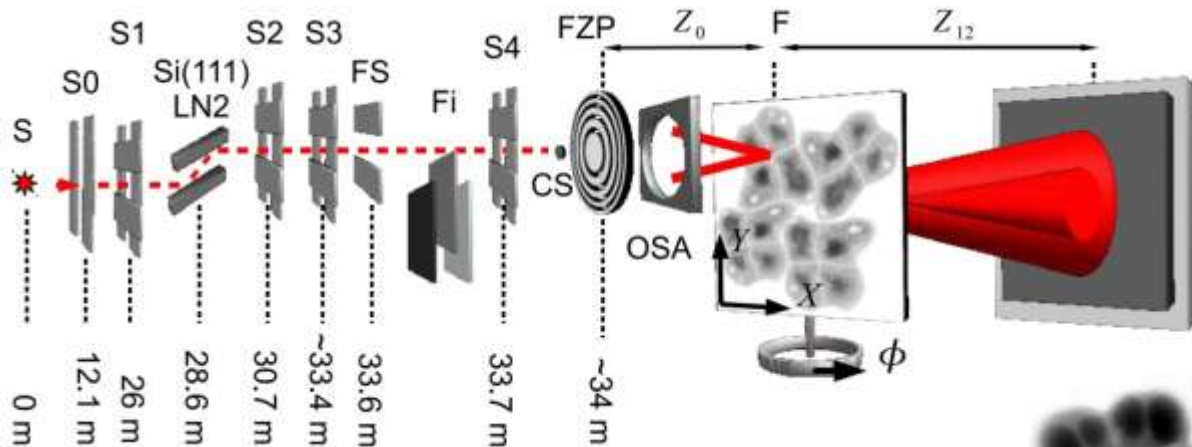
- ❑ „ePIE” (Maiden and Rodenburg, Ultramicroscopy 109, 1256 (2009))
- ❑ Difference Map (Thibault et al., Science 321, 379 (2008))
- ❑ Conjugate gradient (Guizar-Sicairos et al., Opt. Expr. 16, 7264 (2008))

e-PIE algorithm

Diff patterns on a 6x6 grid, beam size $D=80$ pix,
8 iterations.



Hard X-ray imaging of bacterial cells using ptychography

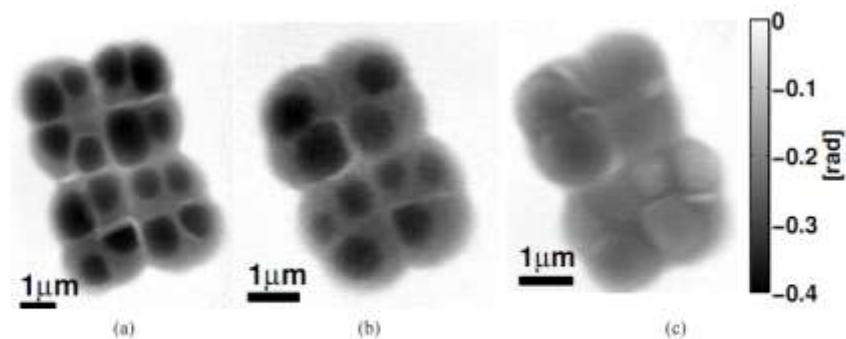


cSAXS - X12SA:
Coherent Small-Angle X-ray
Scattering Beamline

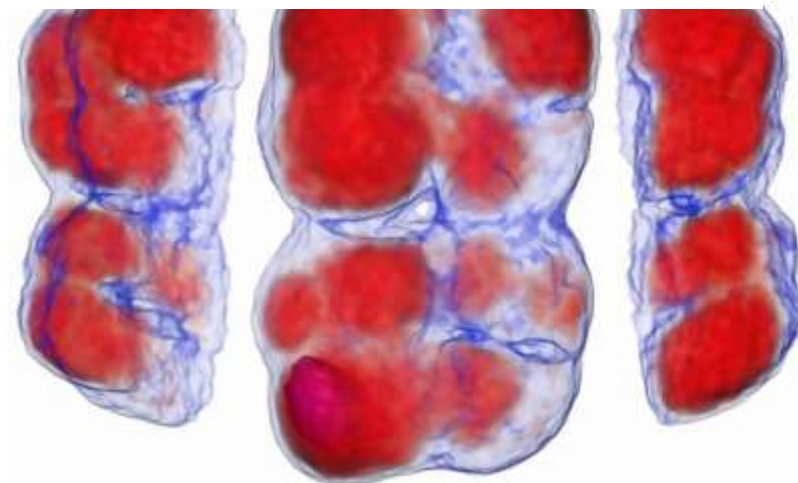
Probe size: $0.75 \times 1.5 \mu\text{m}^2$

Spatial resolution $\sim 90 \text{nm}$ @ 6.2keV

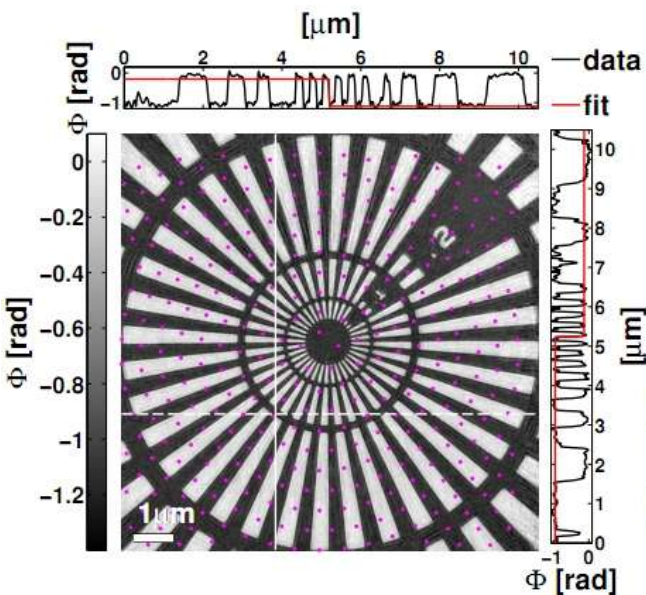
Au on Si_3N_4 FZP, $\text{dr} = 100 \text{nm}$



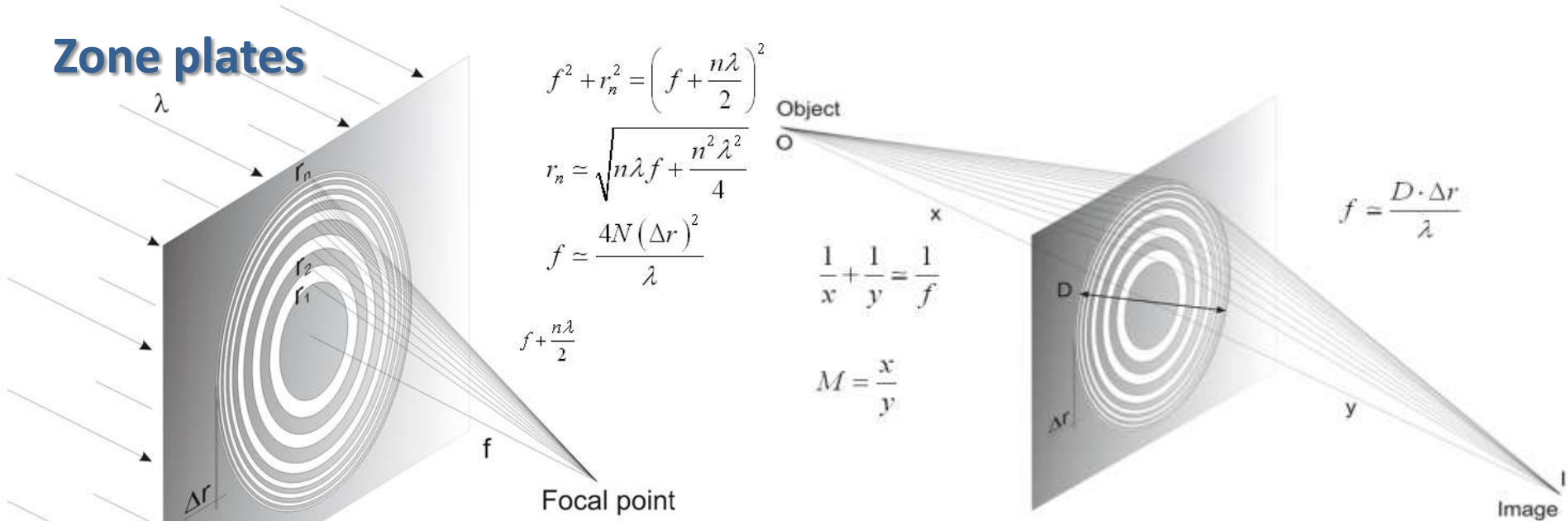
Phase reconstructions of *D. radiodurans*



- Multiple
diffraction
patterns enhance
**resolution beyond
the probe size**



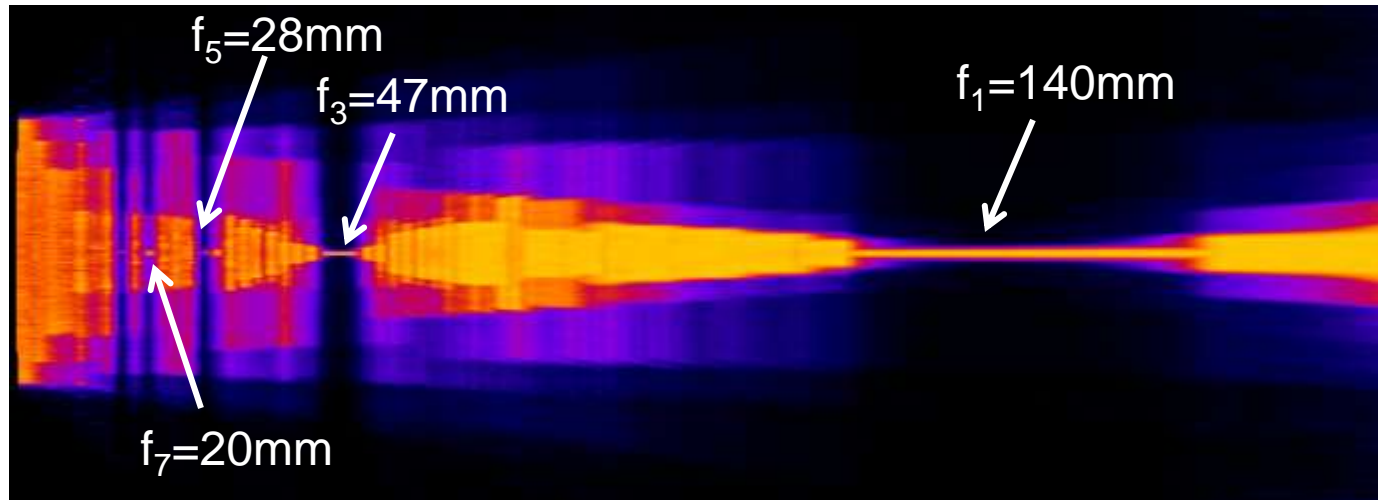
Zone plates



- use diffraction of light
- highly chromatic devices
- used typically to focus light and in the imaging scheme
- very small, thin and fragile

MATLAB + Image J

$D=200\mu\text{m}$,
 $dr=3.5\mu\text{m}$, $\lambda=5\text{nm}$



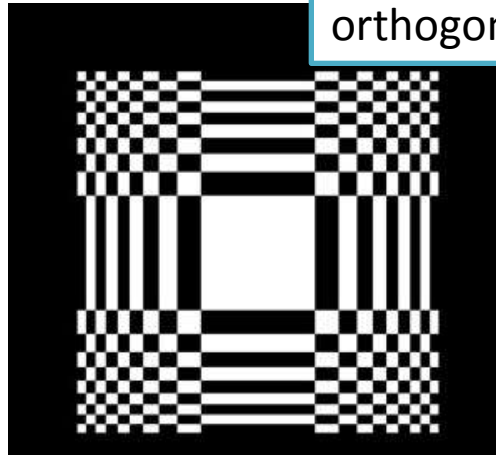
Various Zone Plates

Free standing



www.cxro.lbl.gov

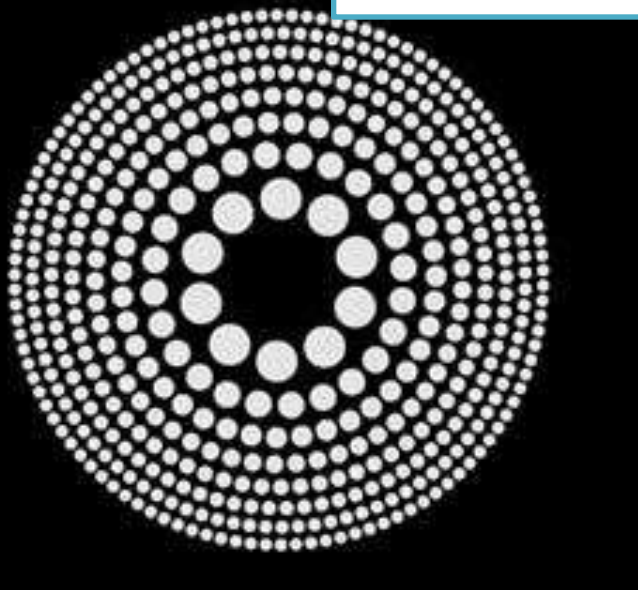
orthogonal linear ZP -
orthogonal Fresnel array



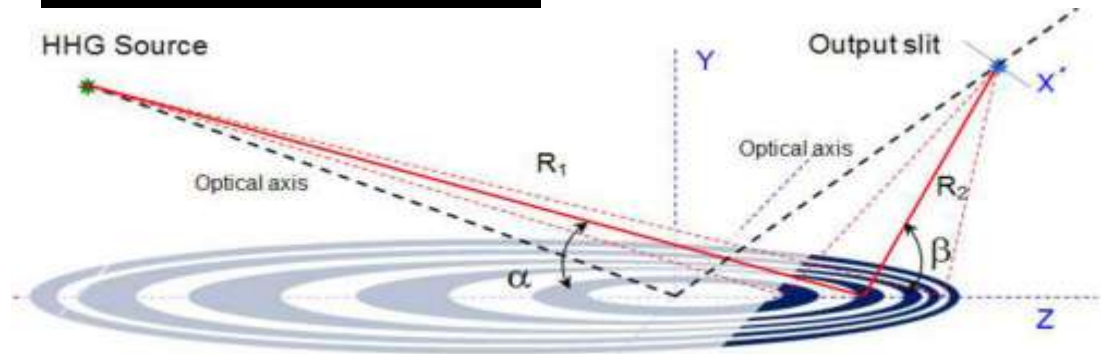
http://bonryu.com/atelier_bonryu_e/ZP_Salon_5.html

J. Metje, et al., Optics Express 22, 9, 10747-10760 (2014)

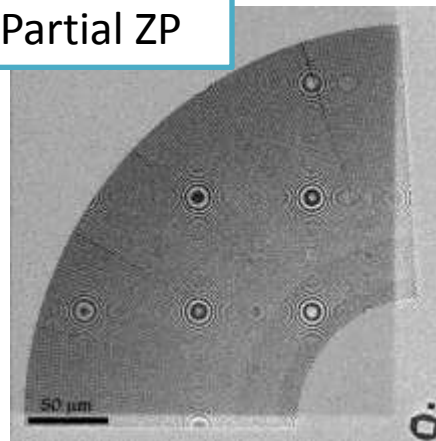
Photon Sieve



<http://www.oocities.org/penate@rogers.com/sieve/photonsieve.html>

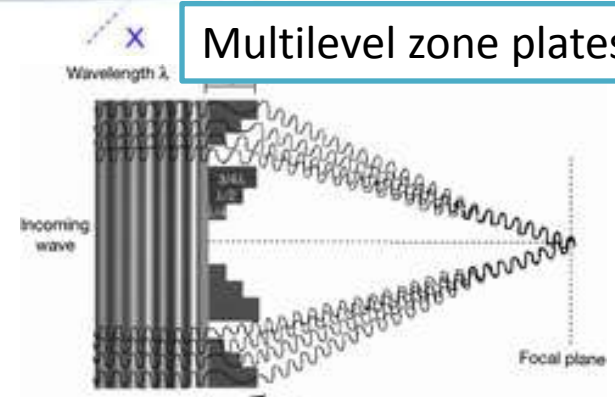


Partial ZP



www.zoneplates.ltd

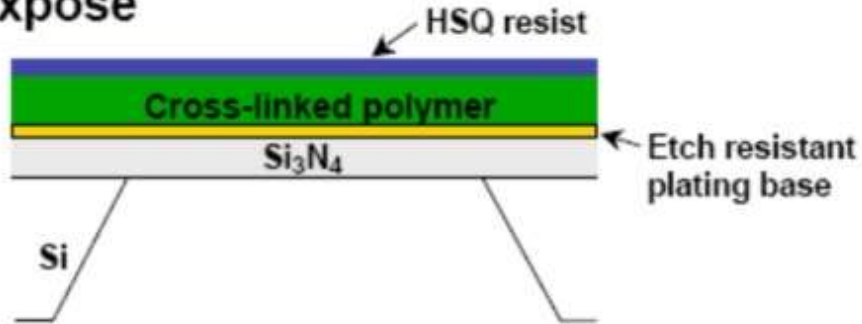
Multilevel zone plates



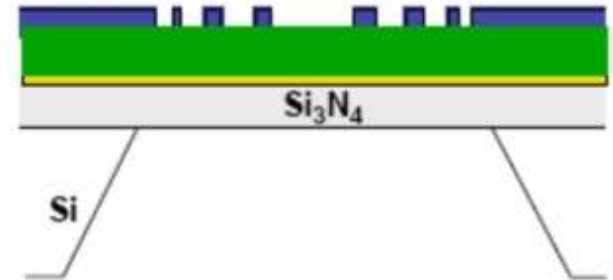
E. Di Fabrizio et al., Nature 401, 895-898, 1999

Zone plates - Fabrication for 46.9nm

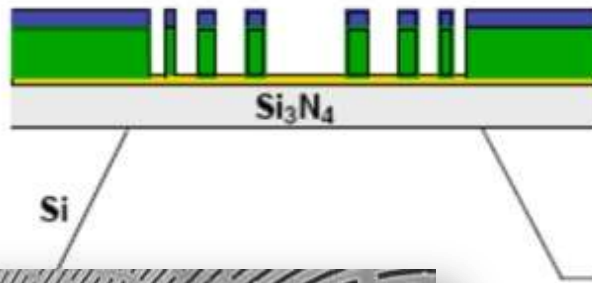
1. Expose



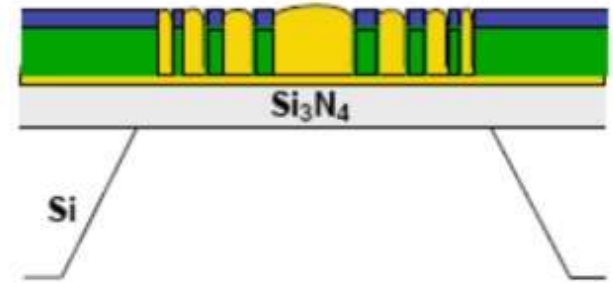
2. Develop



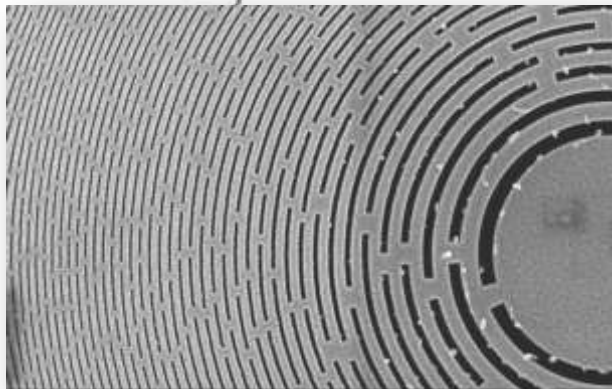
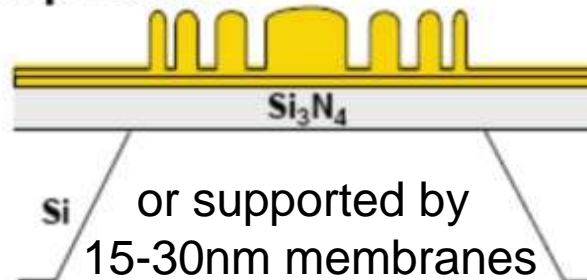
3. Cryogenic ICP Etch



4. Plate



5. Strip Resist



Free-standing design

*E. H. Anderson, IEEE JQE
42, 27 (2006)*

Zone plates for $\lambda=13.8\text{nm}$

PMMA: 200 nm

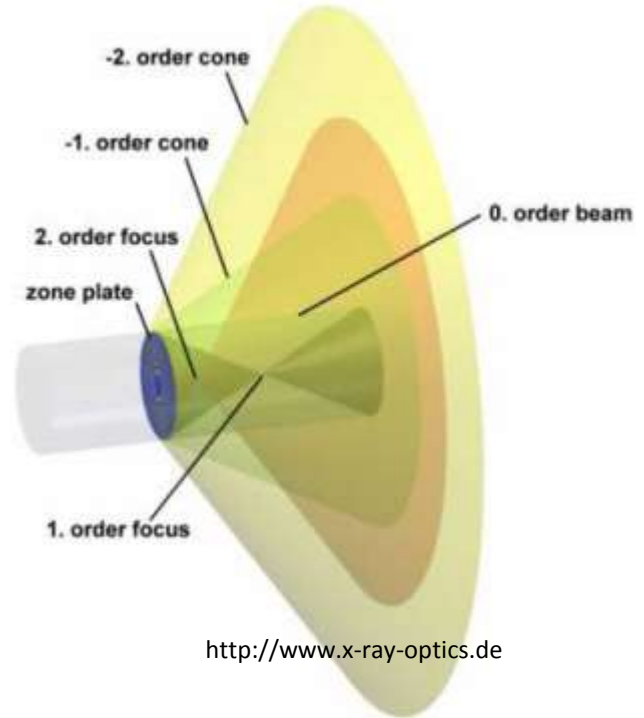
SiNx: 100 nm

$$r_n^2 = n\lambda f + \frac{n^2\lambda^2}{4}$$

$$NA \ll 1$$

$$r_n \simeq \sqrt{n\lambda f}$$

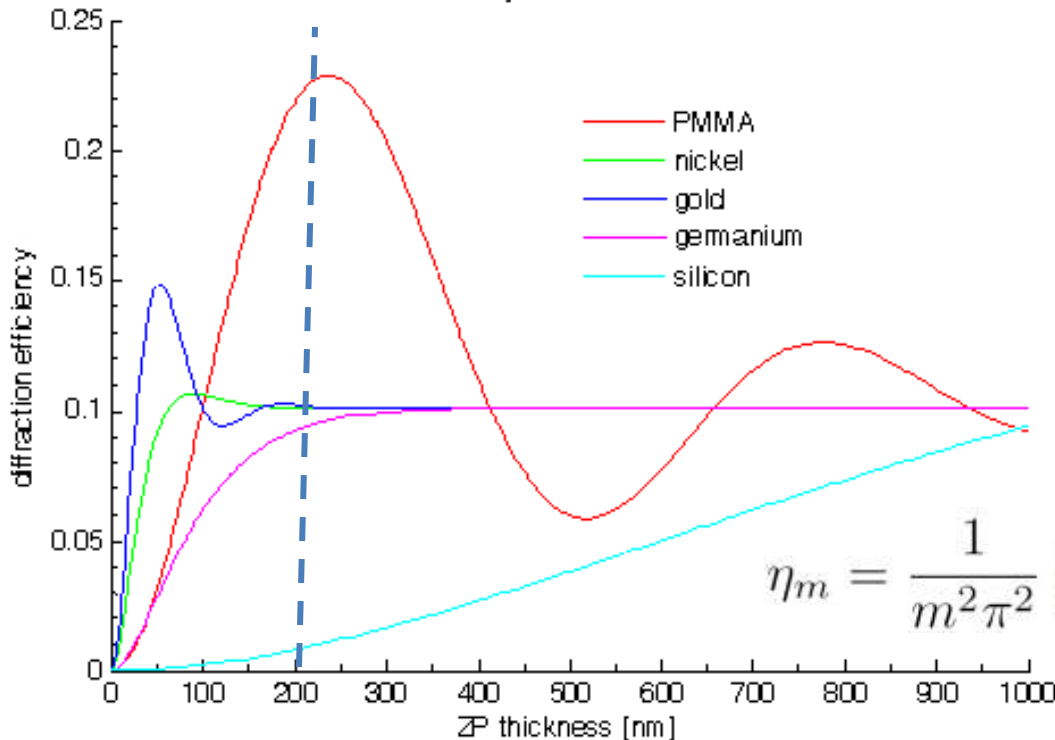
10 mm



<http://www.x-ray-optics.de>

Dr. H. T. Kim and
Dr. S. Ch. Jeon
Korea

diffraction efficiency for ZP materials @13.84nm



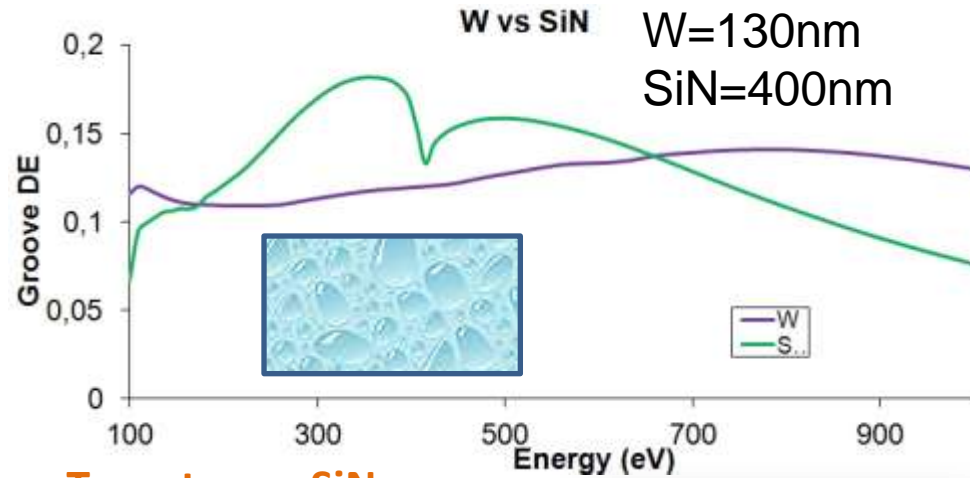
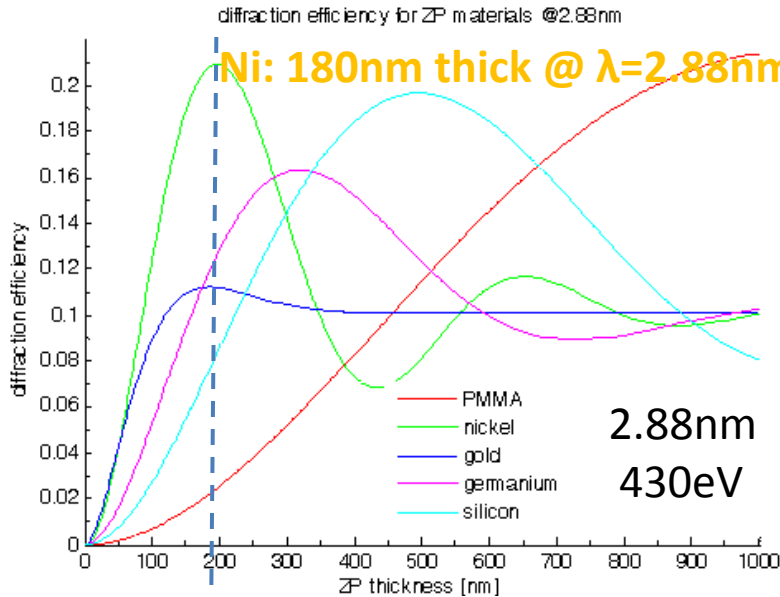
$$\eta_m = \frac{1}{m^2\pi^2} \left(1 + e^{-2\phi\beta/\delta} - 2e^{-\phi\beta/\delta} \cos \phi \right)$$

$$\phi = 2\pi t\delta/\lambda$$

J. Kirz, *JOSA* 64, 301 (1974)

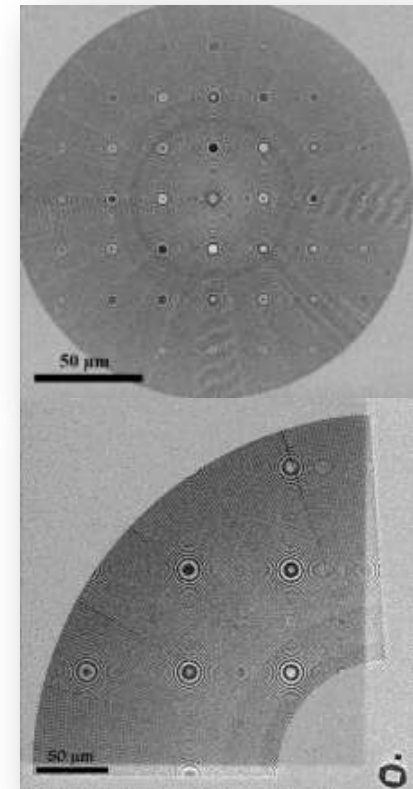
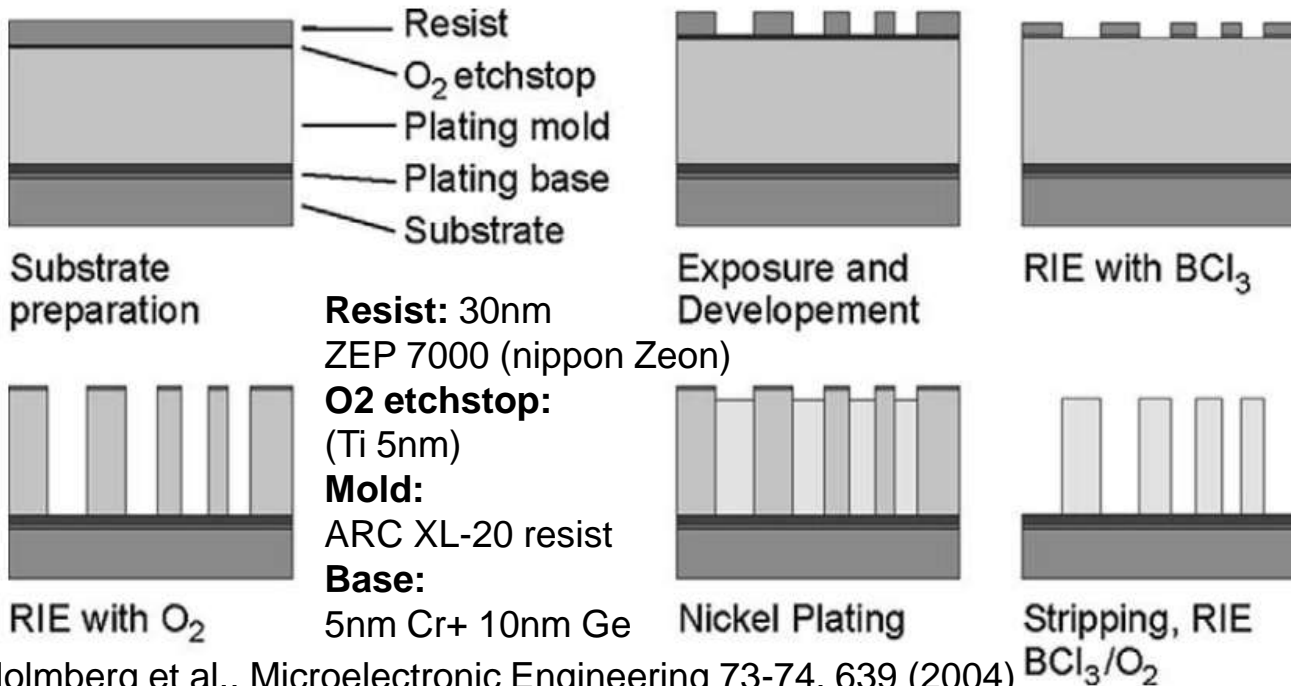
Zone plates for „water window”

Thickness:
W=130nm
SiN=400nm



Tungsten or SiN

<http://www.zoneplates.com/index.html>



Desktop-size capillary discharge $\lambda=46.9$ nm EUV laser



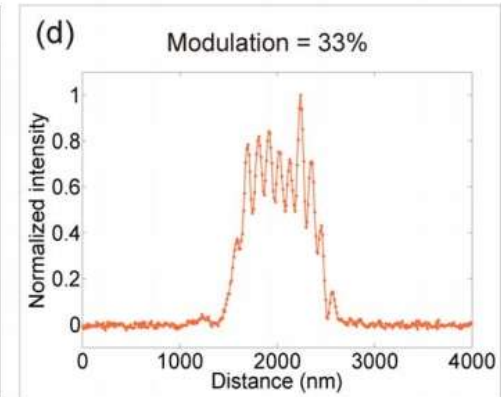
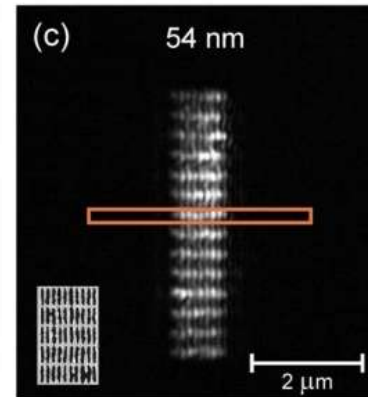
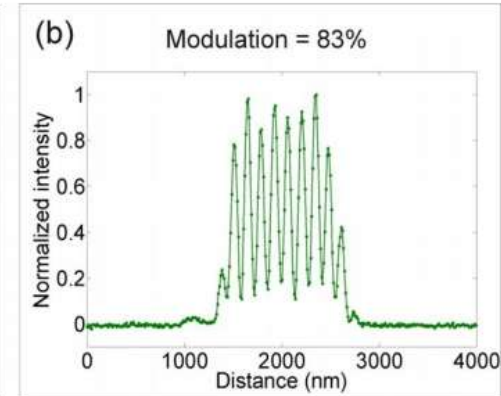
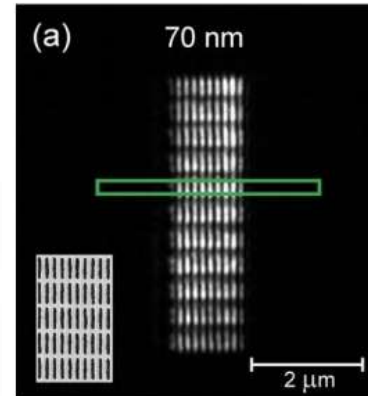
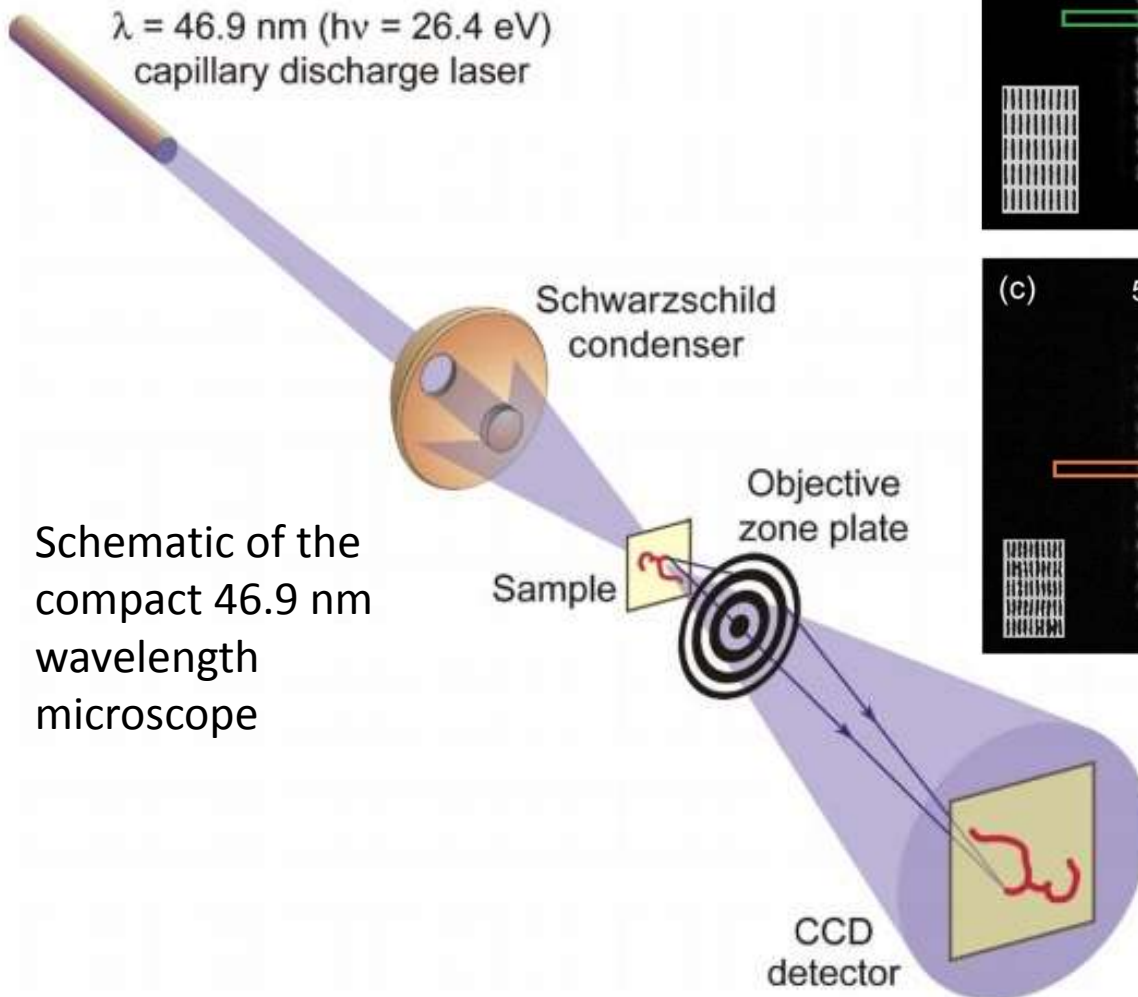
Laser Output Characteristics:

- Output Energy:
> 10 $\mu\text{J}/\text{pulse}$
- Average power:
0.15 mW
- Repetition rate:
up to 12 Hz
- Pulse duration:
 ~ 1.5 ns
- Bandwidth:
 $\Delta\lambda/\lambda < 1 \times 10^{-4}$

S. Heinbuch , et al., Opt. Exp. 13, 4050 (2005)

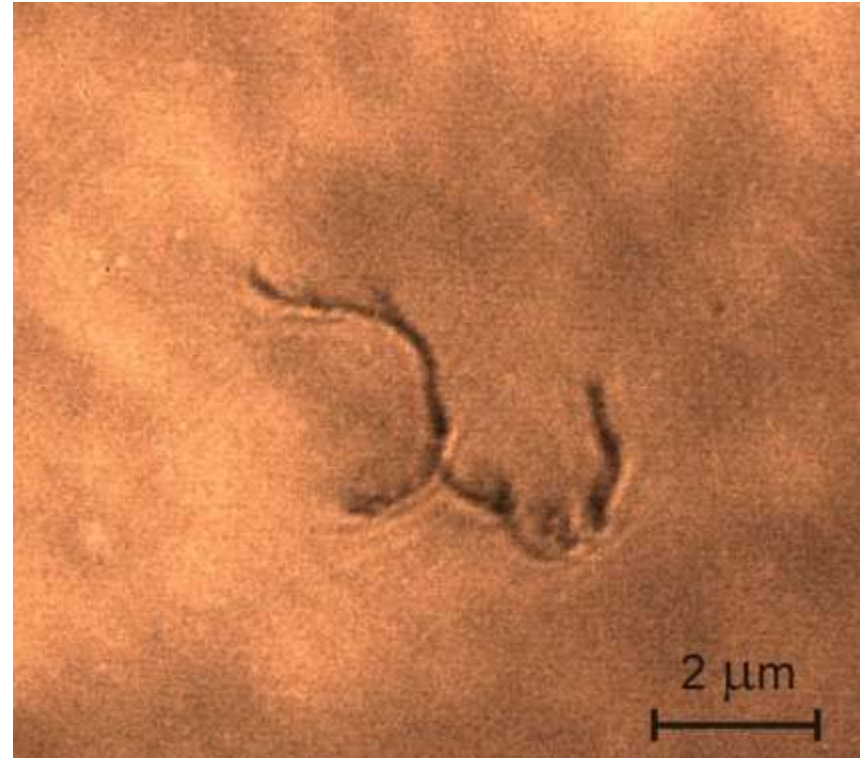
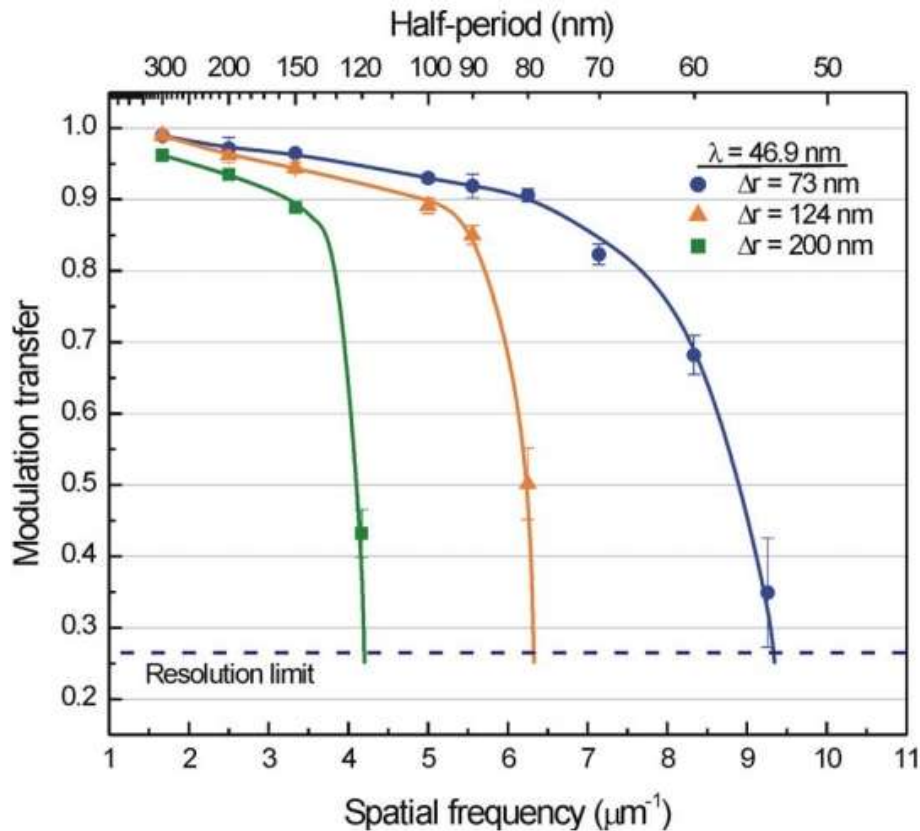
Courtesy of Courtney Brewer, Colorado State University

Single-shot EUV laser imaging of nanostructures with wavelength resolution



Single-shot EUV images

Single-shot EUV laser imaging of nanostructures with wavelength resolution



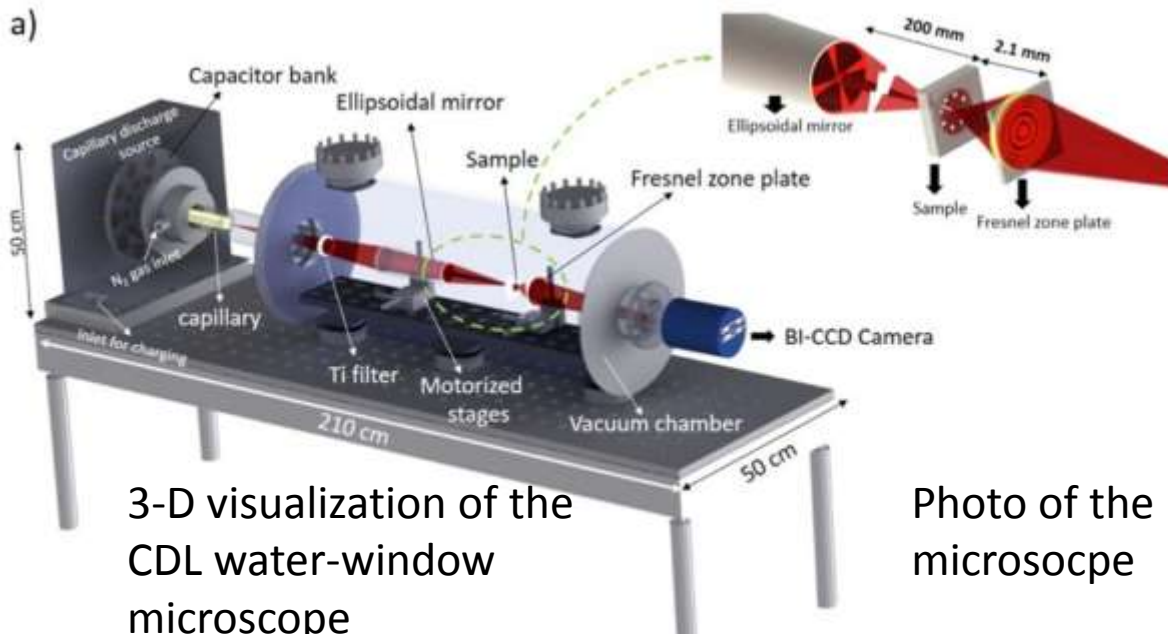
Measured MTFs for objective zone plates with outer zone widths of $\Delta r=200$, 124, and 73 nm (NA=0.12, 0.19, and 0.32, respectively).

Rayleigh-like resolution values of 120, 80, and 54 nm

Single-shot image of an entanglement of 50 nm diameter carbon nanotubes. This image was obtained using the $\Delta r=73 \text{ nm}$ objective lens and a wavelength of 46.9 nm.

Water – window CDL microscope (CTU Prague)

a)



3-D visualization of the CDL water-window microscope

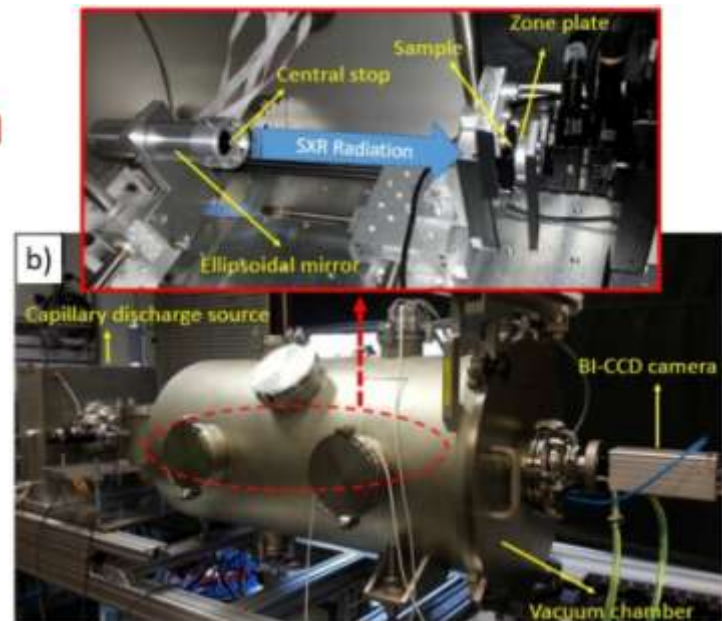
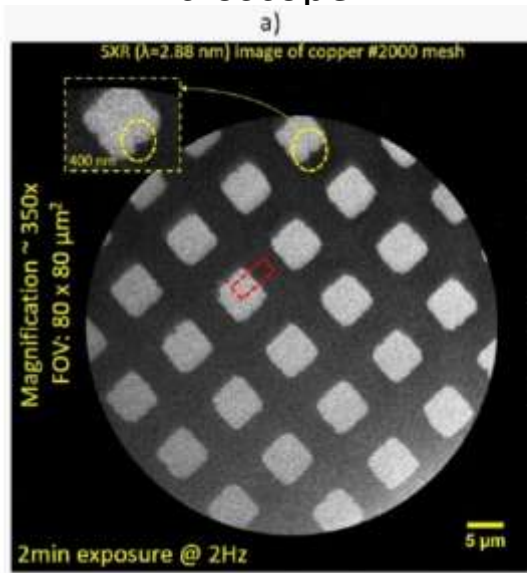
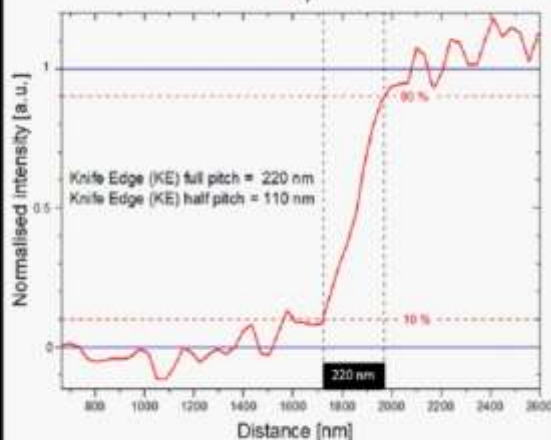


Photo of the microscope



Spatial resolution 110nm



Test object and spatial resolution

Number of photons: $\sim 1.2 \times 10^9$ /pulse

@ $\lambda = 2.88 \text{ nm}$

Energy: 83 nJ/pulse and at 3-Hz repetition rate

FZP objective

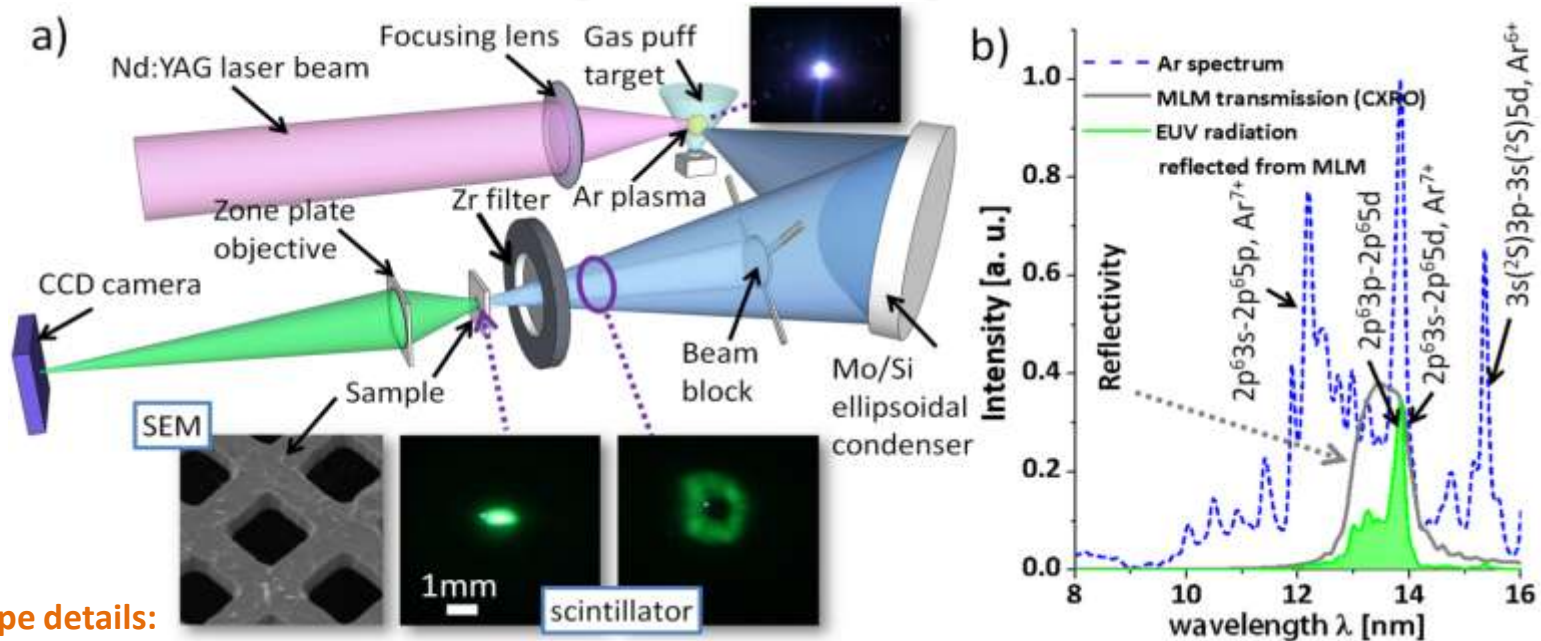
- tungsten zoneplate 130 nm thick

- $D = 180 \mu\text{m}$, $\Delta r = 33 \text{ nm}$,

- $f = 2.1 \text{ mm}$, $\lambda = 2.88 \text{ nm}$, $NA_{O_{in}} = 0.043$

- magnification: $350 \times$

A 50nm spatial resolution EUV imaging with FZP and gas puff target based source



Microscope details:

CCD camera: iKon-M (Andor) with 1k x 1k pix, $13 \times 13 \mu\text{m}^2$

Condenser NA: $H=0.11, V=0.15$

Objective NA: 0.137

Magnification: (min) 470x

Pixel size: (max) 27.7nm

Acquisition time: 10-100sec/2Hz

Zone plate objective parameters:

D: $200 \mu\text{m}$, $\Delta r: 50 \text{ nm}$, f: 0.724 mm, N: 1000

zones, NA: 0.137, Rayleigh resolution:

$1.22\Delta r=61 \text{ nm}$, DOF: $\pm 0.5\lambda/NA^2 \pm 385 \text{ nm}$

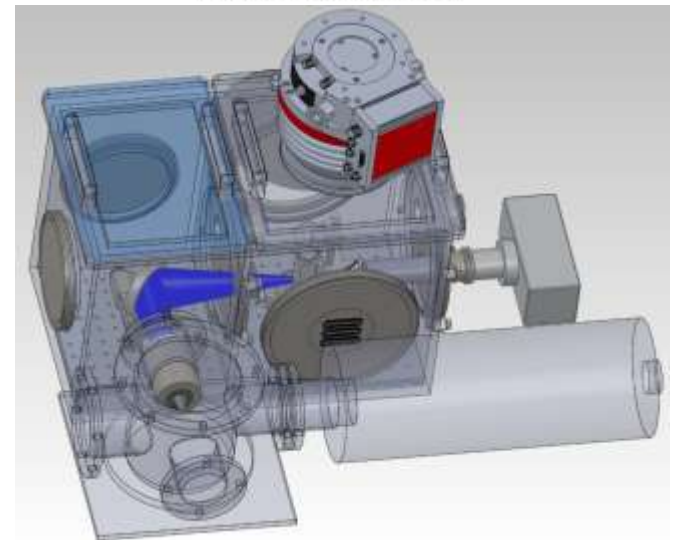


LIDER 2012 Mikroskop EUV

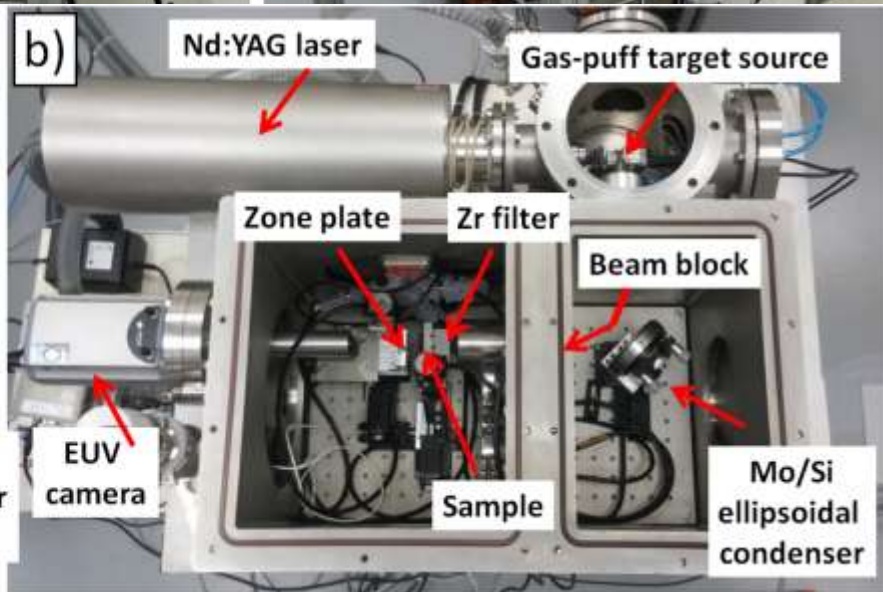
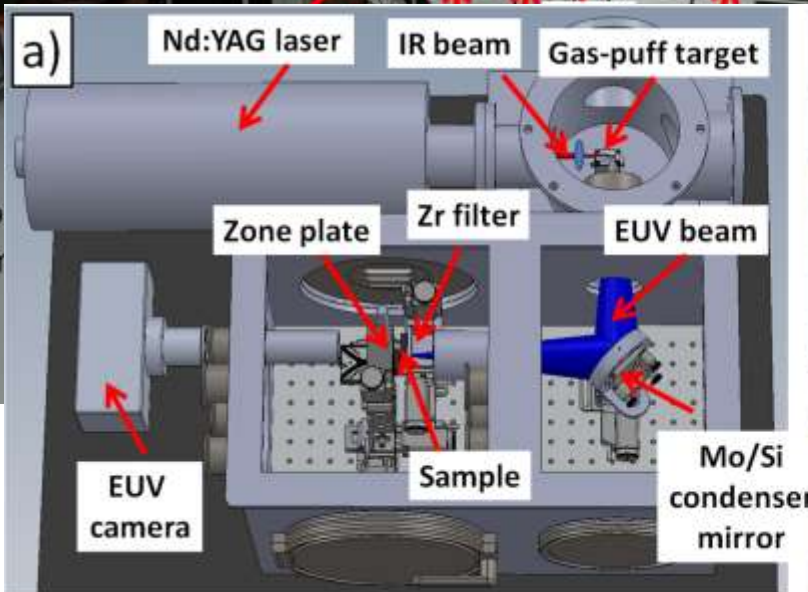
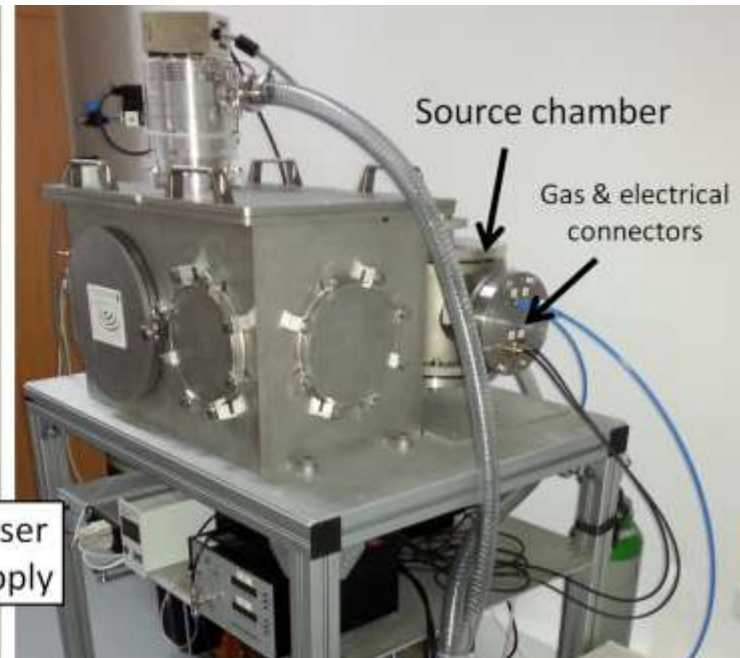
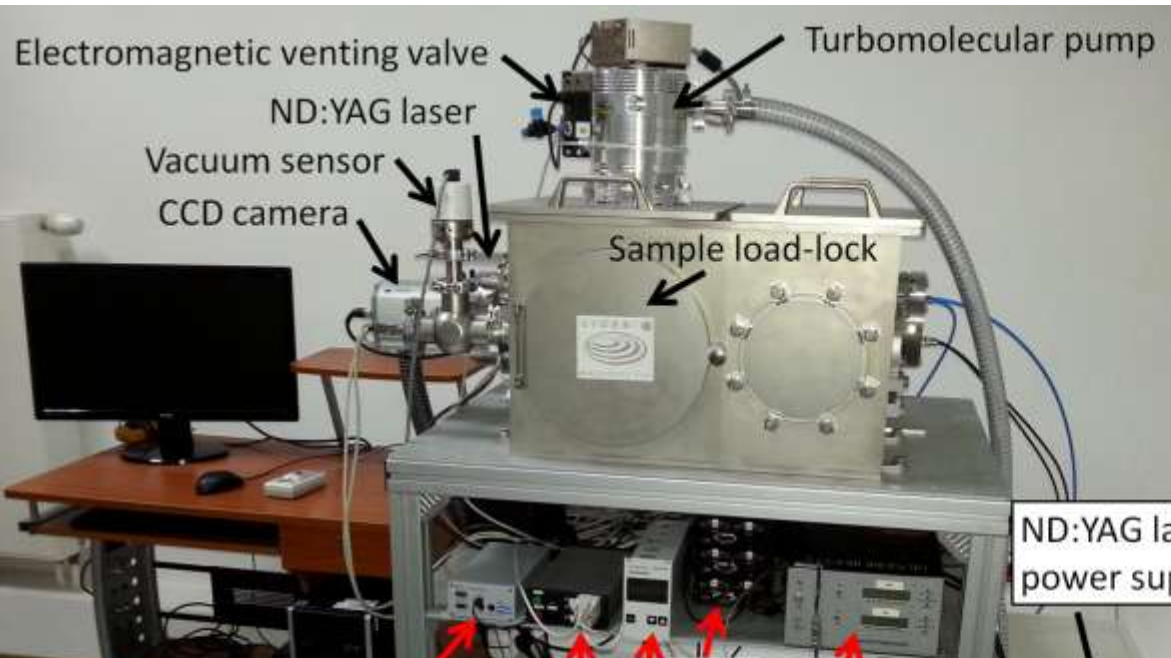
Realizacja projektu została sfinansowana przez



Narodowe Centrum Badań i Rozwoju

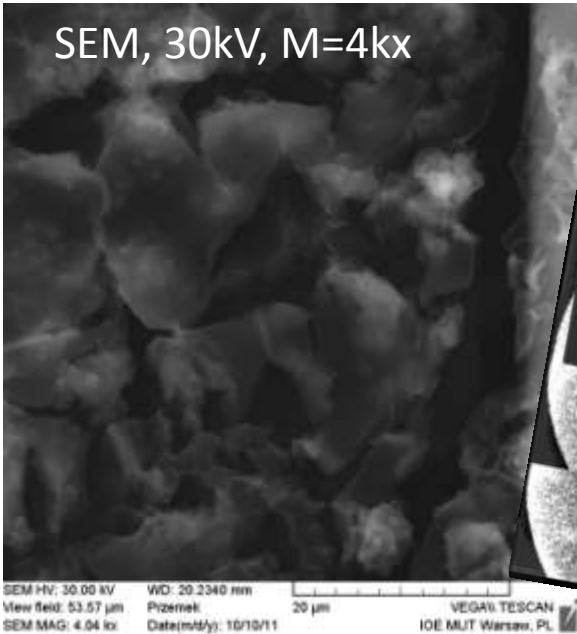


Developed EUV microscope system

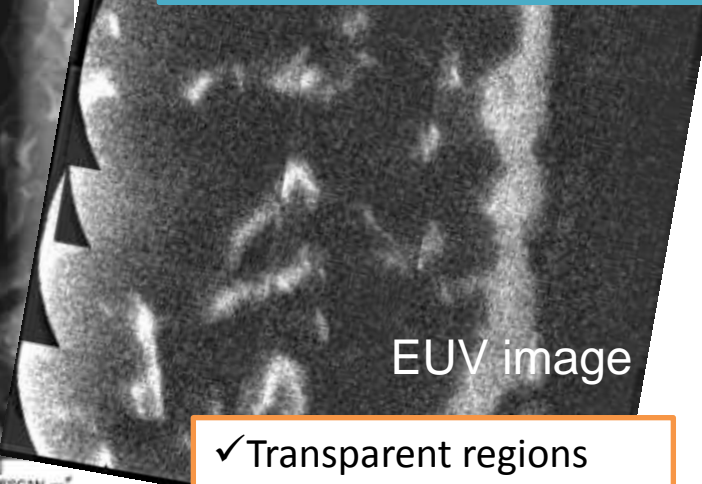


PC for the m

...studying thin films



✓ 100nm features are visible



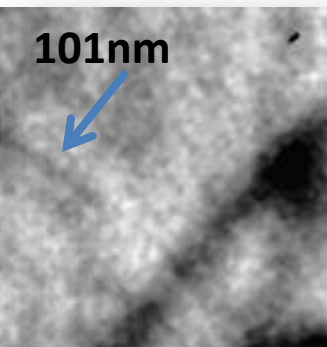
✓ Transparent regions with thickness <100nm (cracks, holes, etc.)

..imaging of nanostructures: ZnO nanowires

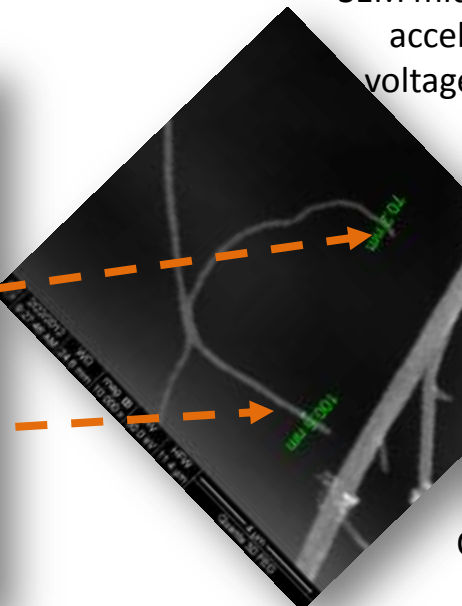
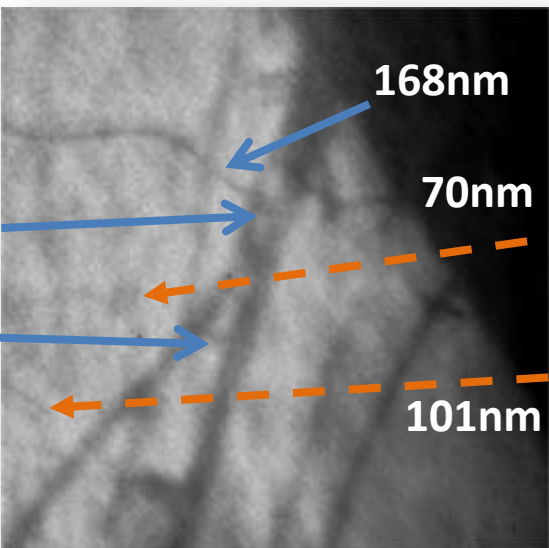
P. Wachulak et al., Radiation Physics and Chemistry 93 (2013) 54–58

EUV microscope image, 100 EUV pulses exposure

SEM micrograph at acceleration voltage of 30kV

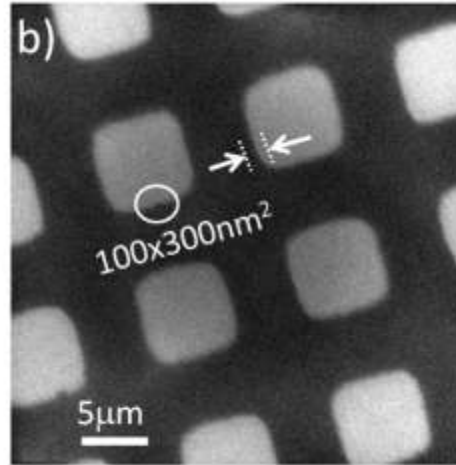
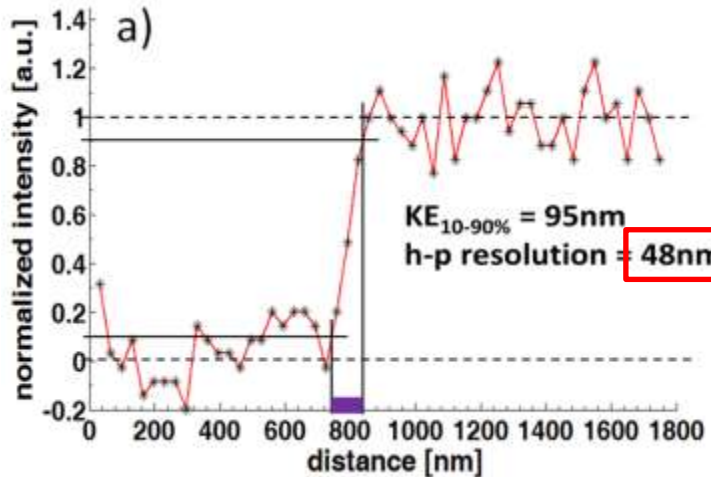


Hole 300nm
165nm



Optical micrograph of ZnO nanofibers

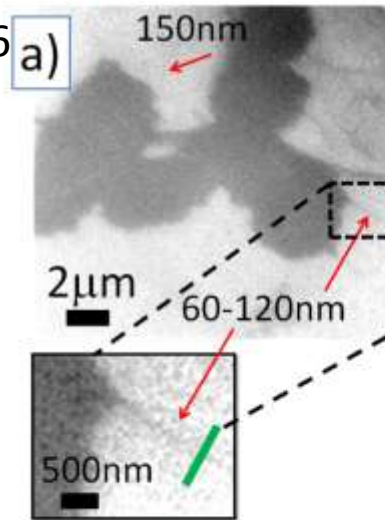
Resolution and applications



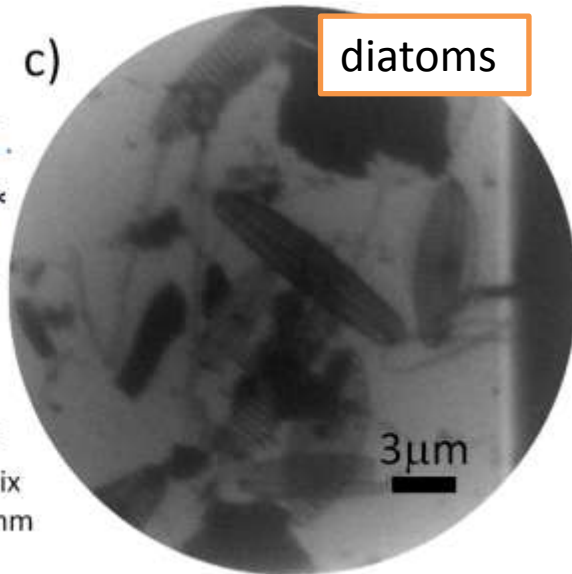
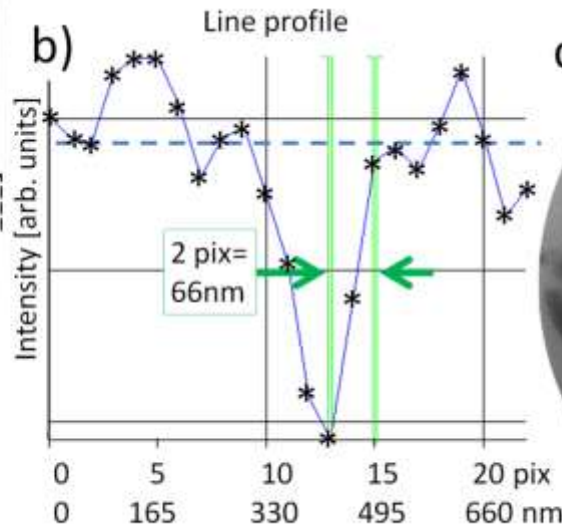
Spatial resolution estimation based on knife-edge test

A. Torrisi, et al., Journal of Microscopy 265, 2, 251-260 (2017)

EUV image of CT 26 fibroblast cells...



...obtained with exposure of 200 EUV pulses (22 seconds), with visible features as small as 60 nm.



Microscope details:

CCD camera: iKon-M (Andor) with 1k x 1k pix, 13x13 μm²

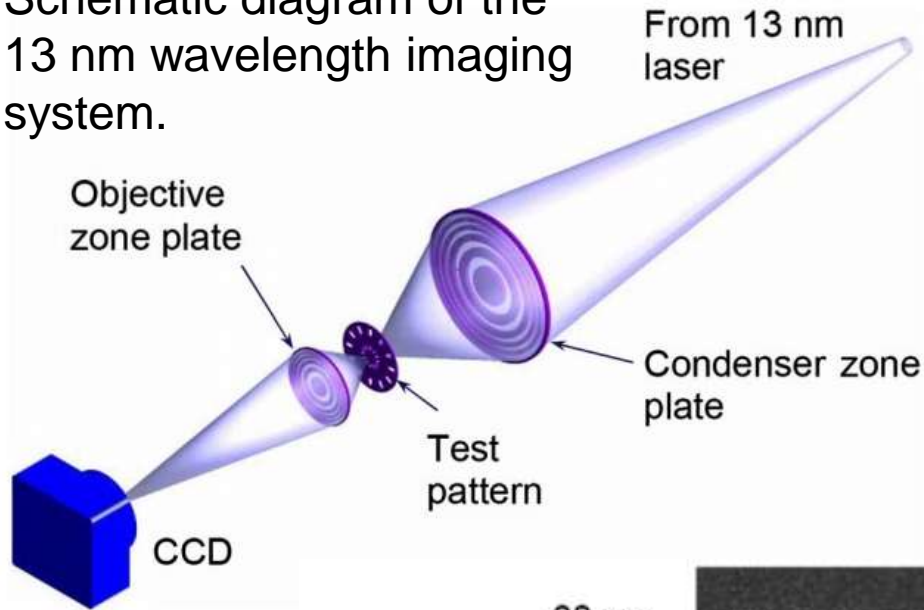
Condenser NA: H= 0.11, V=0.15, Objective NA: 0.137

Magnification: (min) 410x, Pixel size: (max) 33nm

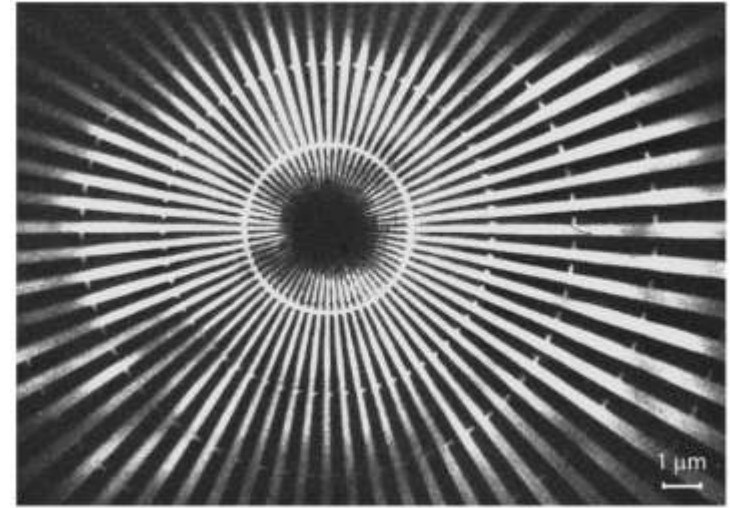
Acquisition time: 10-100sec/10Hz

Sub-38 nm resolution tabletop microscopy with 13 nm wavelength laser light

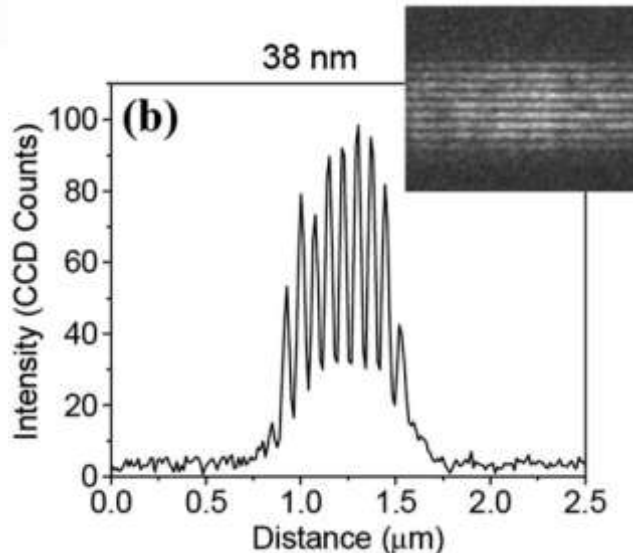
Schematic diagram of the 13 nm wavelength imaging system.



Ni-like Ag, pumped by 1J/8ps Ti:Sa laser
 $\lambda=13.9\text{nm}$, $E=1\text{-}2\mu\text{J/pulse}$



Ni-like Cd
 $\lambda=13.2\text{nm}$
 $dr=50\text{nm}$ ZP
20 sec exp.
38nm lines
resolved



Condensor: $D=5\text{mm}$, 12500 zones, $dr=100\text{nm}$, $NA=0.07$, $f=38\text{mm}$

Objective 1: $D=0.2\text{mm}$, 625 zones $dr=80\text{nm}$, $NA=0.0825$

Objective 2: $D=0.1\text{mm}$, $dr=50\text{nm}$, $NA=0.132$

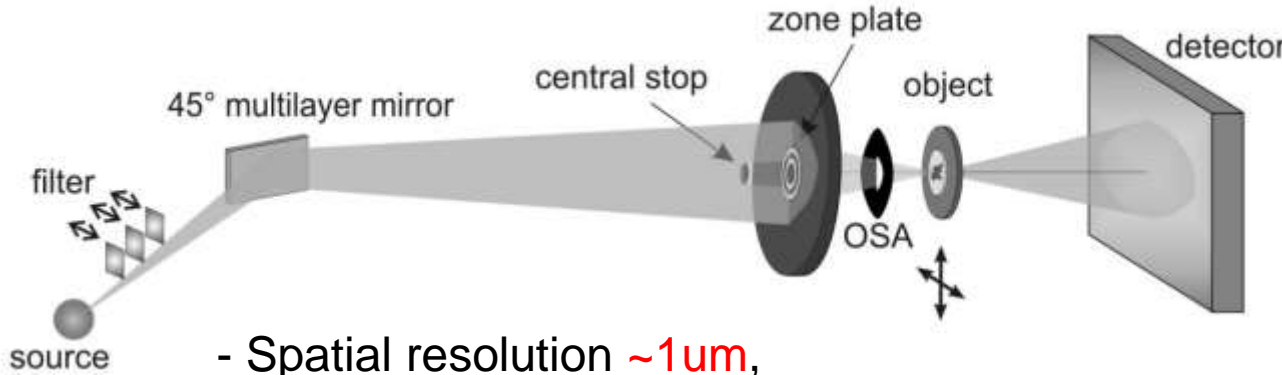
Large-field-of-view image of a set of radial spokes

- $dr=80\text{ nm}$ zone plate

- 20 s exposure.

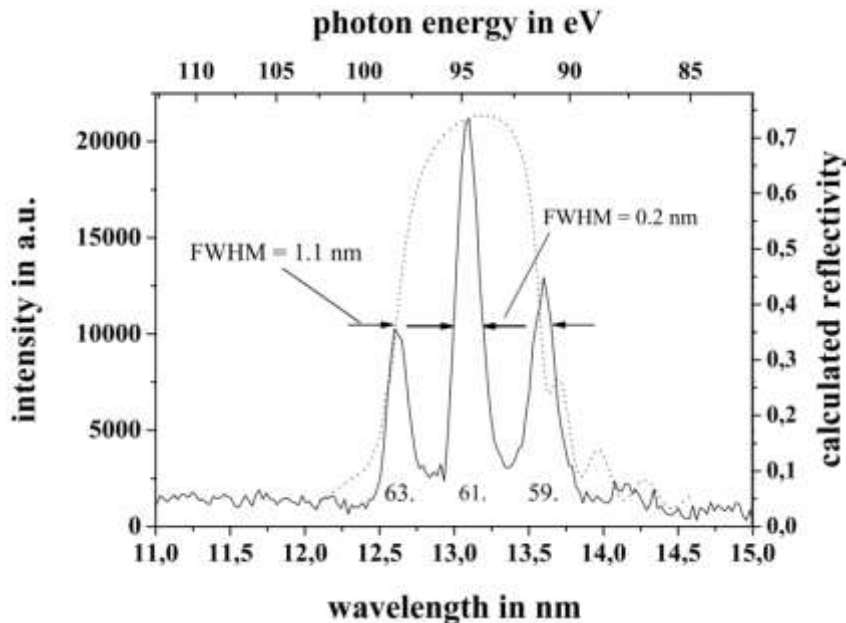
- 60 nm half-period resolution

EUV scanning transmission microscope operating with high-harmonic and laser plasma radiation



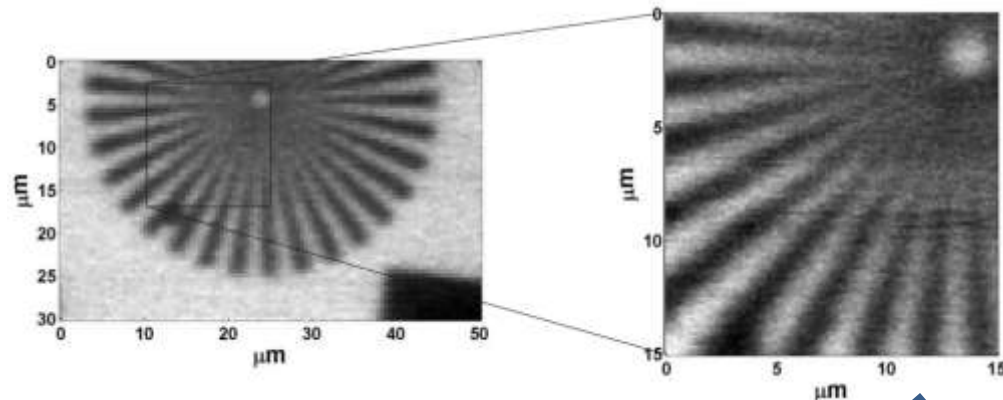
scanning transmission EUV microscope set-up

- Spatial resolution $\sim 1\mu\text{m}$,
- Ti:Sa pumping laser, 1kHz, 800nm/30fs pulses, 1mJ focused onto a SS tube,



Spectrum extracted from the HH source using a Mo/Si multilayer mirror

- Zr filters and Mo/Si mirror $\lambda/\Delta\lambda \sim 13$



Magnified images of a scanned Siemens star

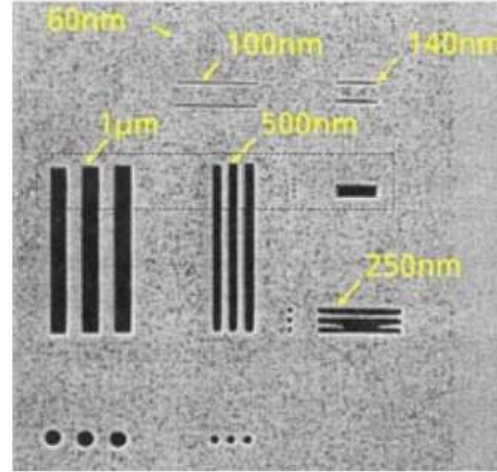
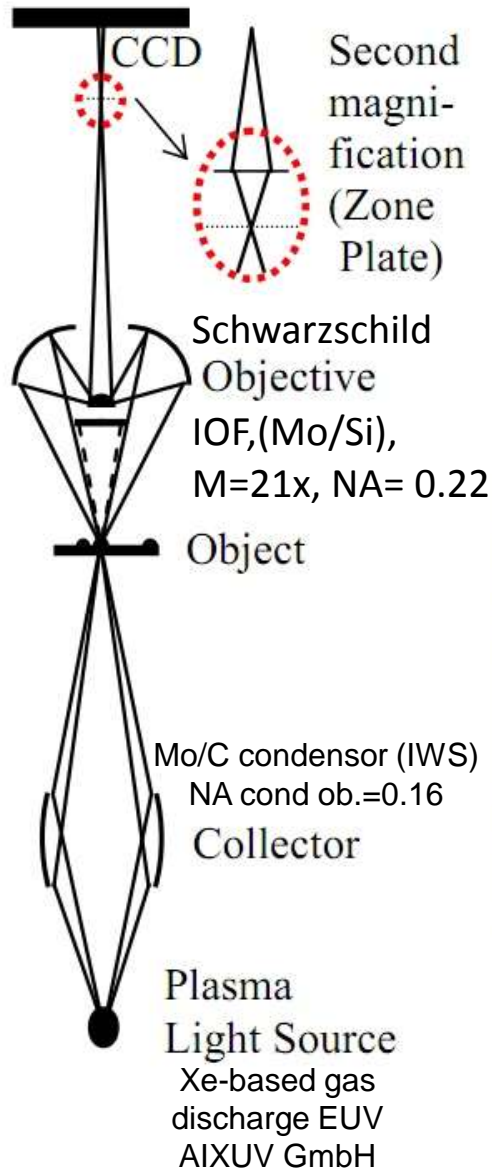
100 × 100 pixel image a scan step size of 150 nm



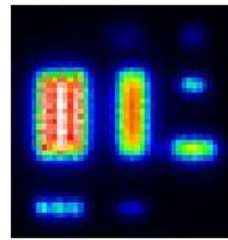
EUV microscopy for defect inspection by dark-field mapping and zone plate zooming



EUV microscope scheme

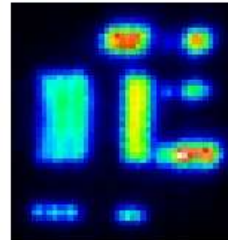


100nm holes,
Dist. 100nm

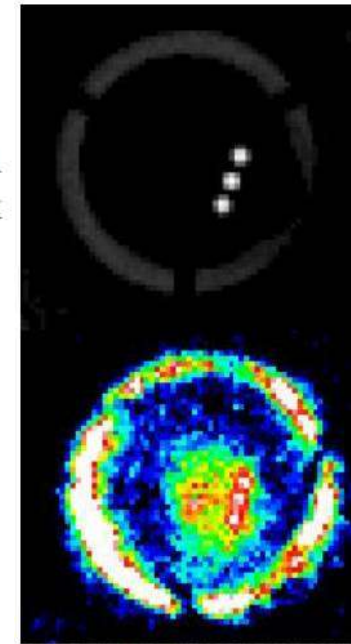


Test transmission mask

Bright field
mag=21



Dark field
mag=21



NASSO,det = 0.01, Mag = 10
 NA ZP=0.011 (30 zones), res ~1µm
 Combined resolution ~100nm

Compact water-window transmission X-ray microscopy

- 100Hz/3ns/100mJ pulses, Nd:YAG (Coherent Infinity)
- ethanol droplet target, W/B₄C condensor mirror, $\lambda=3.37\text{nm}$,
- ZP: D=56 μm , 468 zones, dr=30nm, f=498 μm .
- magnification: 650x-1000x

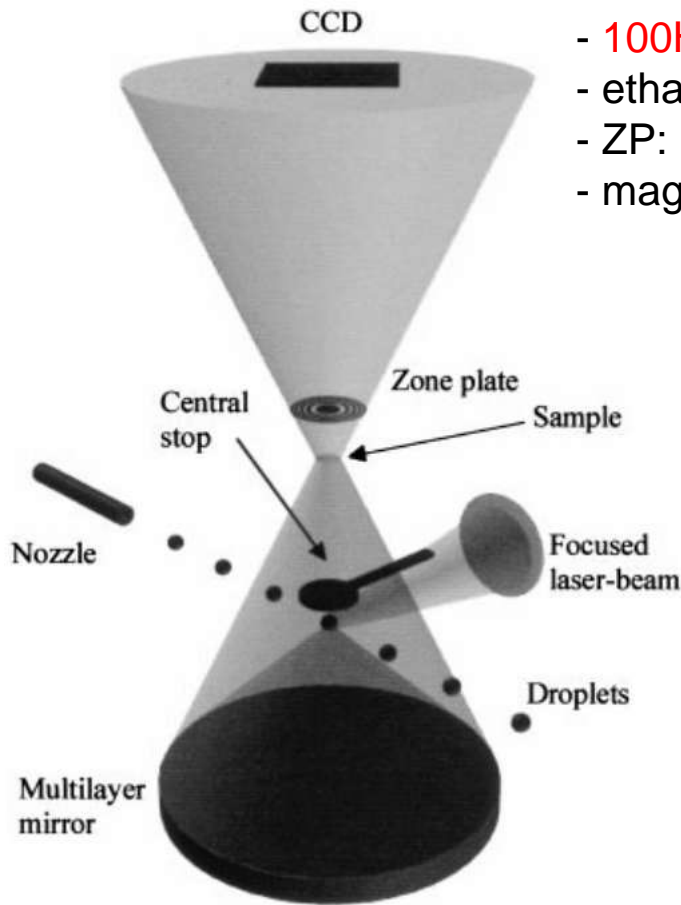


Table-top water-window X-ray microscopy arrangement

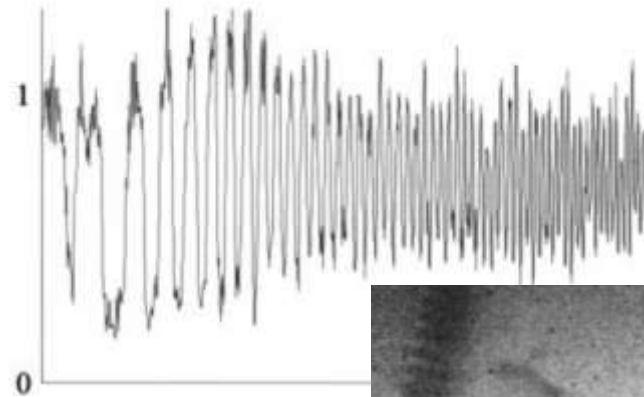
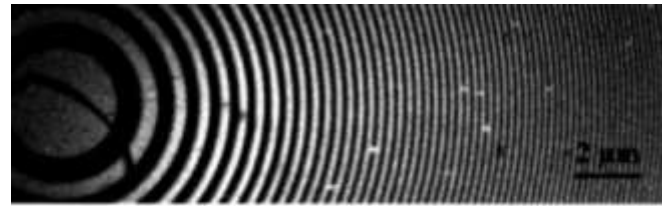
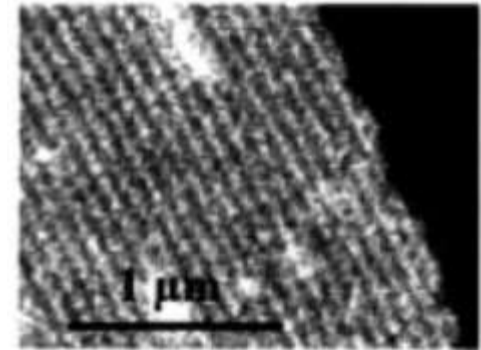
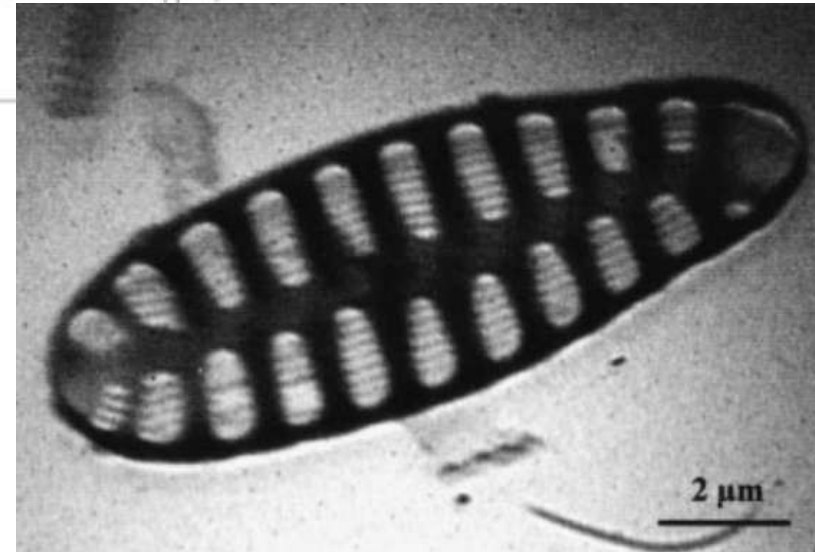


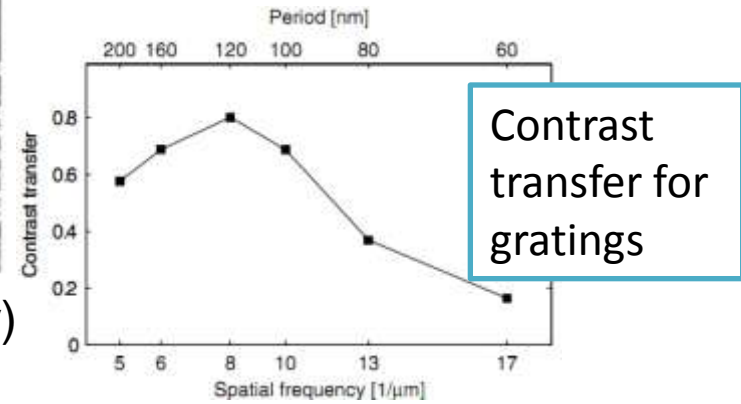
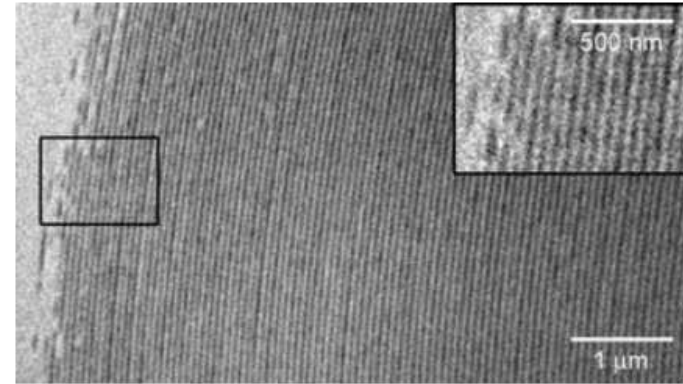
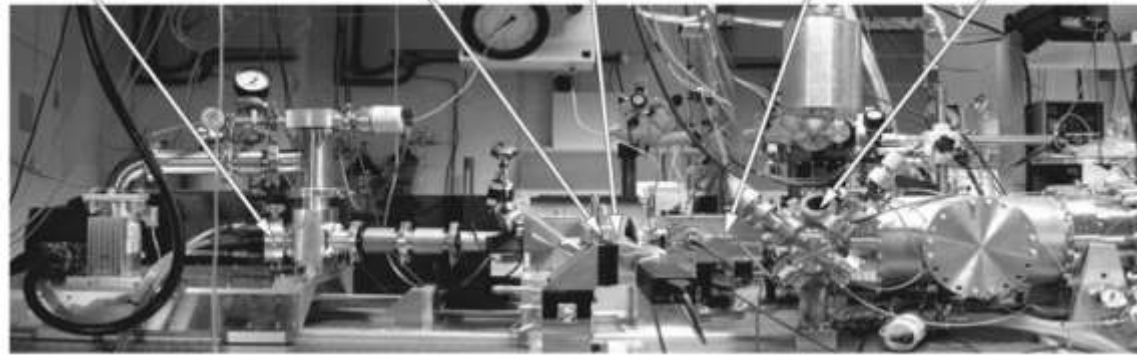
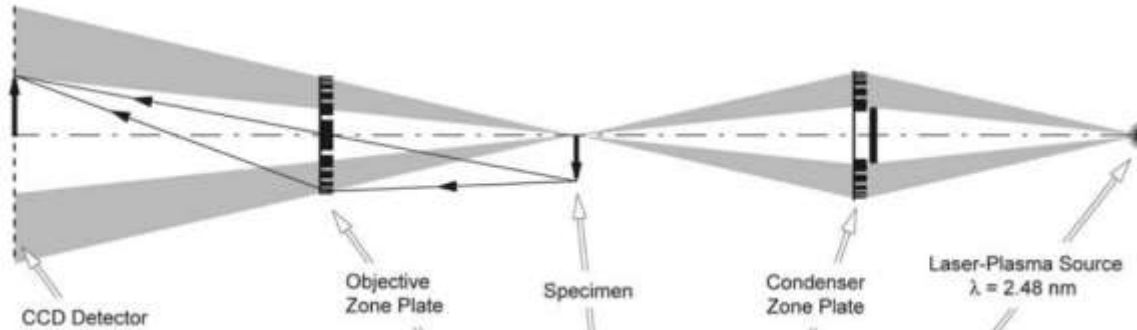
Image of diatom with M=650x



1000x magnification
60nm features visible



High-resolution compact X-ray microscopy



- **100Hz/3ns/100mJ** pulses, Nd:YAG (Coherent Infinity)
- 100-Hz regenerative liquid-nitrogen-jet laser-plasma source , $\lambda=2.48+2.49\text{nm}$
- **sub-60-nm** resolution
- exposure: 5 min

Image of a diatom

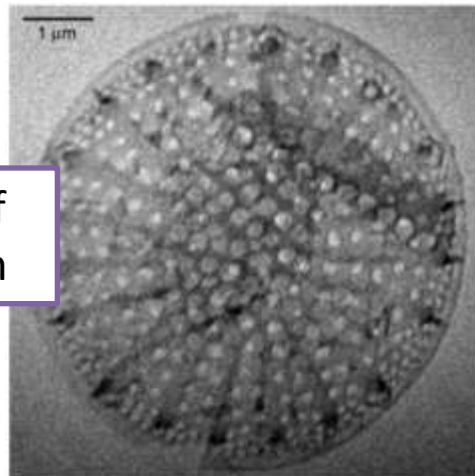
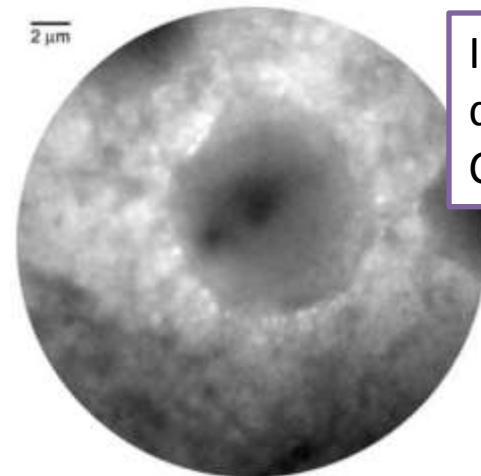
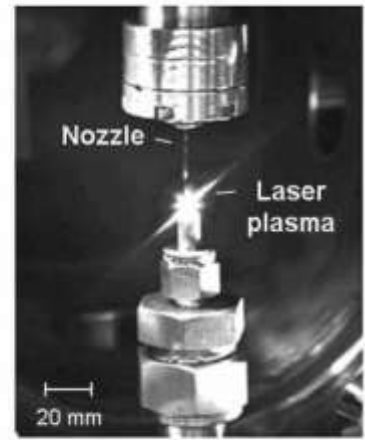
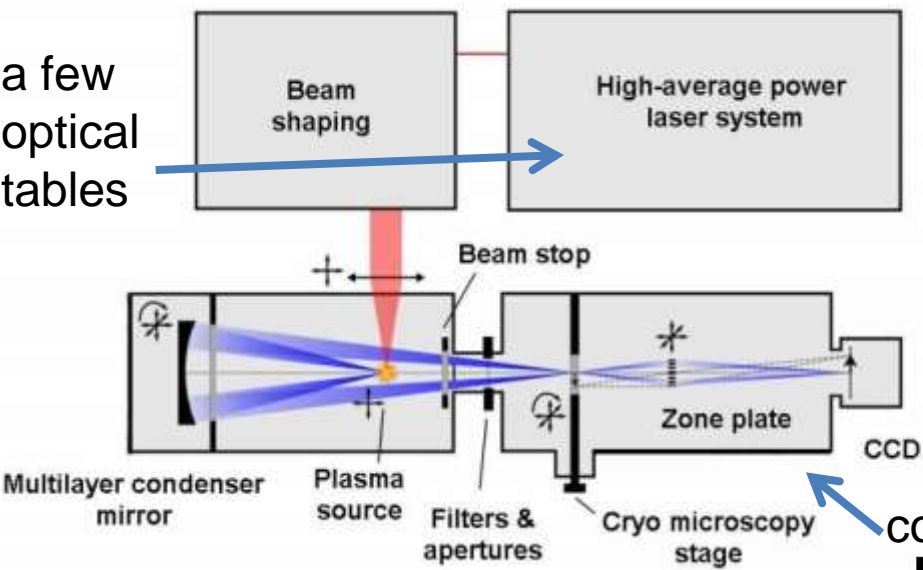


Image of a dehydrated COS-7 cell



Compact x-ray microscope for the water window based on a high brightness laser plasma source

a few optical tables

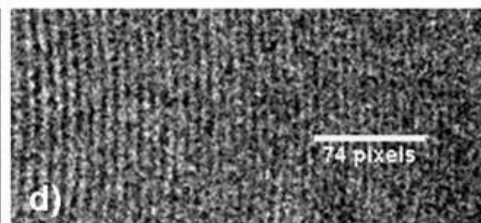
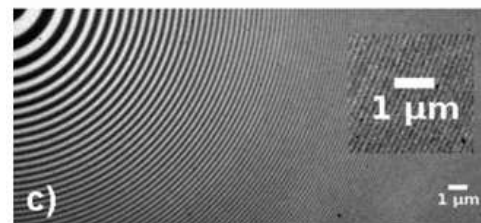
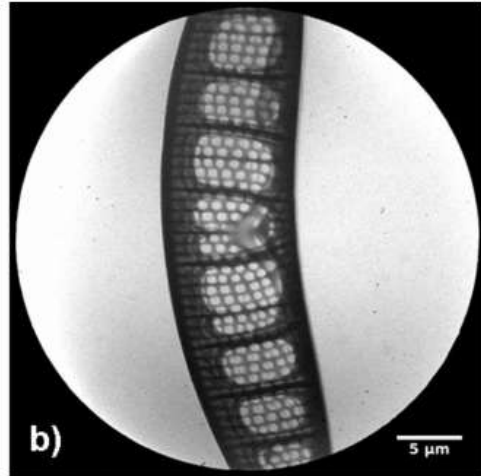
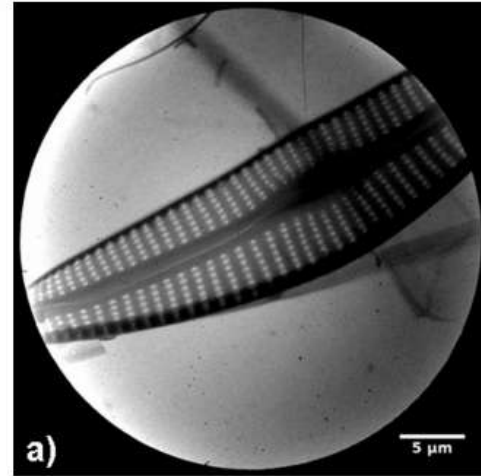
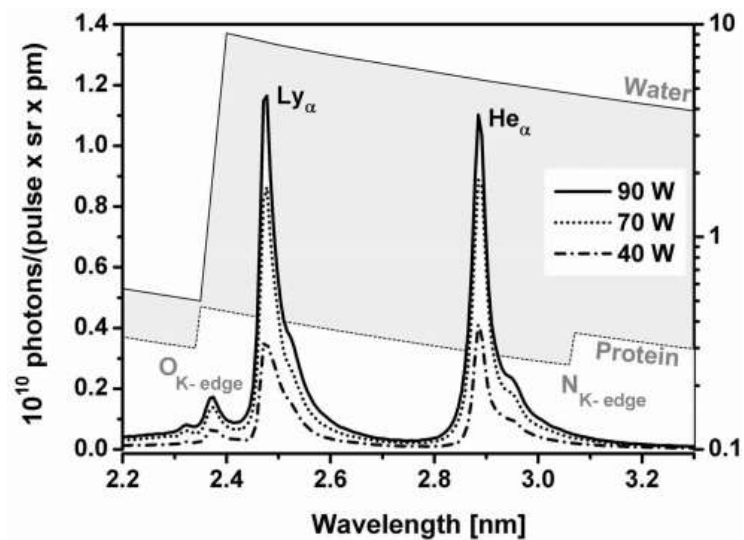


Pumping source: $\lambda=1064\text{nm}$ (Nd:YAG), $450\text{ps}/1.3\text{kHz}$
 -Target L-N₂, $\lambda=2.48\text{nm}$
 -Cr/V multilayer condenser mirror (FhG-IOF Jena), NA=0.05
 - ZP: dr=25nm, NA=0.05,
 - half-pitch resolution: **40-50nm**

compact

Images of the diatoms and ZP

The arrangement and optical layout of the table top water window x-ray microscope



Compact soft x-ray microscope using a gas-discharge light source

$\lambda=2.88\text{nm}$ (He-line N_2), 1kHz, electrical discharge

Grazing incidence condenser

ZP: 468 zones $\text{dr}=30\text{nm}$, $D=56\mu\text{m}$, $f=0.585\text{mm}$,

$\text{NA}=0.048$

spatial resolution: **40nm**

exposure: 20sec

photons: **$2.9\text{E}6/\mu\text{m}^2/\text{s}$, aim $1\text{E}7$**

Plasma source

Central stop

Ellipsoidal condenser

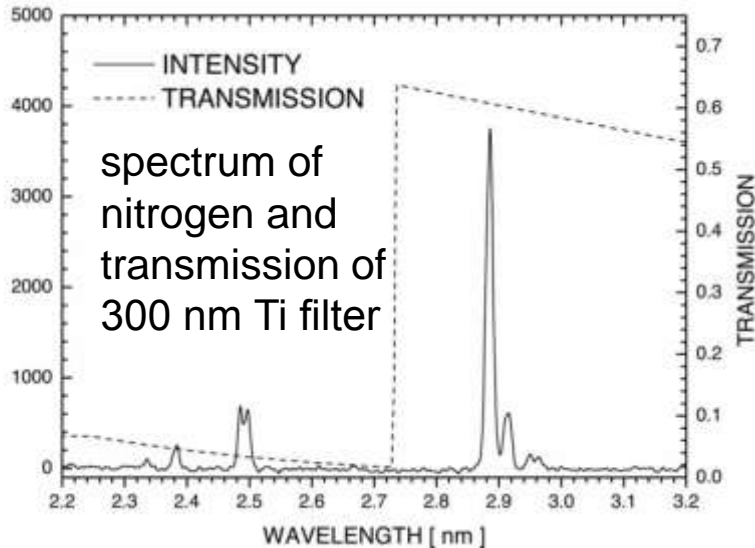
Titanium filter

Zone plate

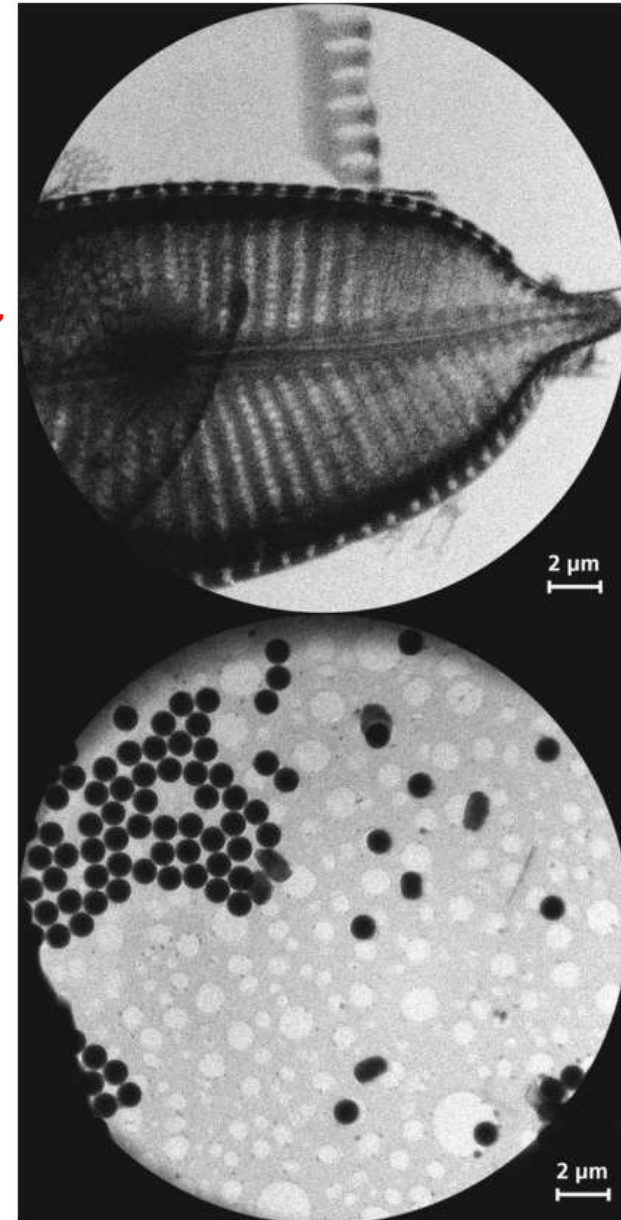
Specimen

CCD detector

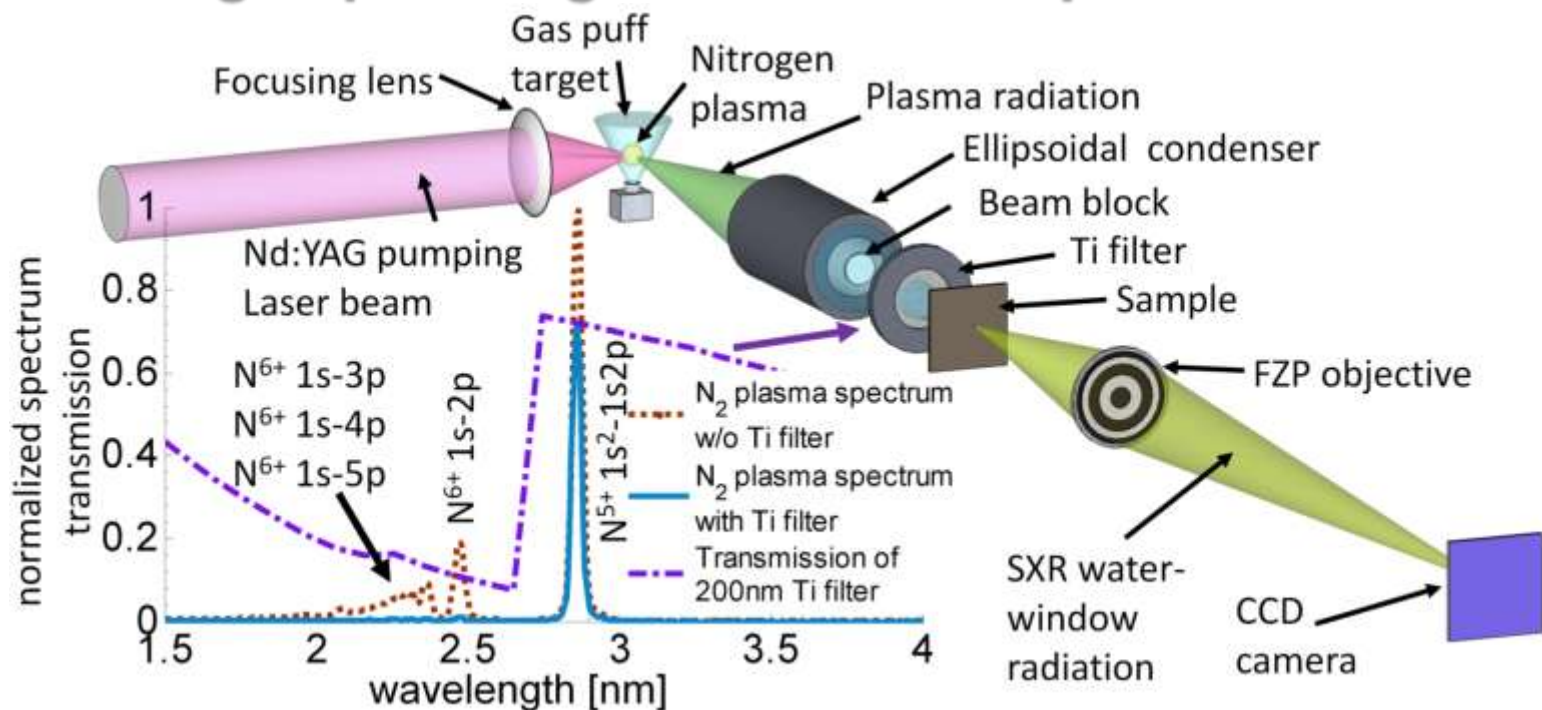
Schematic of the experimental microscope setup



Microscopic images of diatoms and latex spheres



Compact SXR transmission microscope using double stream gas puff target with 60 nm spatial resolution



Scheme of the setup and spectrum of the nitrogen plasma source

Double stream N_2/He gas puff target,

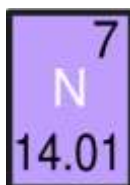
$N_2=8\text{bars}$, $He=6\text{bars}$

$TD1=400\mu\text{s}$,

$PW1=350\mu\text{s}$,

$TD2=500\mu\text{s}$,

$PW2=250\mu\text{s}$,



Number of photons: $\sim(7.9\pm 0.2)\times 10^9$ /pulse @ $\lambda=2.88\text{nm}$

Energy: $(561\pm 17)\text{nJ/pulse}$ and at 10-Hz repetition rate

Inverse relative bandwidth (FWHM): $\lambda/\Delta\lambda\sim 70$, (spec. limit)

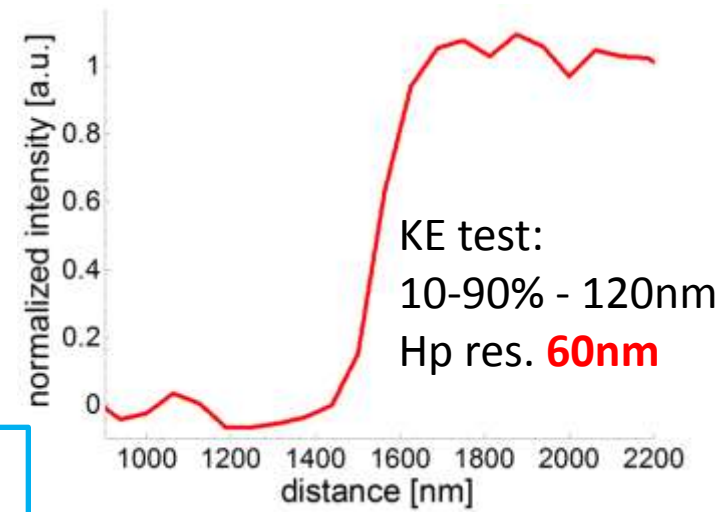
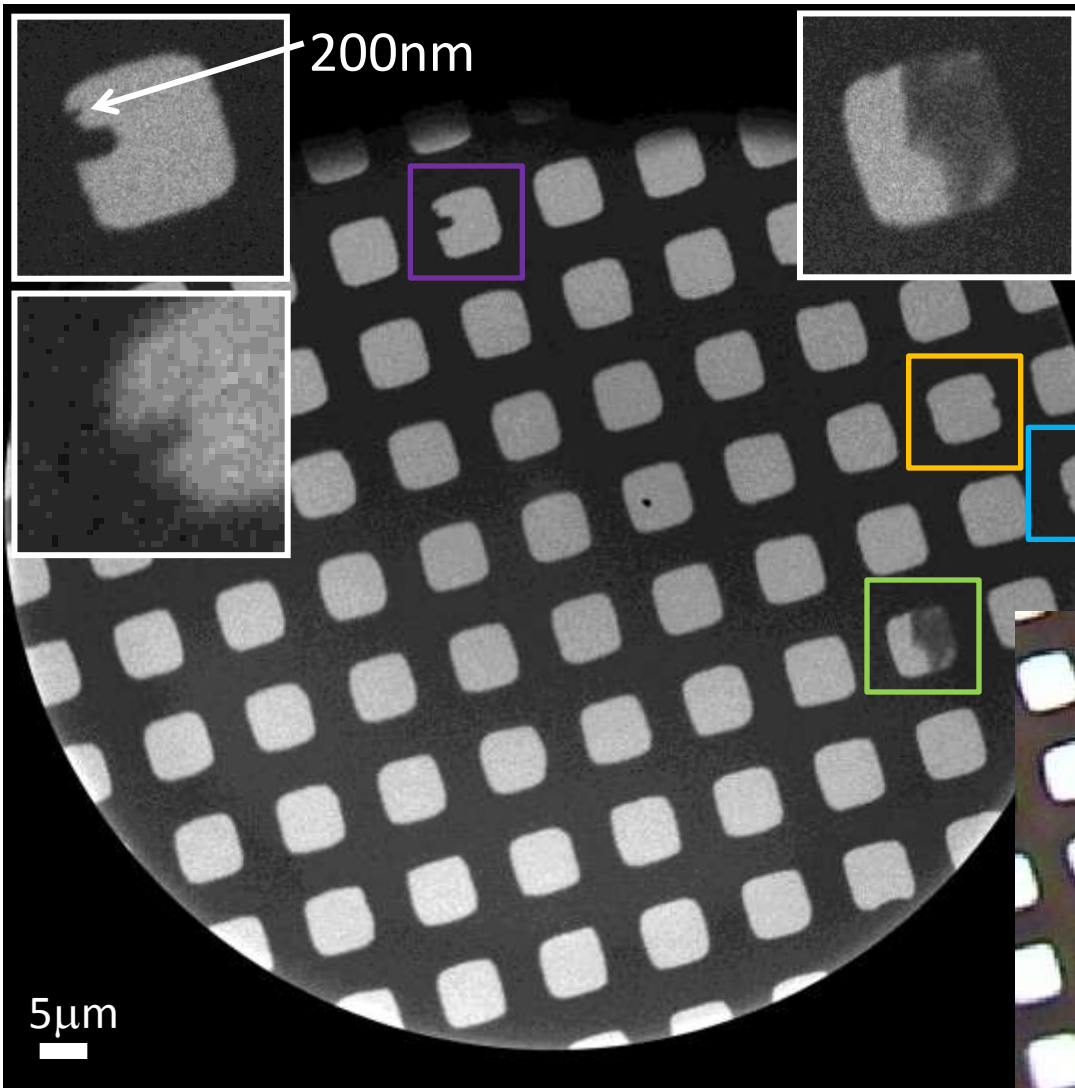
Plasma size: $\sim 310\times 470\mu\text{m}^2$

FZP objective:

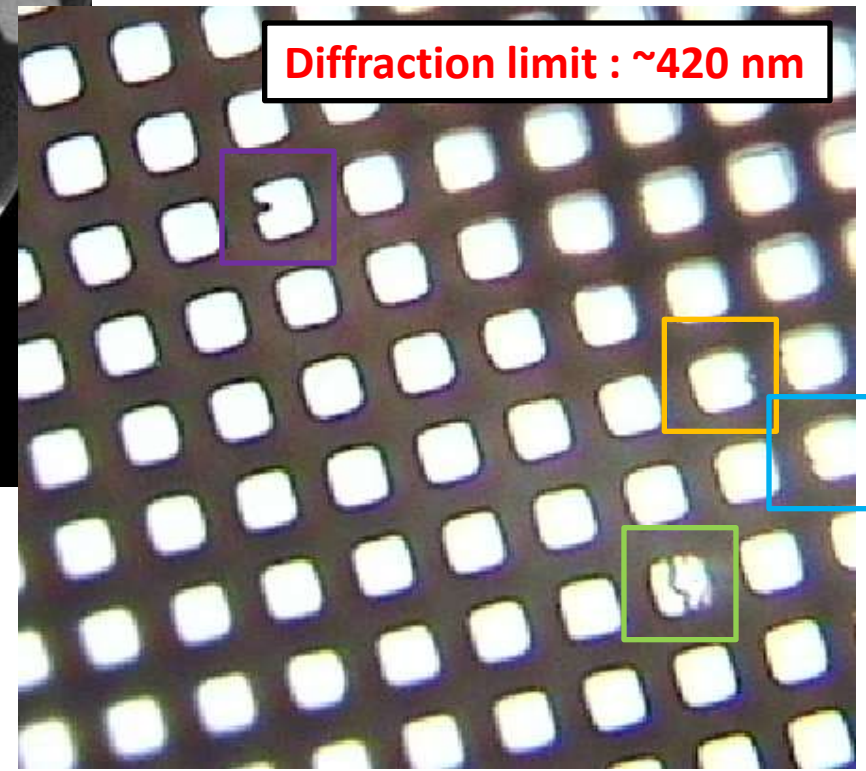
- Si_3N_4 zoneplate 400nm thick

- $D=250\mu\text{m}$, $\Delta r=30\text{nm}$,

- $f=2.6\text{mm}$, $\lambda=2.88\text{nm}$, $NA=0.048$



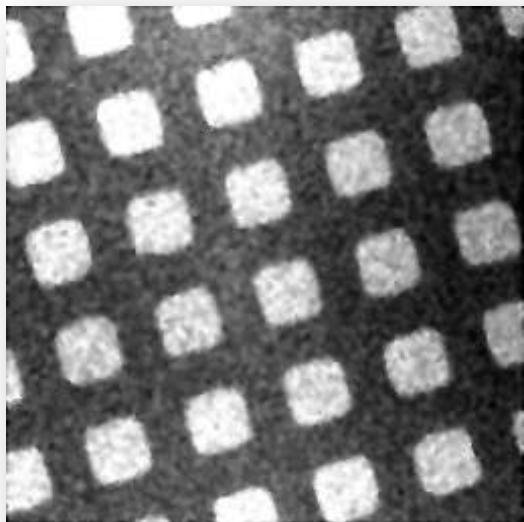
Optical image: 40x objective, NA=0.7



Cu mesh 2000 periods per inch

Exposure: 100 SXR pulses, @10Hz rep. rate, exposure time 12 seconds, T=-20°C, FOV=120µm, res. 60nm, mag. 220x, Great eyes camera 2048x2048 pix, GE2048

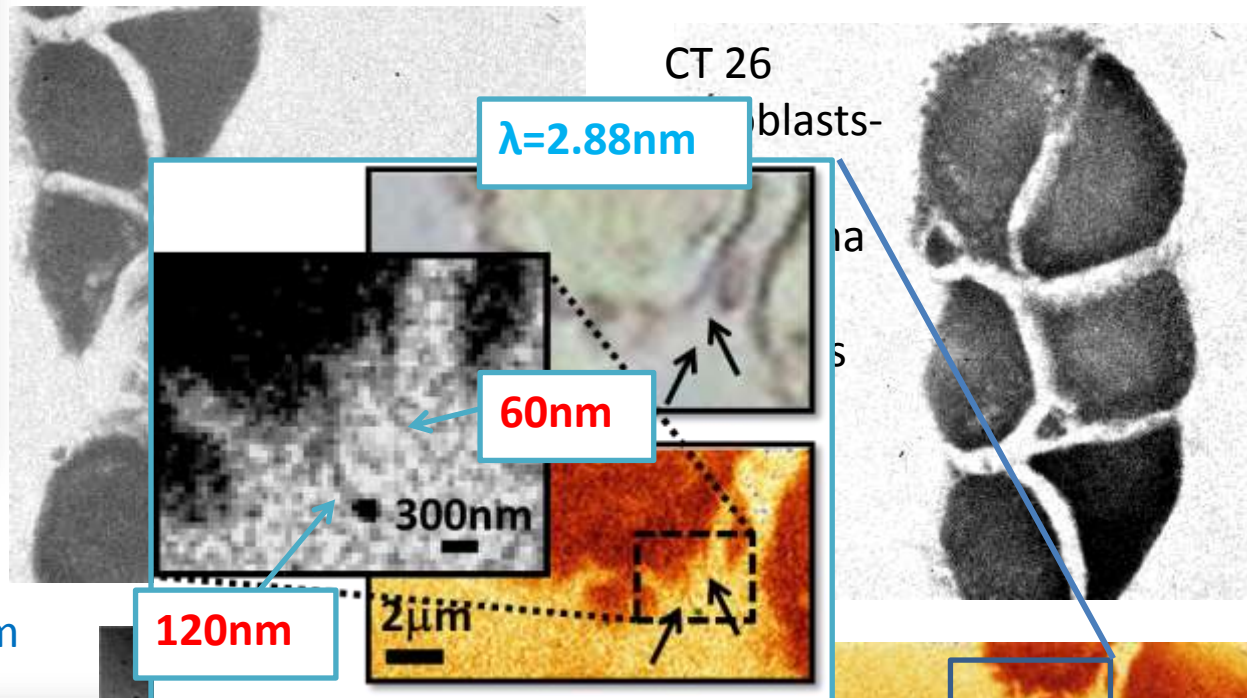
Single shot operation and biological samples



Exposure 1 SXR pulse – 3ns
4x4 CCD binning
spatial resolution ~250-300nm

Supercoiled pBR322
DNA (4361bp)-
circular double-
stranded DNA from
Inspiralis, UK, on top
of 50nm Si₃N₄
membrane,
100ng/μl

sample
thickness
~160nm



120nm

$\lambda=2.88\text{nm}$

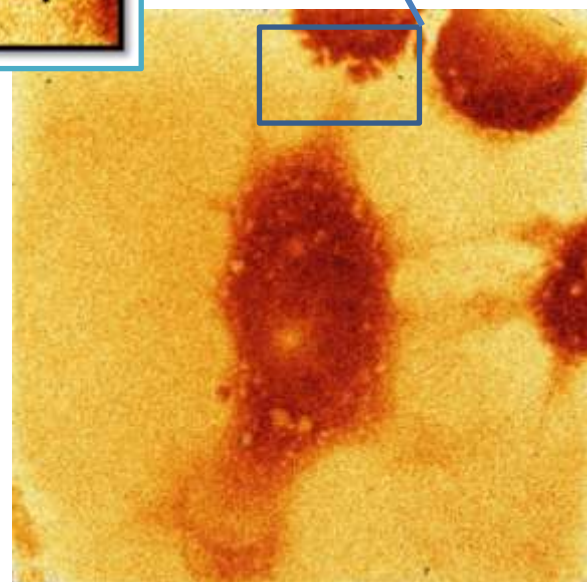
60nm

300nm

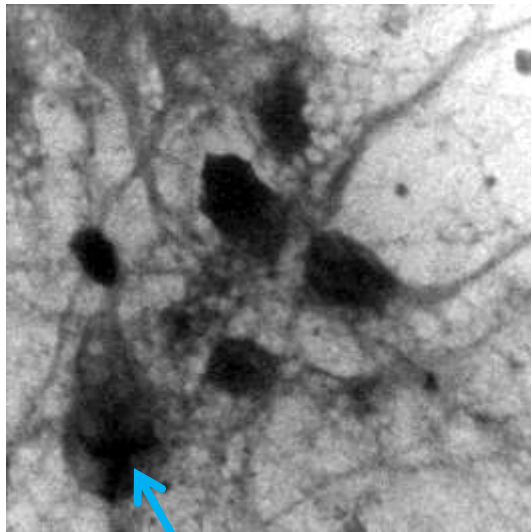
2μm

10μm

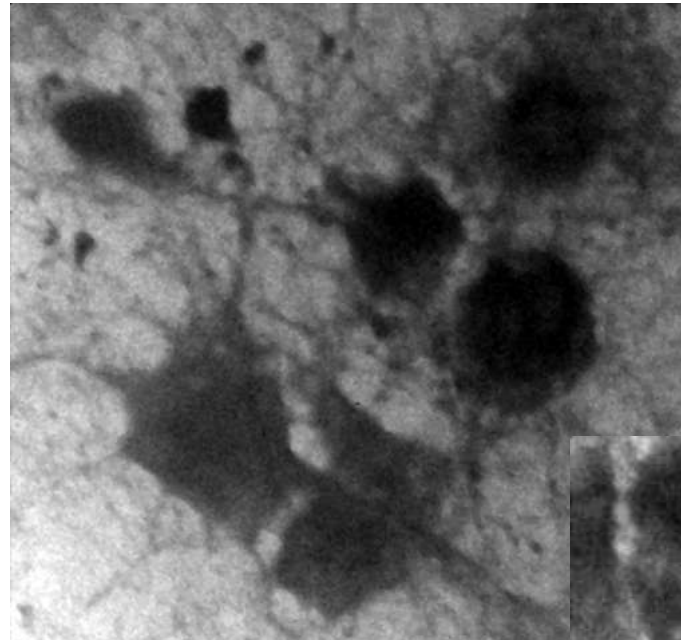
supercoiled
plasmid DNA



Hippocampal neurons from E17 mouse embryos

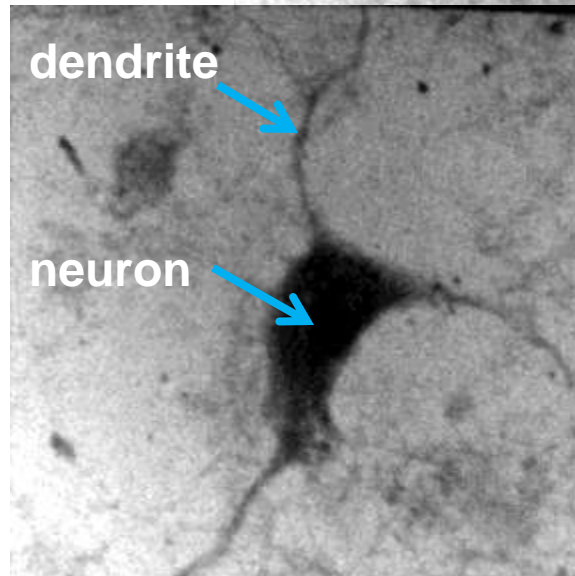
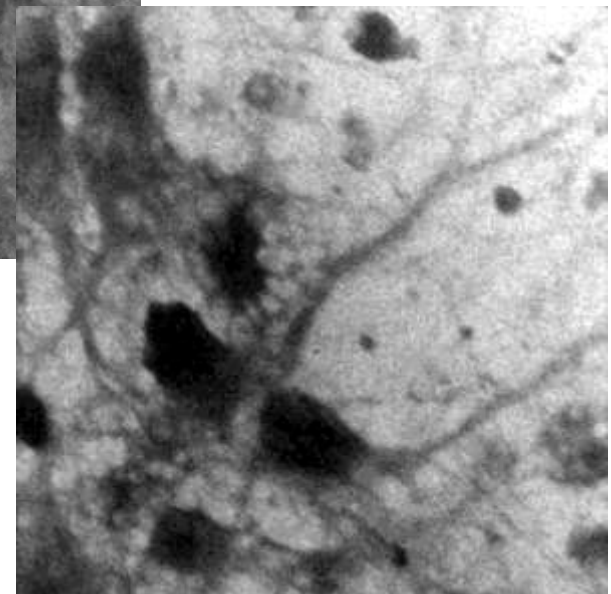


Internal structure visible



Multiple
interconnected
neurons

on 50 nm Si_3N_4 membrane,
200 SXR pulses
22 sec, 10 Hz

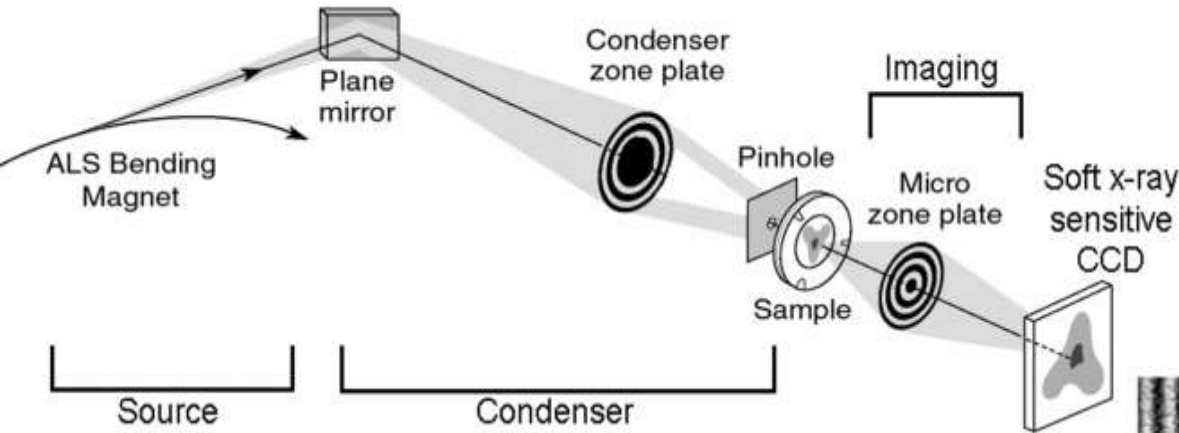


Individual
neurons with
dendrites

*Samples courtesy of M.
Odstrčil, Paul Scherrer
Institut (Switzerland)*



20-nm-resolution SXR microscopy demonstrated by use of multilayer test structures

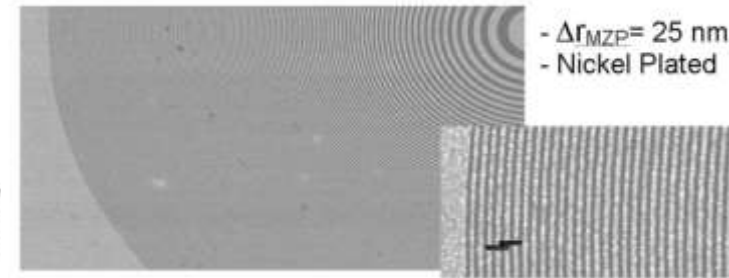


Schematic diagram of the soft x-ray, full-field imaging microscope, XM-1, at the Advanced Light Source (ALS)

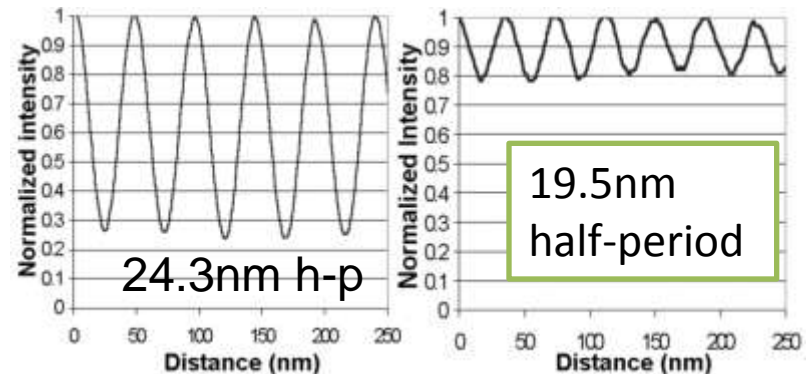
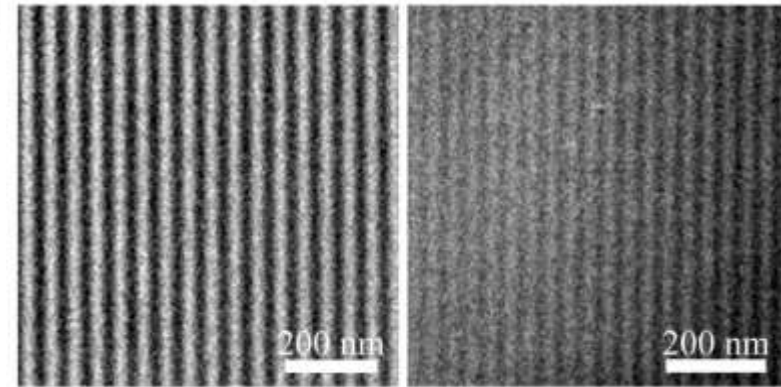
$\lambda=2.07\text{nm}$ (600eV), spatial resolution 20nm (diffraction limit 19nm)

Condensor: D=10mm, 5mm CS, 41700 zones, dr=60nm, NA=0.017, f=289mm

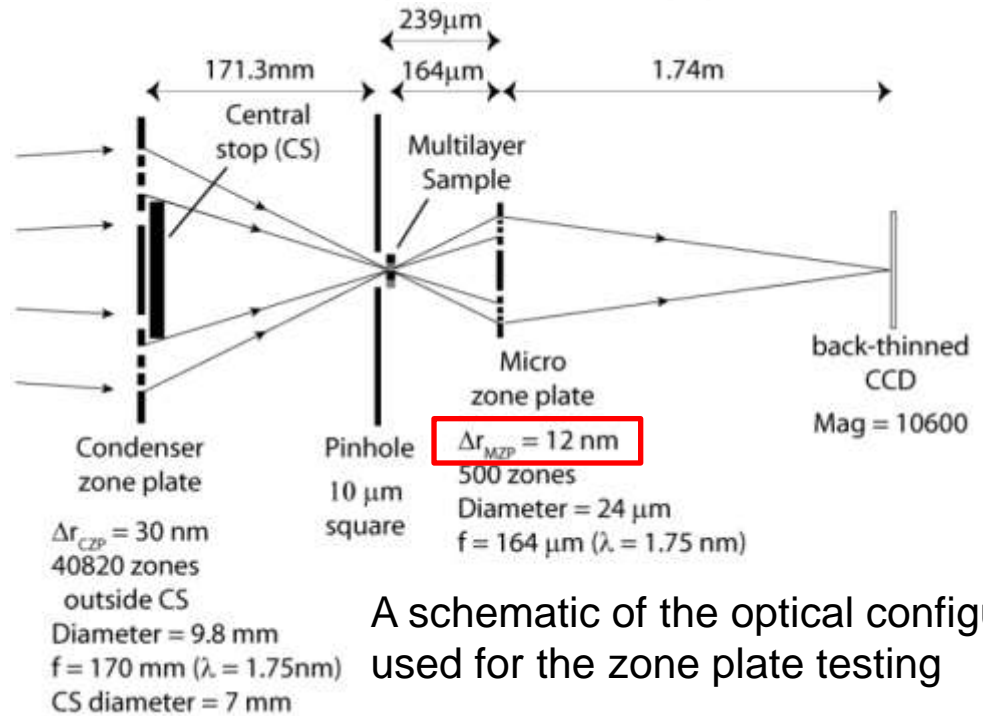
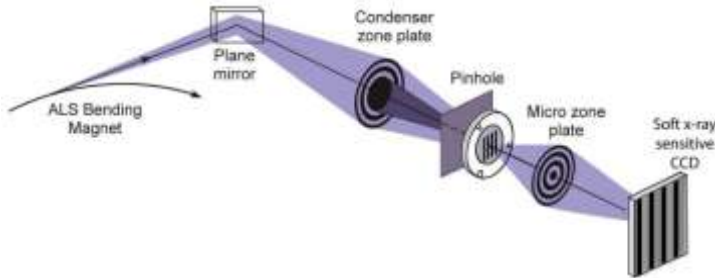
Objective: D=30 μm , 300 zones dr=25nm, NA=0.04, f=0.362mm



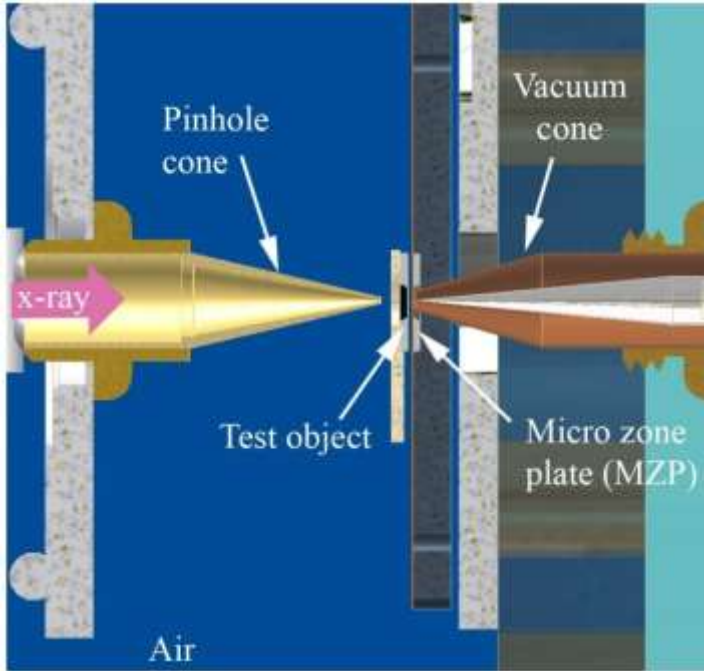
SEM micrograph of a 25-nm outermost-zone-width MZP



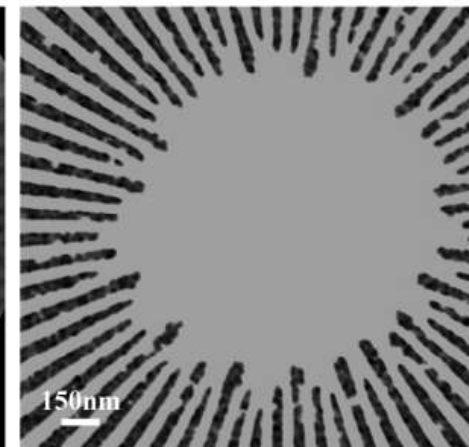
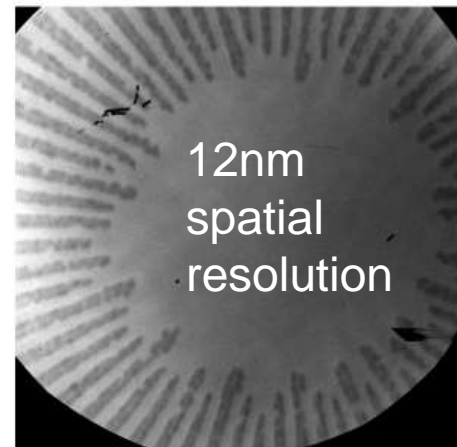
Demonstration of 12 nm Resolution Fresnel Zone Plate Lens based Soft X-ray Microscopy



A schematic of the optical configuration used for the zone plate testing



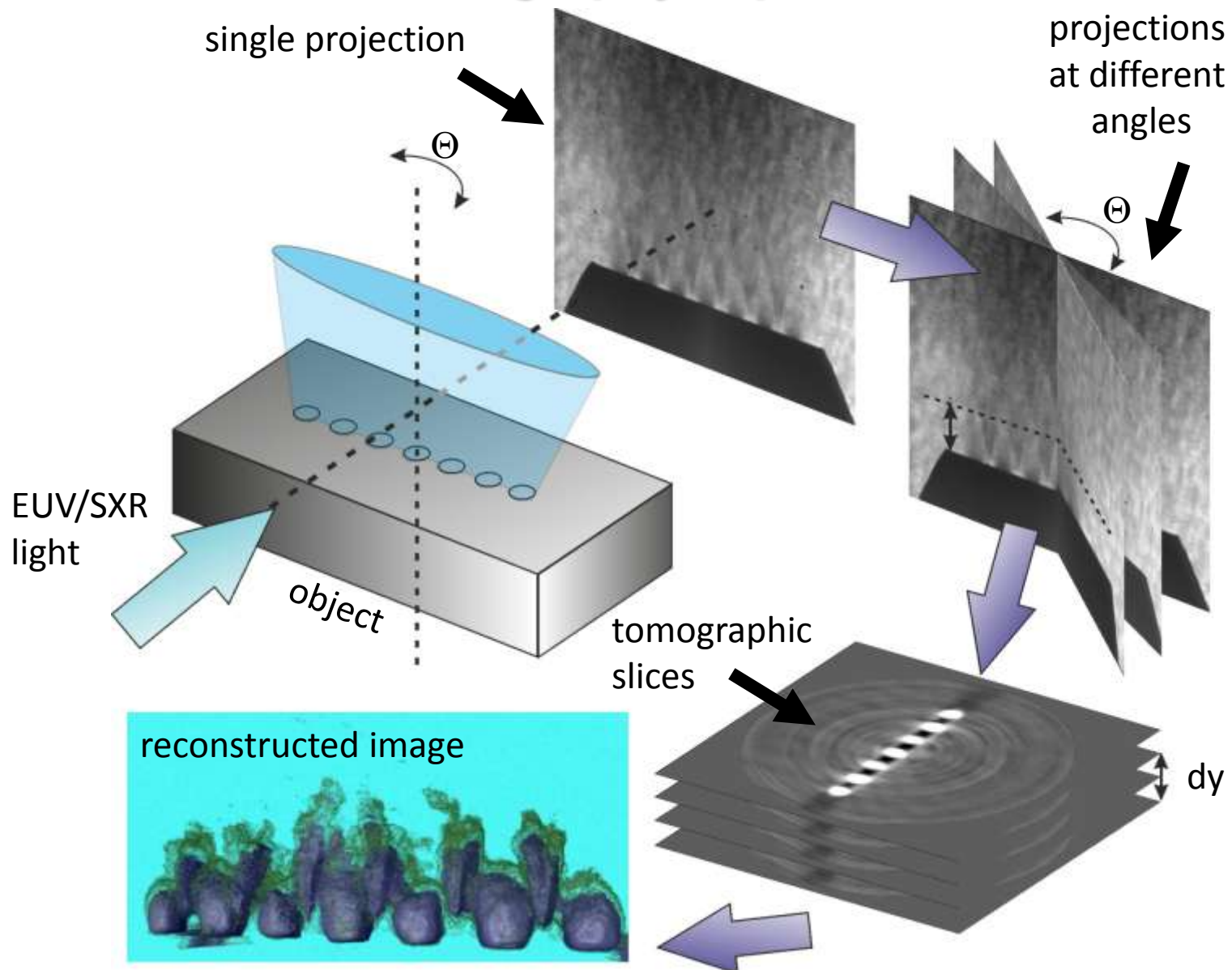
Imaging setup of the test object and the 12 nm zone plate at the XM-1 microscope



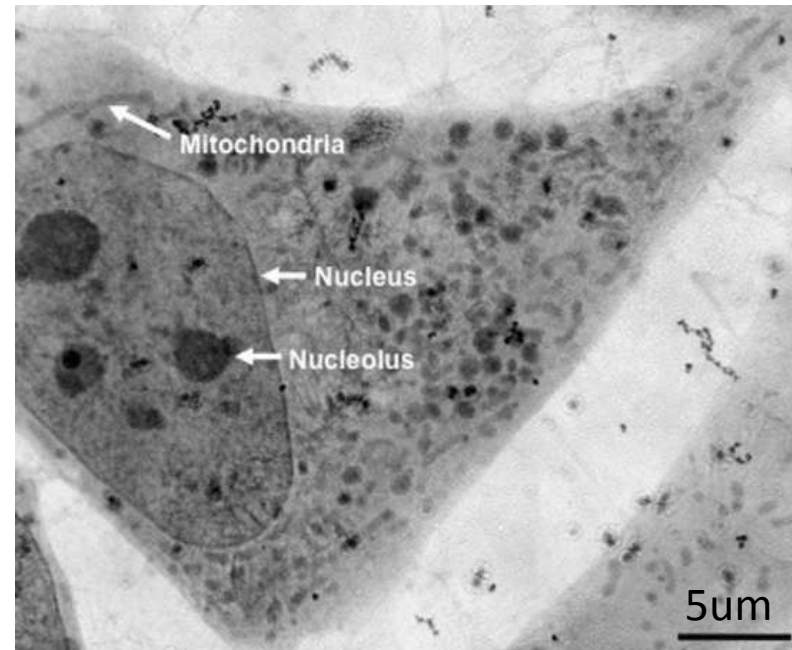
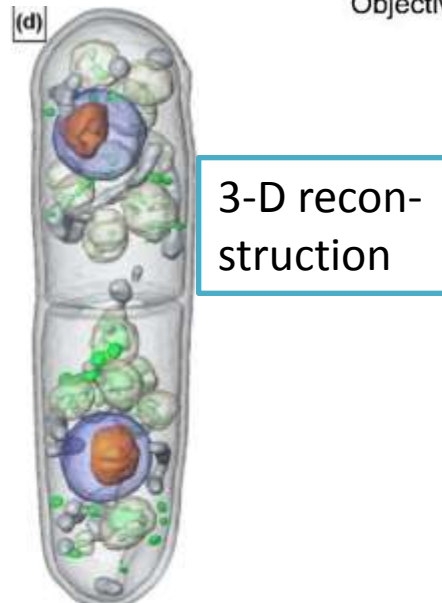
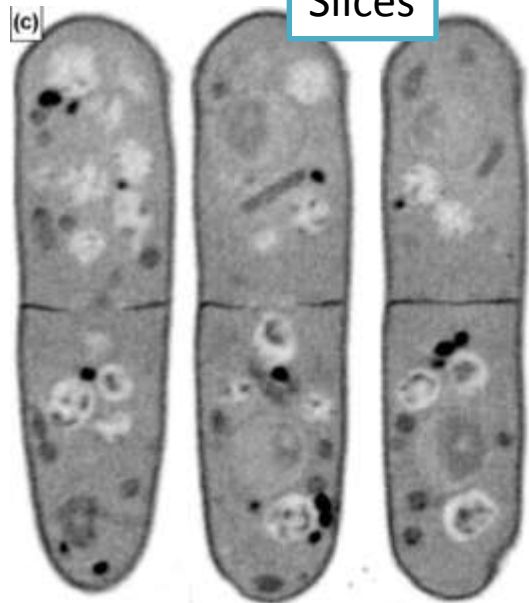
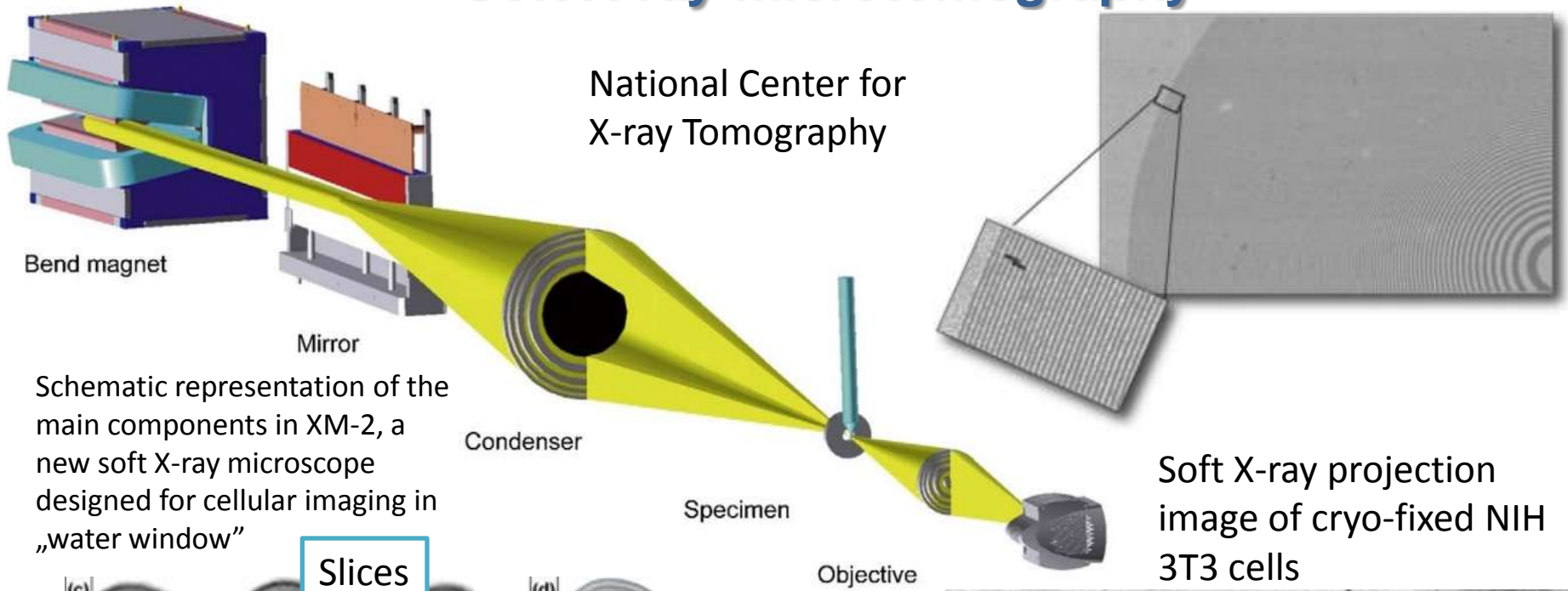
SXR image @ $\lambda=1.75\text{nm}$

TEM image

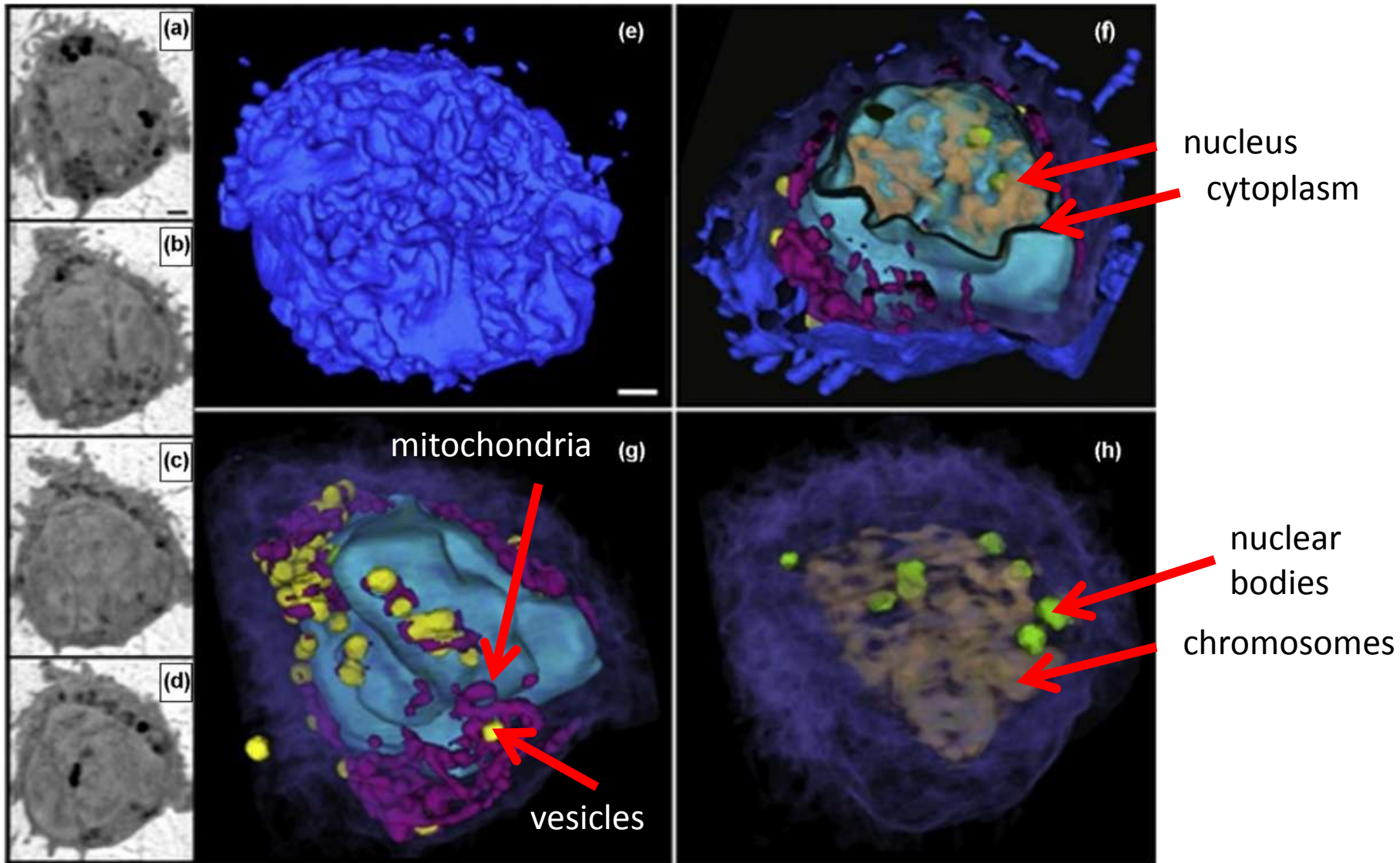
Flowchart of the EUV/SXR tomography experiment



Soft X-ray microtomography



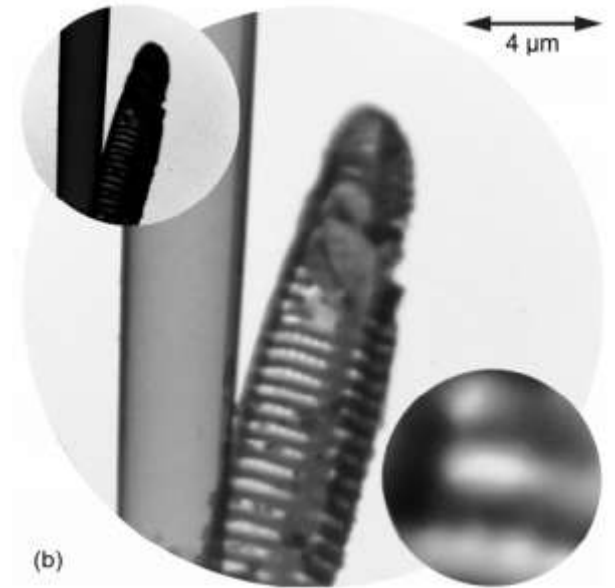
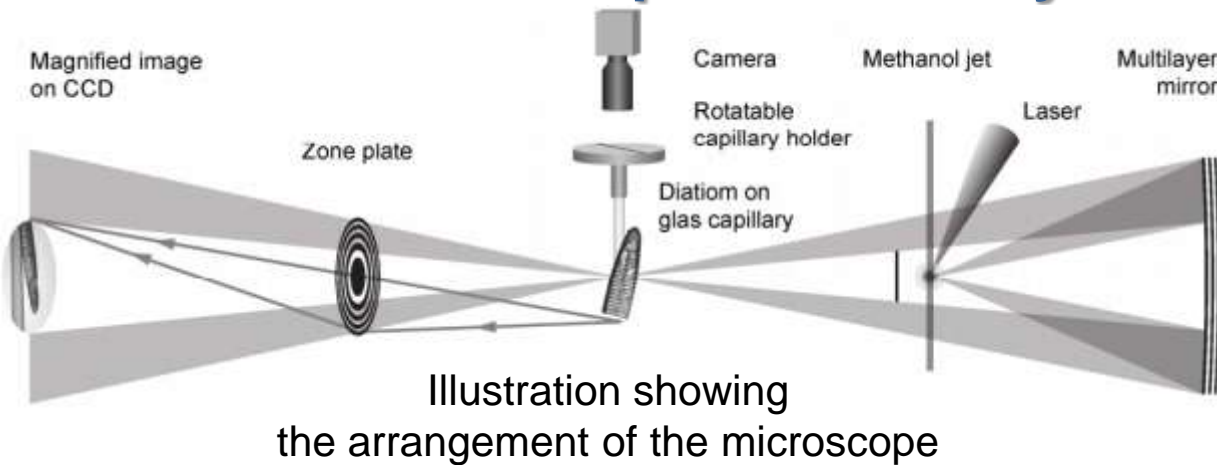
Soft X-ray microtomography



(a-d) slices from the tomographic reconstruction of a cryo-fixed T-cell

(e) cell surface with numerous filopodial extensions
(f-h) section views

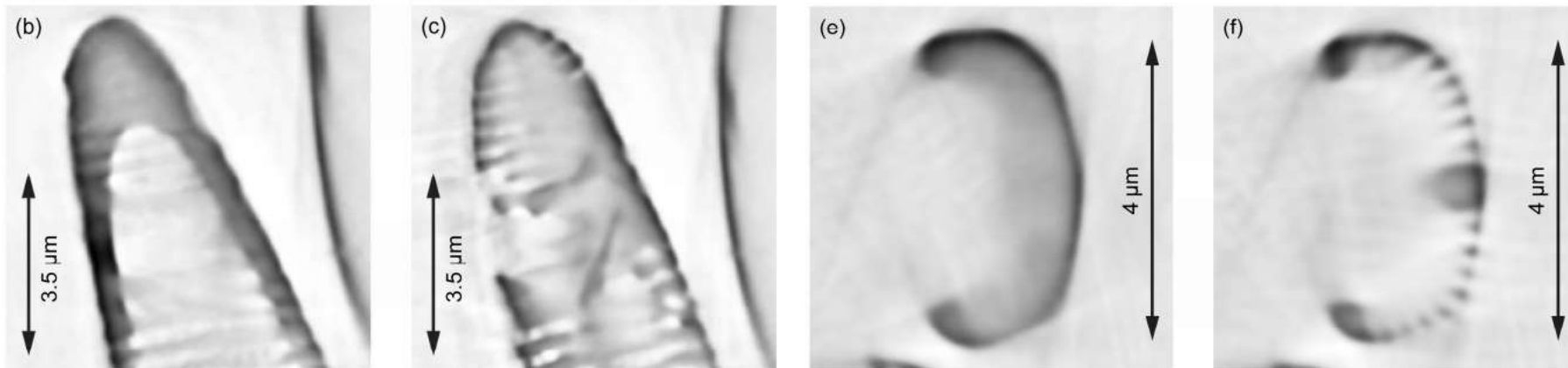
High-resolution computed tomography with a compact soft x-ray microscope



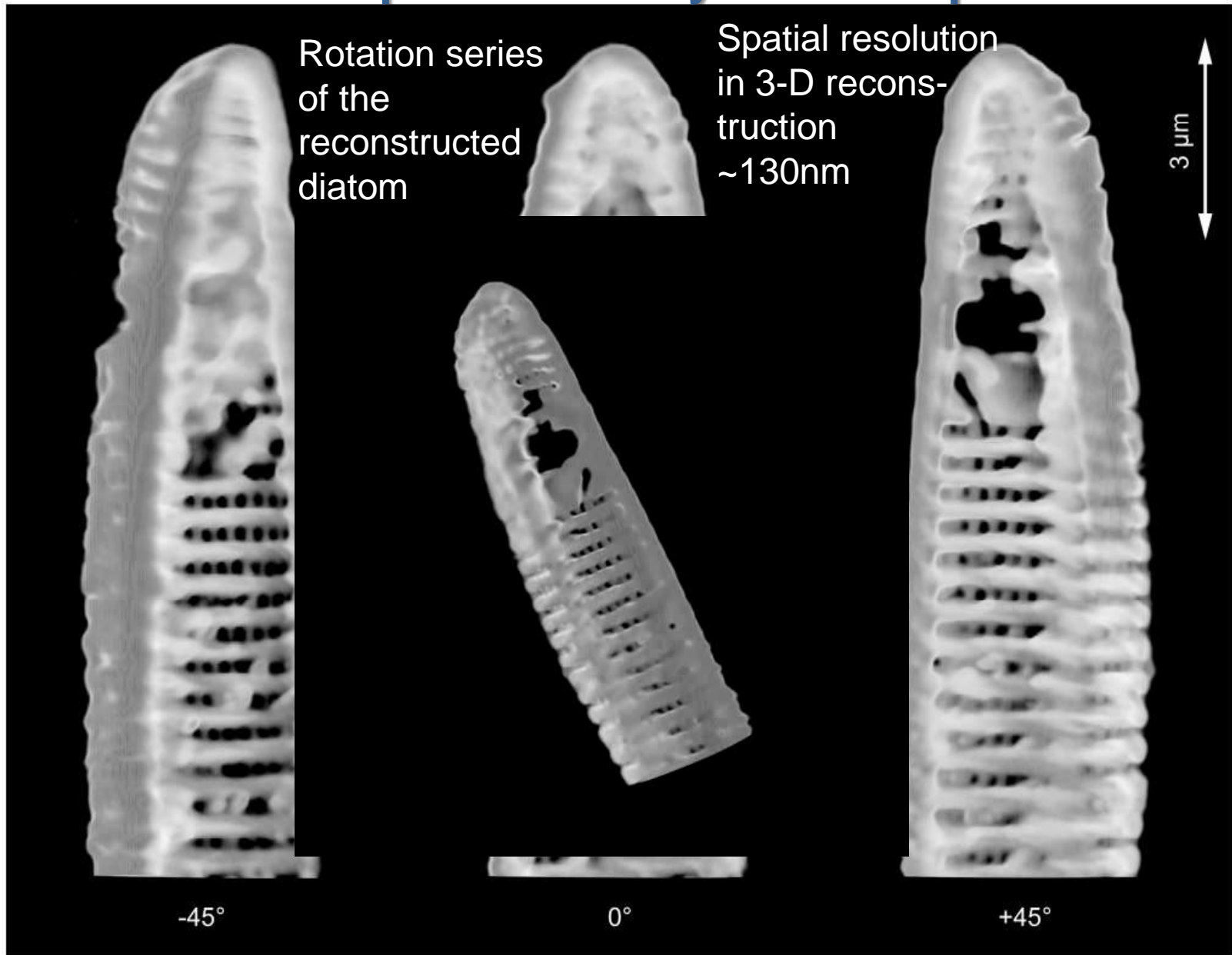
projection image of a diatom

- 100Hz/3ns/100mJ pulses, Nd:YAG 2nd harmonic 532nm
- ethanol droplet target, Cr/Sc condensor mirror, $\lambda=3.37\text{nm}$,
- 300nm Ti filter, 2E6 photons/ $\mu\text{m}^2/\text{s}$
- ZP: Ni, D=56 μm , 468 zones, dr=30nm, f=498 μm .

Selected slices through the reconstructed tomogram



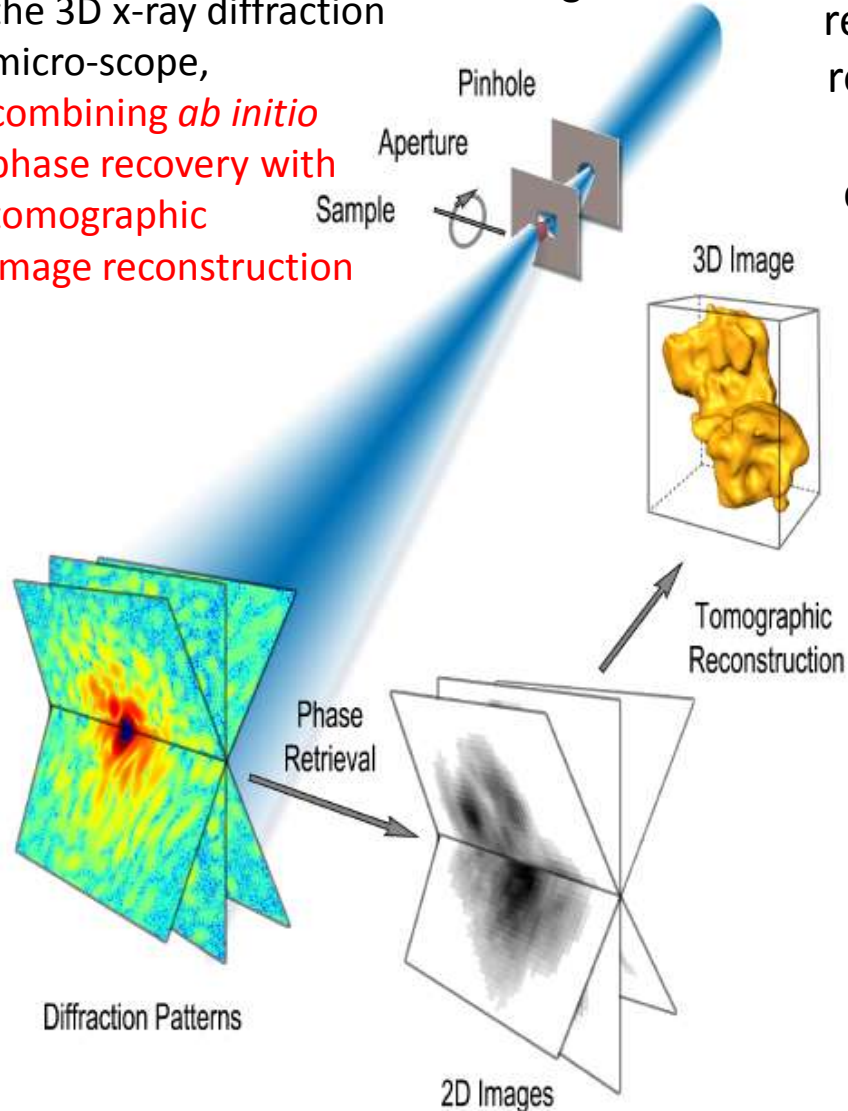
High-resolution computed tomography with a compact soft x-ray microscope



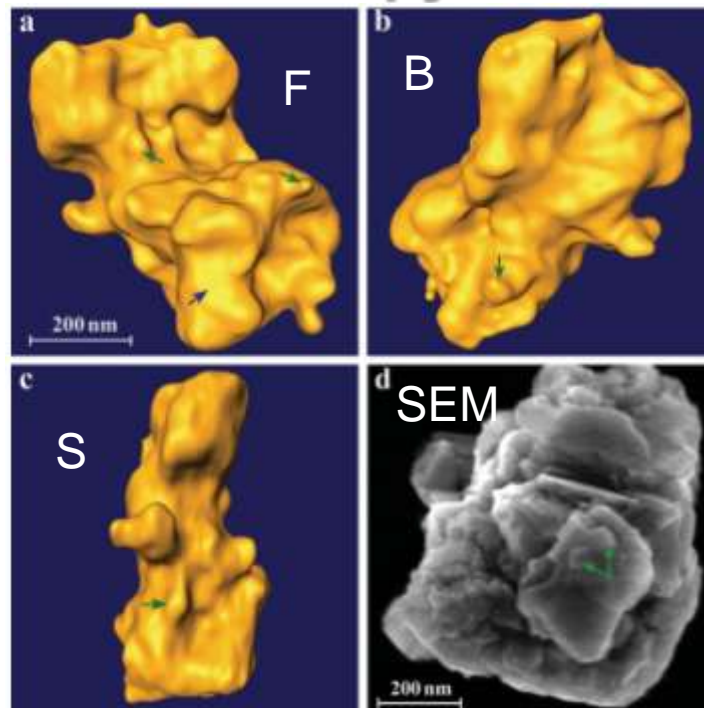
Three-Dimensional GaN-Ga₂O₃ Core Shell Structure Revealed by X-Ray Diffraction Microscopy

Schematic layout of the 3D x-ray diffraction microscope, combining *ab initio* phase recovery with tomographic image reconstruction

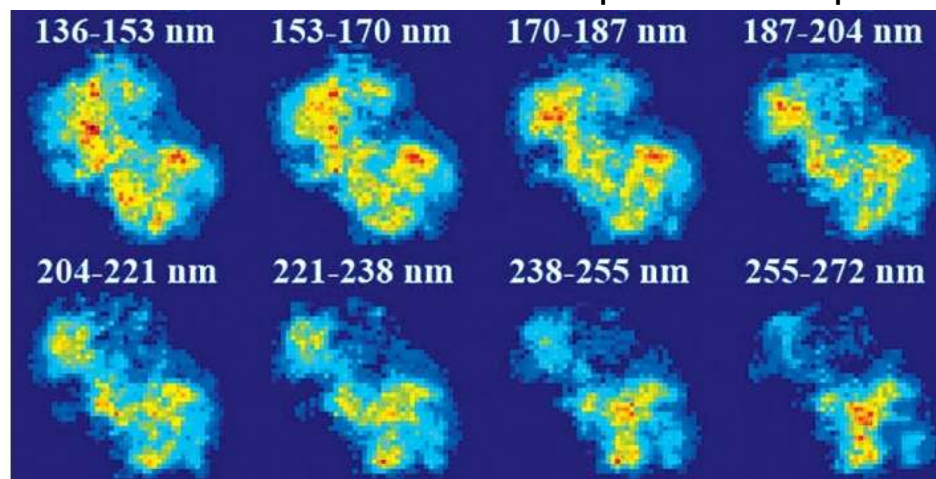
SPring-8 @ 5 keV



Isosurface rendering of a reconstructed 3D GaN quantum dot particle

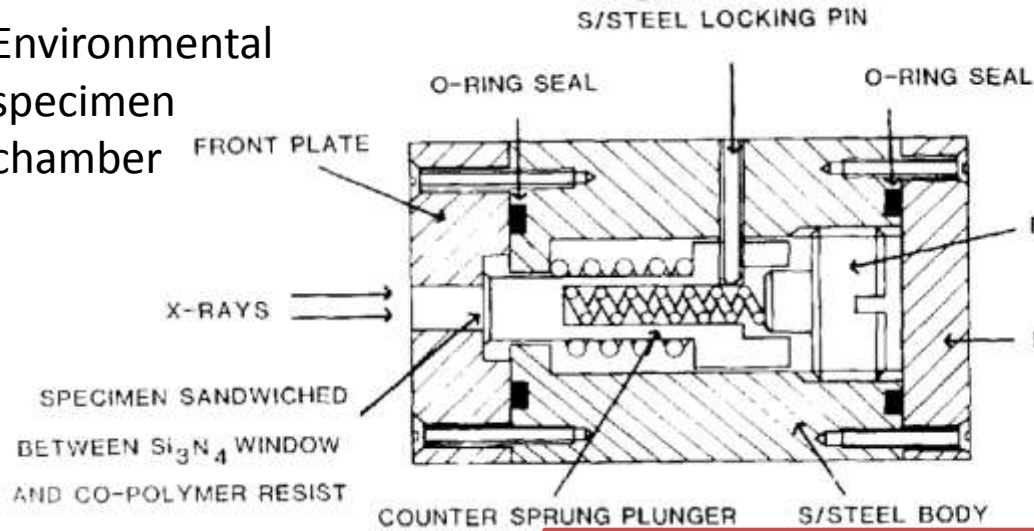


3D internal structure of the GaN quantum dot particle

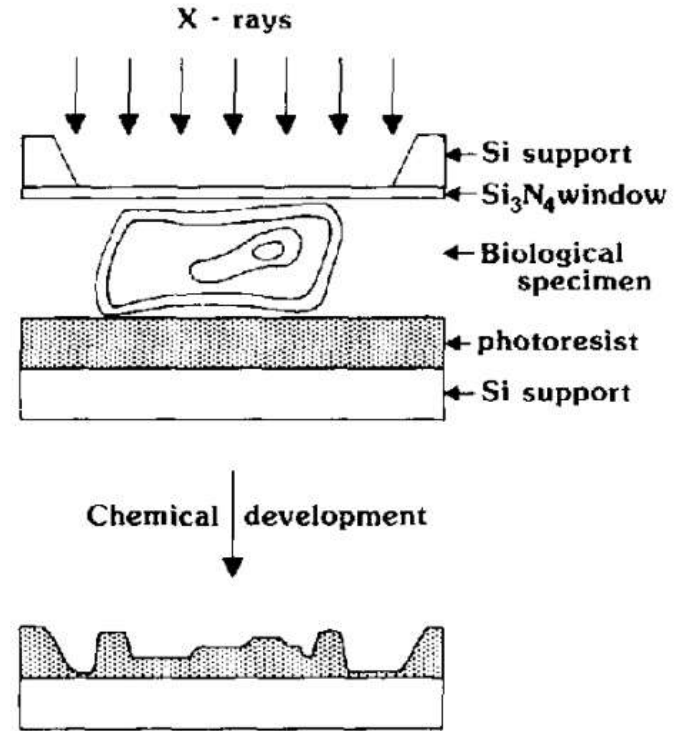


Soft X-ray contact microscopy (SXCM)

Environmental specimen chamber

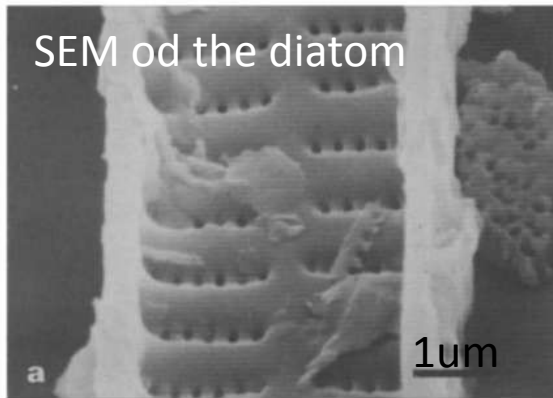


Contact microscopy method

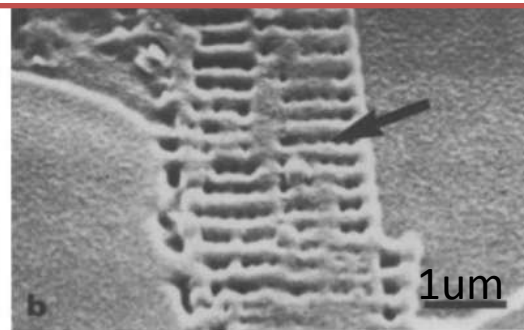


T. W. Ford, et al., Electron Microsc. Rev. 4, 269-292 (1991)

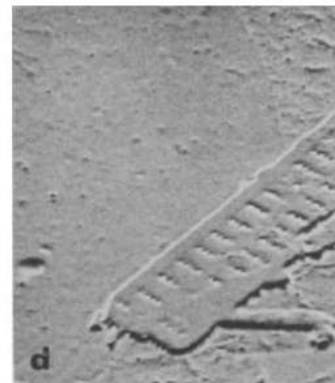
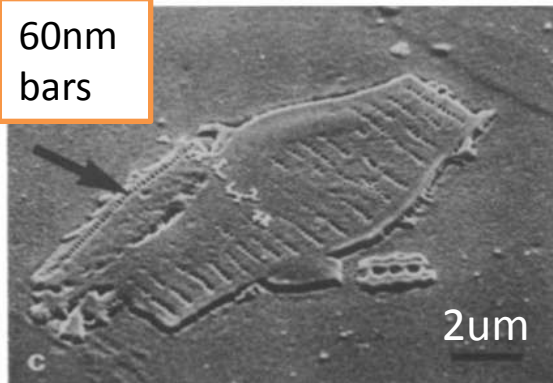
SEM of the diatom



SEM of imprint in polymer



60nm bars

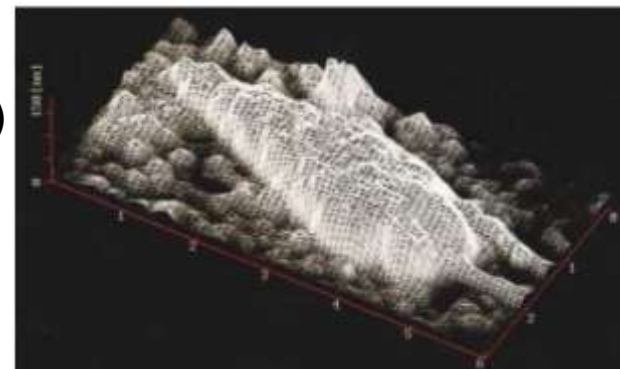


Advantages:

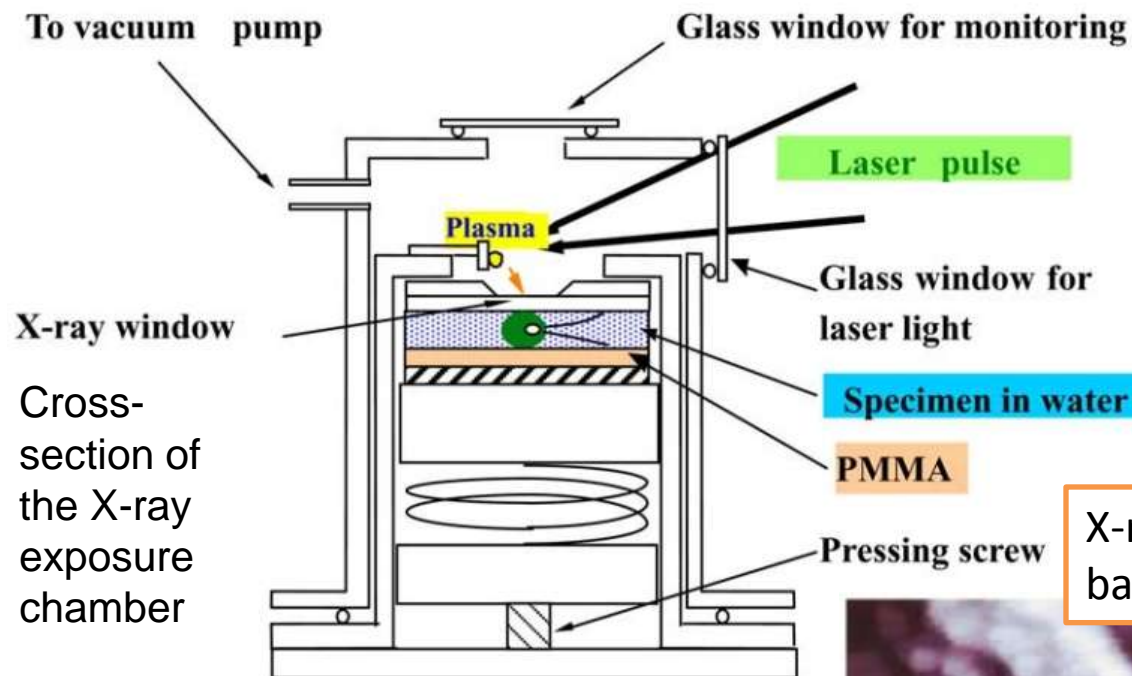
- ✓ the simplest implementation of X-ray microscopy
- ✓ applicable to the large range samples
- ✓ nanoscale resolution and natural contrast
- ✓ no sophisticated optics
- ✓ does not need coherent radiation source
- ✓ possibility of single-shot imaging

Soft X-ray imaging of living cells in water: flash contact soft X-ray microscope

- PMMA photoresist, 40nm spatial resolution
- imaging in water window from LPP source (yttrium target)



X-ray image of a sea-urchin sperm in seawater

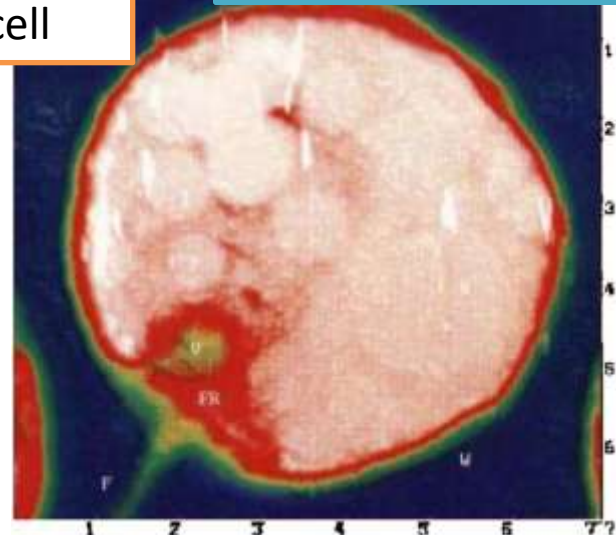


Cross-section of the X-ray exposure chamber

X-ray image of bacterial cell

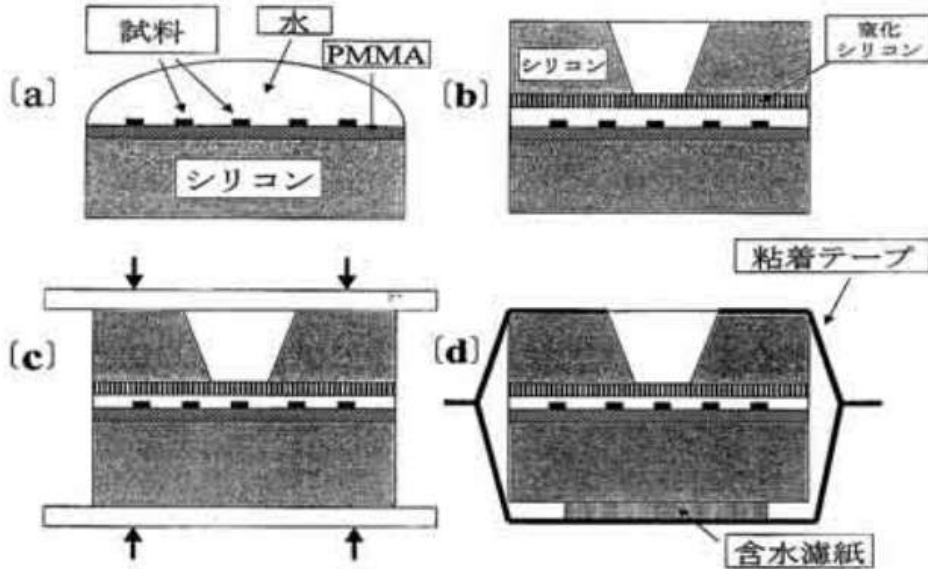


X-ray image of Chlamydomonas



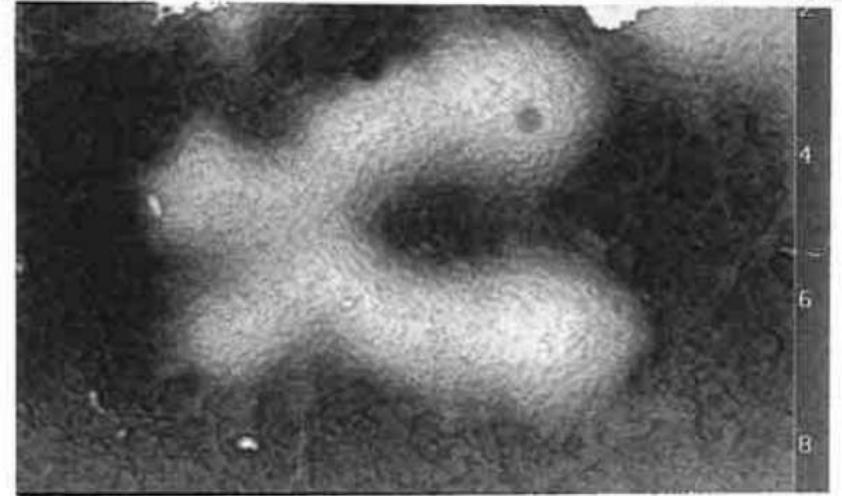
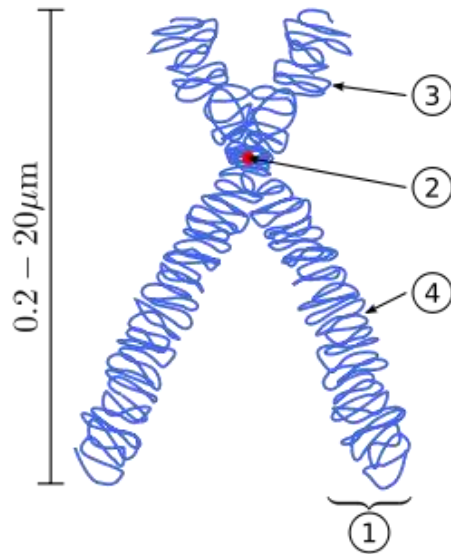
„ The amount of X-ray to make an X-ray image with a spatial resolution of 100 nm is at least more than 10^4 times the dose level that causes biological damage to the specimens”

Imaging of chromosomes using contact microscopy



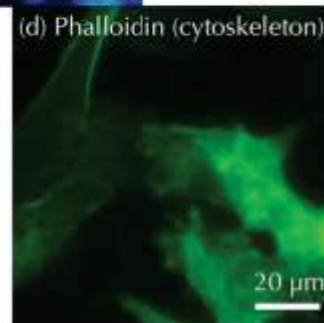
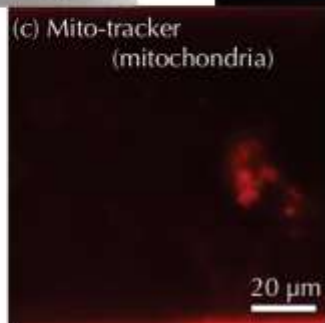
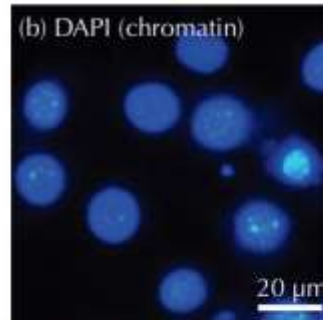
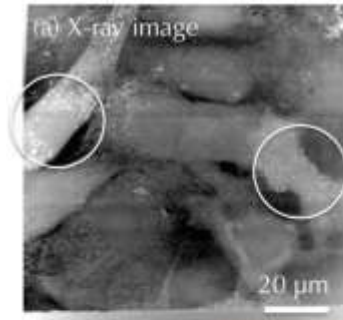
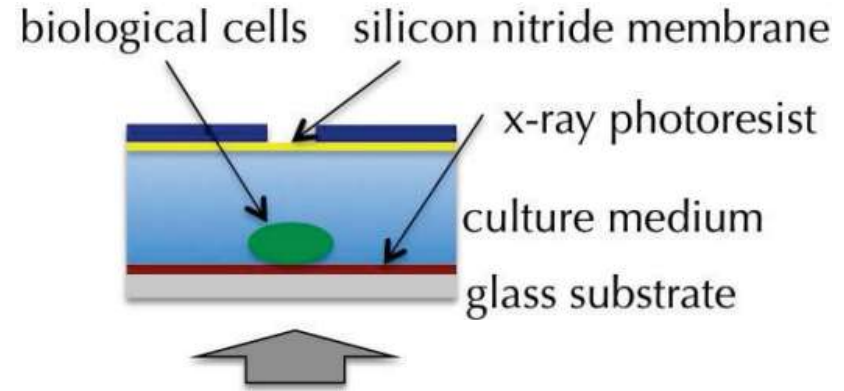
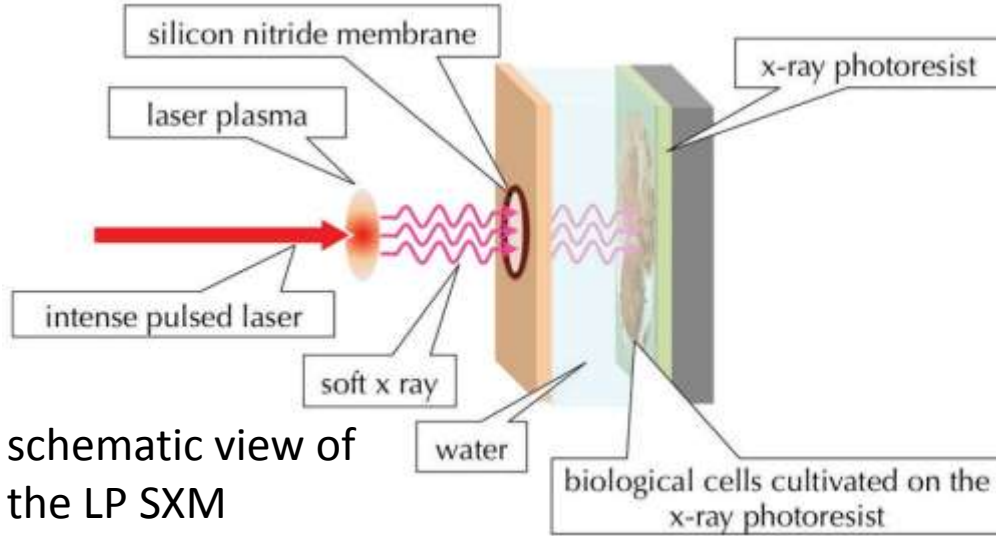
Wet chamber for the sample

<http://en.wikipedia.org/wiki/Chromosome>

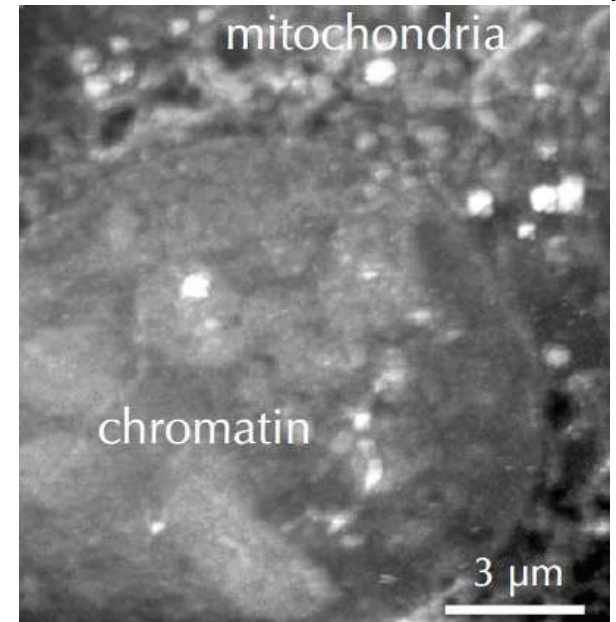


Metaphase chromosomes in the dry state (contact X-ray image)

Imaging of fine structures of cellular organelles in hydrated biological cells by a soft x-ray microscope

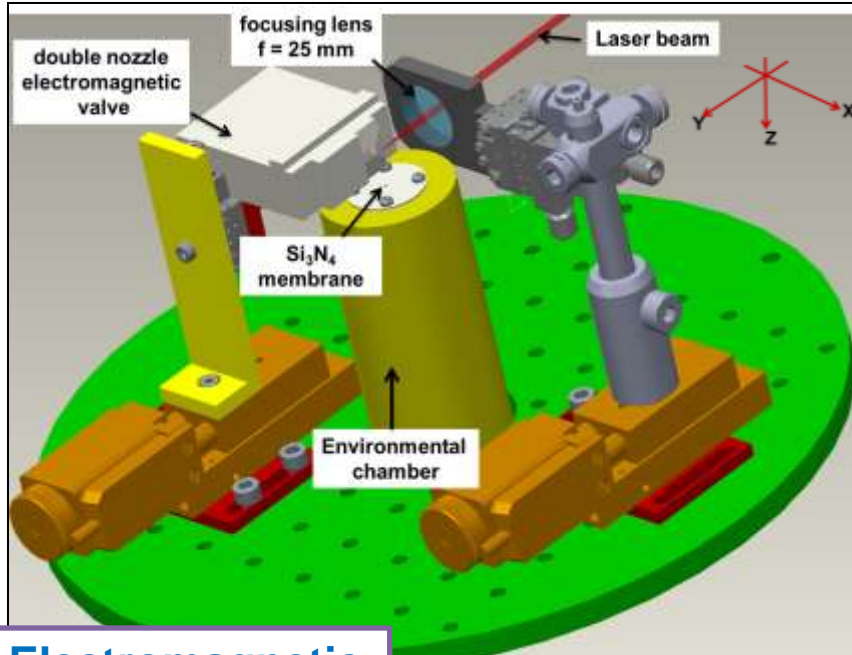


SXR with fluorescence microscope

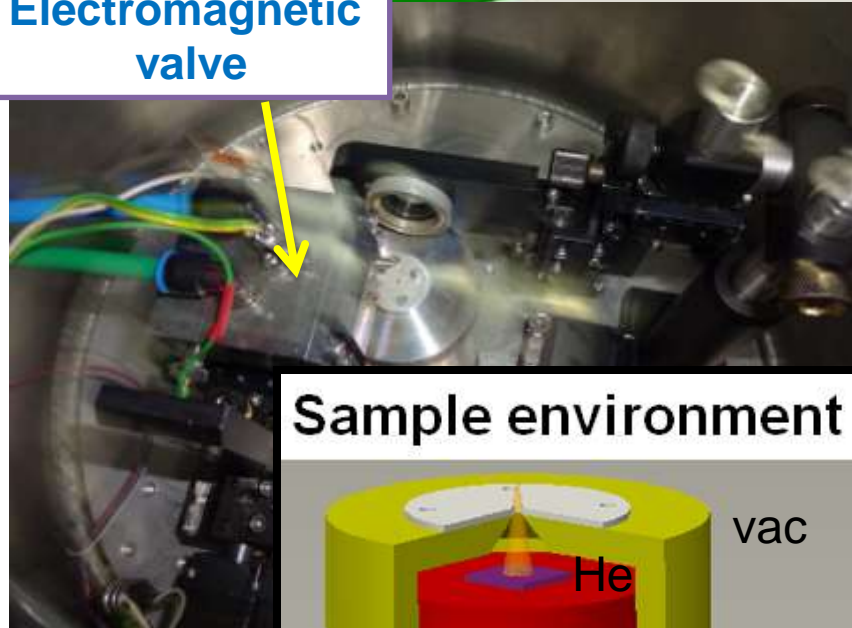


hybrid images of the hydrated biological cells obtained with both of the LPSXM and the fluorescence microscope

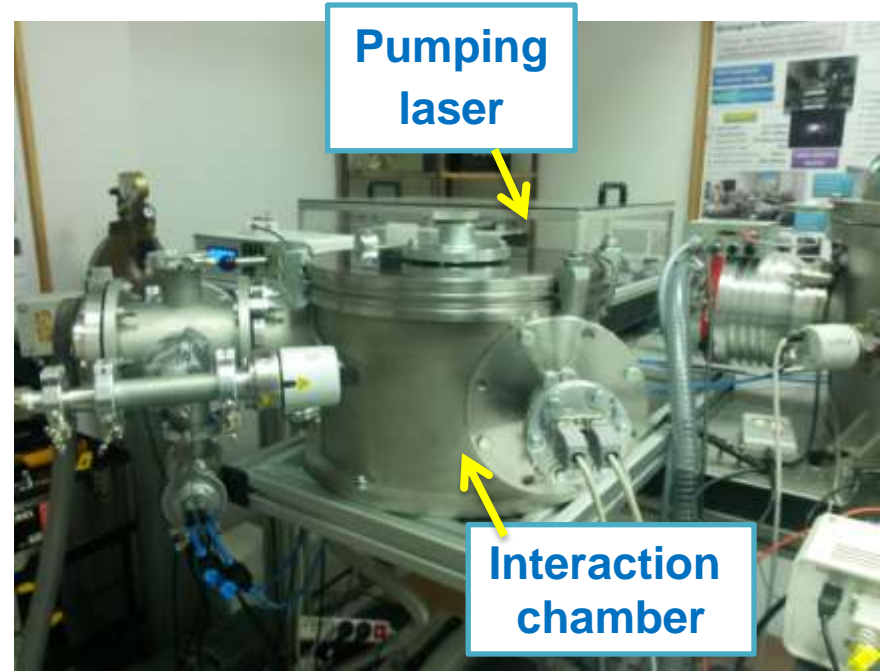
Contact microscopy setup



Electromagnetic valve



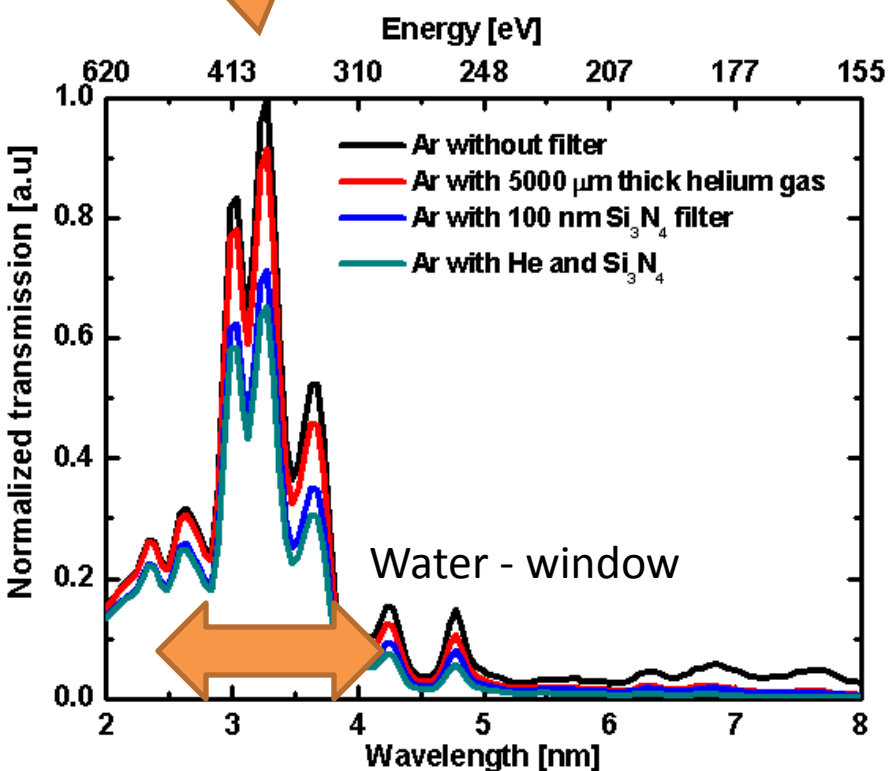
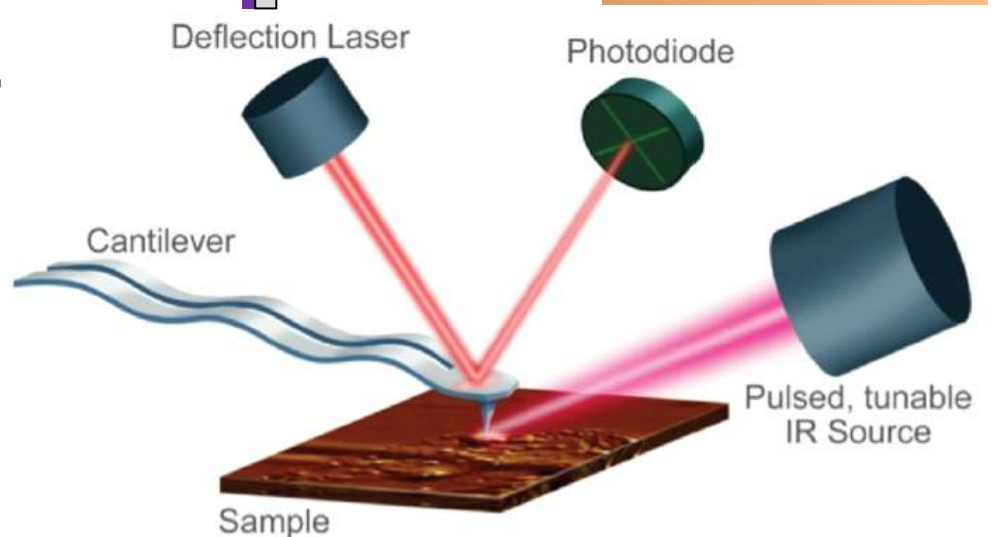
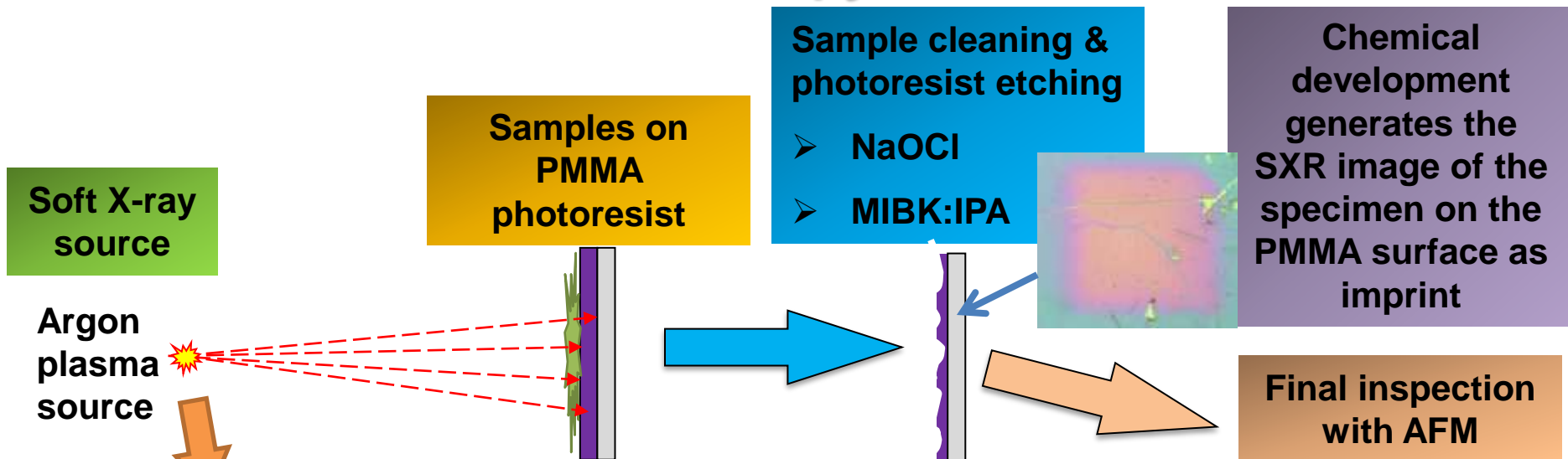
Sample environment



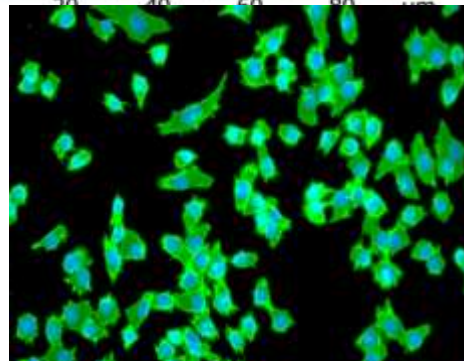
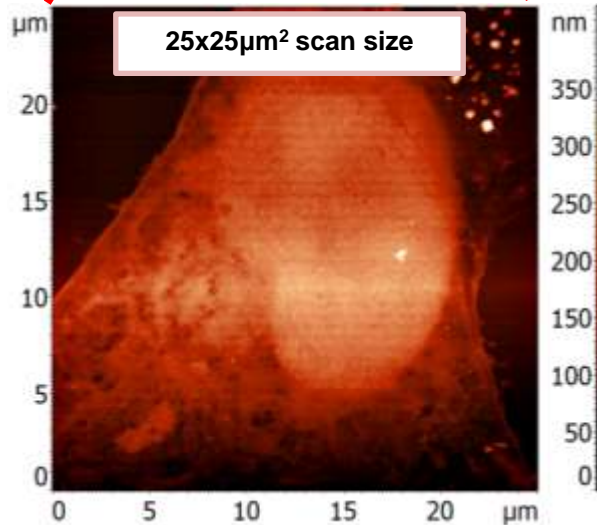
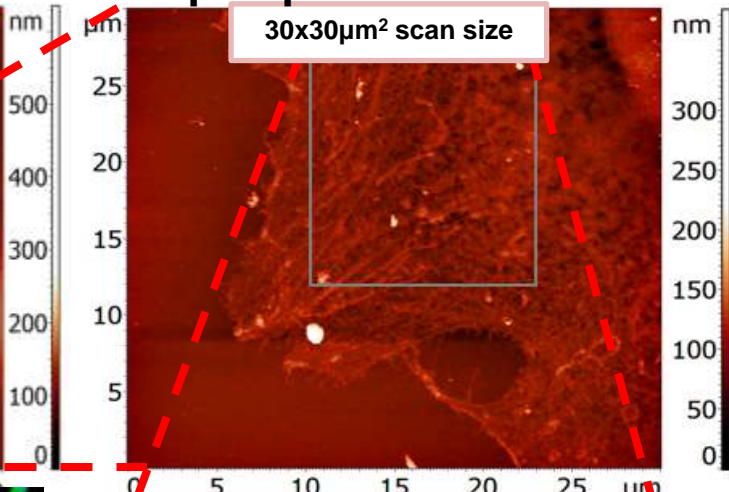
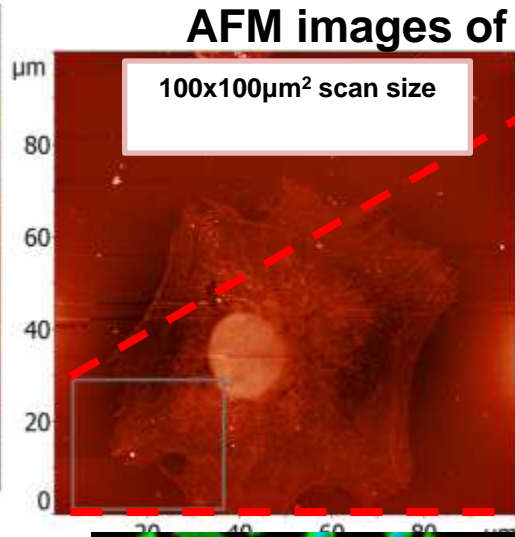
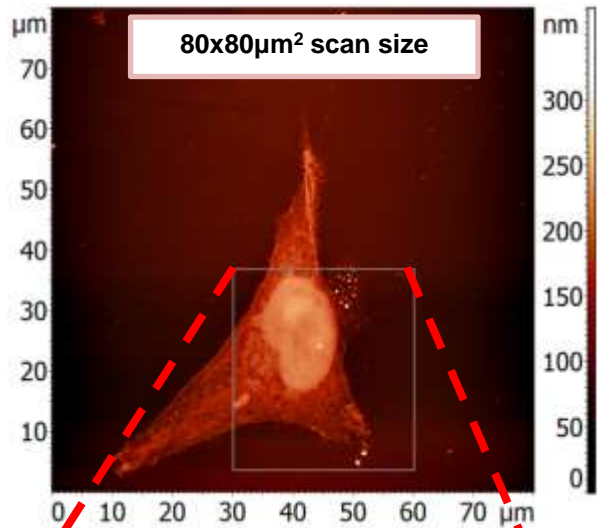
Interaction chamber and components of the source

Pumping laser	Nd:YAG laser (EKSPLA), 740mJ/ 4ns pulse duration, 10 Hz repetition rate, $\lambda = 1064 \text{ nm}$
Double nozzle electromagnetic valve	Inner circular: $\varnothing \sim 0.4 \text{ mm}$ Outer ring: $\varnothing \sim 0.7 \text{ mm} - 1.5 \text{ mm}$
Target gas	Working gas: Ar , Kr, Xe, O ₂ , N ₂ Outer gas : He
Focusing lens	Spherical biconvex lens Focal length: 25 mm

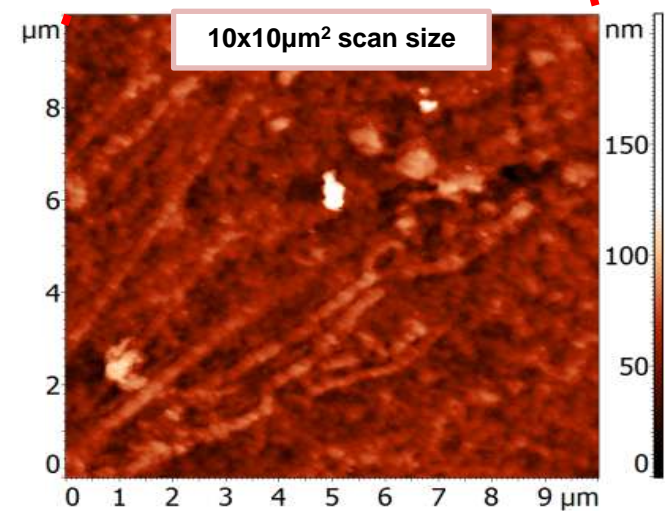
Contact SXR microscopy



Soft X-ray images by contact microscopy of malignant bladder cancer cells (T24)



- Samples stained with Hoechst (Nucleus) and Phalloidin (cytoskeleton)

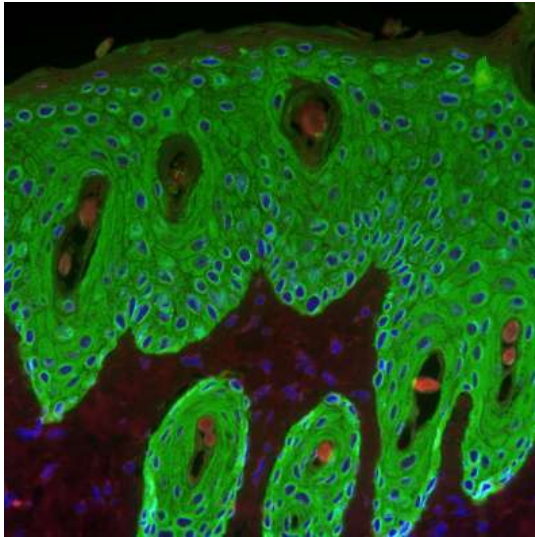


PMMA MW 90k or 350k

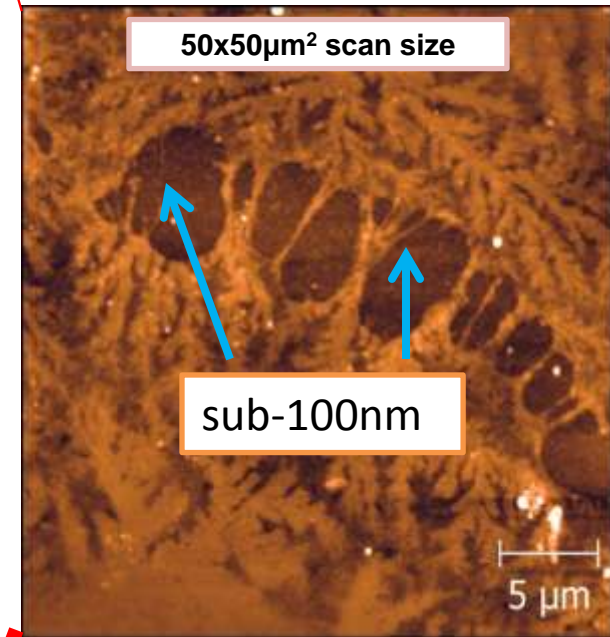
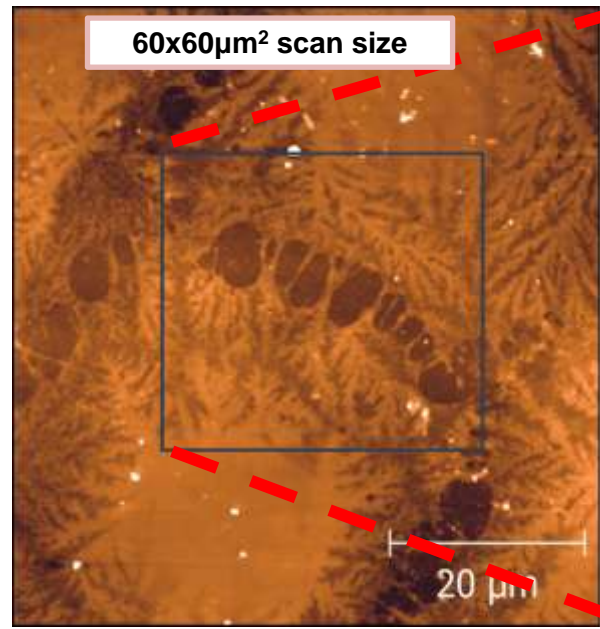
SXCM images of bladder cancer cells (T24),
20 laser shots (2 sec exposure time)

Advantage: during one exposure -> **600x600 μm² imprint.**
If our res. 80nm -> 7.5k x 7.5k (56 Mpix image)

Soft X-ray images by contact microscopy of fixed epidermal cells (Keratinocytes)



Fluorescence microscope image of Keratinocytes



AFM images of a developed photoresist

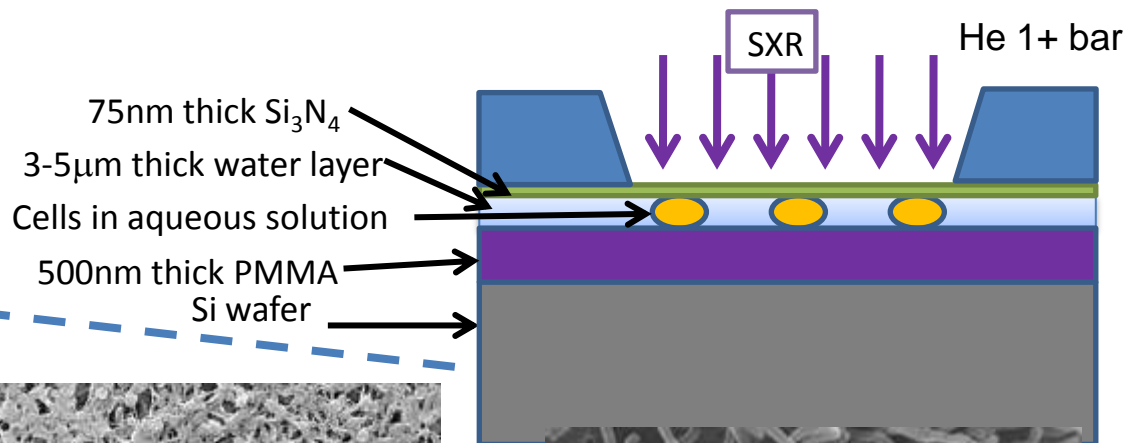
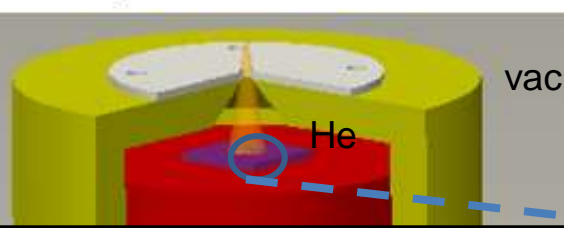
No. of soft X-ray pulses: 200 SXR pulses
Exposure time: 20 sec
Scan : AFM_NDMT
Tip diameter= 20 nm
Scanning mode: semi-contact
No. points/ scan line: 512 pts
Photo resist development time: 90s
Developer concentration: 1:2 v/v (MIBK:IPA)
Knife-edge resolution 80nm (half-pitch)

Mesfin Ayele, EXTATIC PhD student

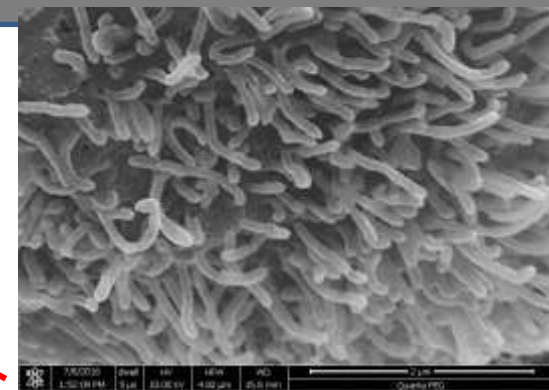
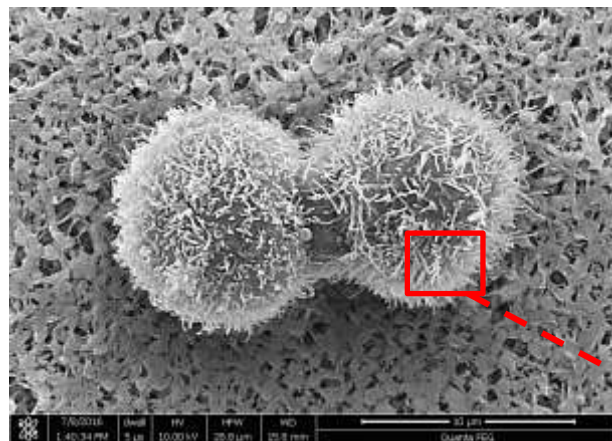


Contact microscopy SXCM – living samples

Sample environment



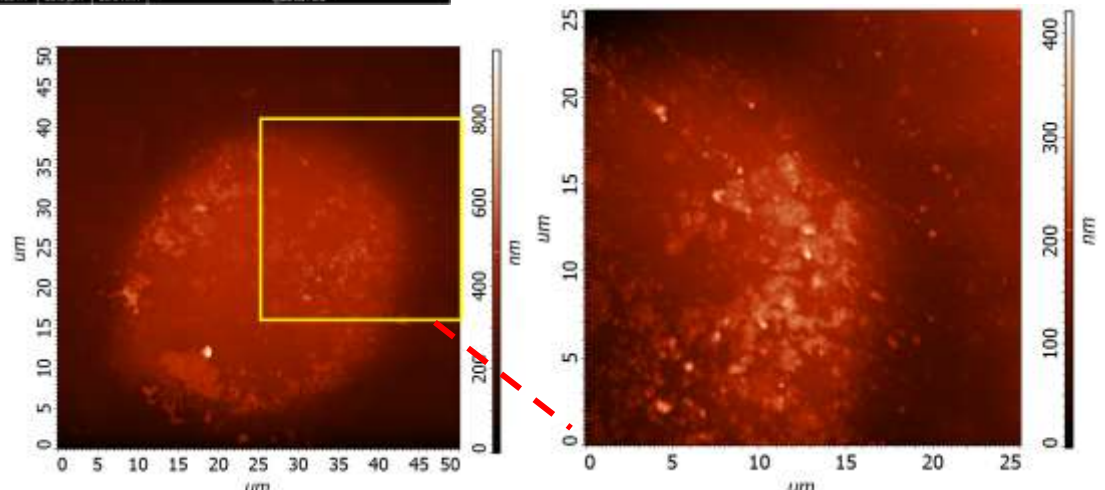
SEM images of the Human prostate cancer cells (DU 145). Scale bar represents 50 µm, 10 µm and 2 µm.



- source emission spectrum: $\lambda=2.7 - 5 \text{ nm}$,
- He pressure inside the environmental chamber: **P=1bar (continuous flow)**
- exposure for wet samples **2000-8000 SXR pulses @ 10Hz**
- Recording medium: **500 nm thick PMMA on Si wafer**
- Postprocessing: PMMA developing procedure: **1:1 MIBK:IPA (3 minutes), AFM scan**

AFM image of **living** prostate cancer cell after the exposure to X-rays in laser plasma based SXCM

Please see: Paulina Osuchowska, et al. „X-ray microscopy imaging versus other microscopic techniques for analysis of human cancer cells morphology” (poster)

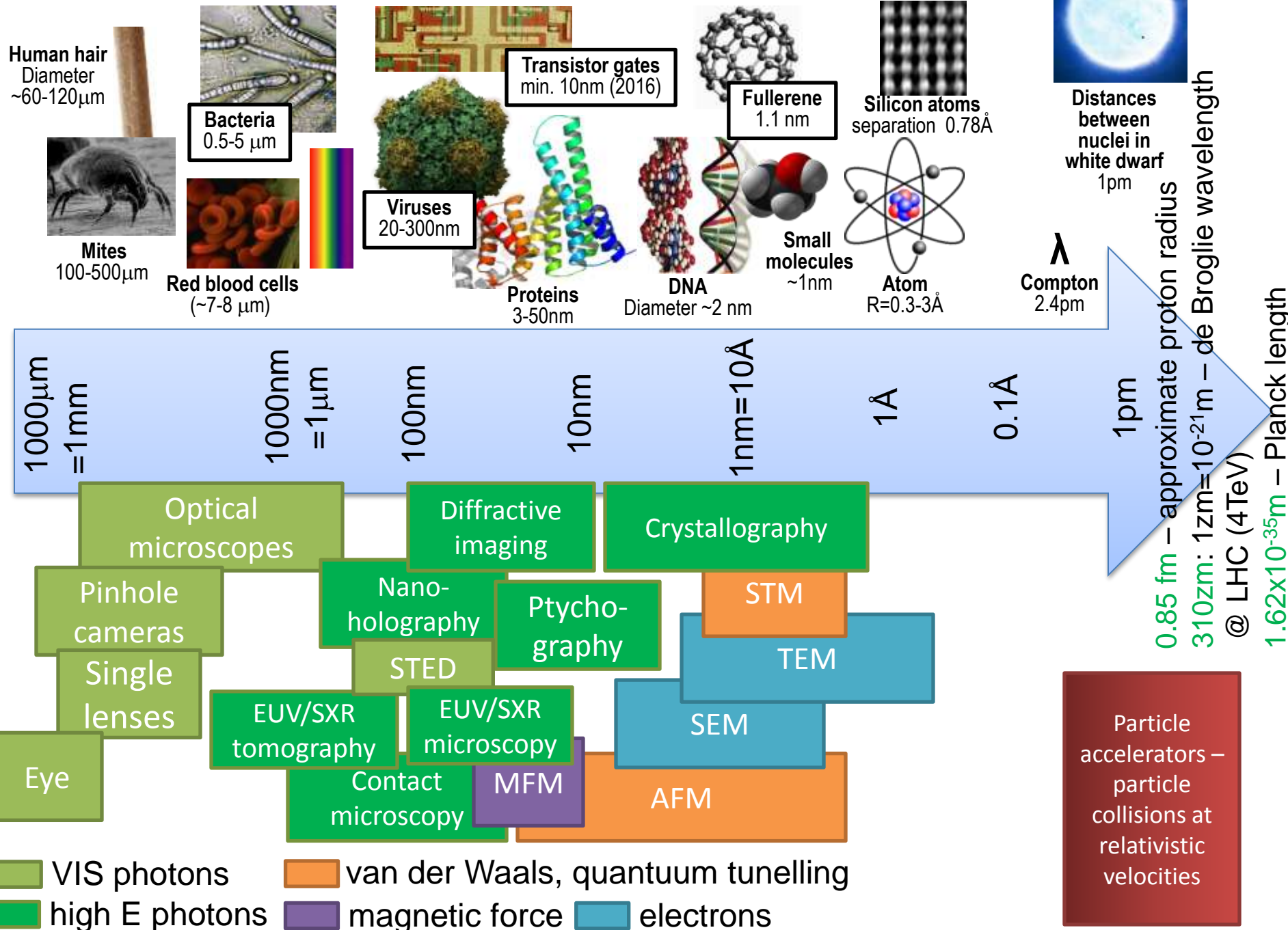


EUV/SXR imaging methods

high energy photons as information carriers

- ✓ EUV holography (Gabor, Fourier)
- ✓ Computer generated holograms
- ✓ Talbot imaging coherent
- ✓ Coherent diffraction (lens-less) imaging
- ✓ Ptychography
- ✓ Zone plates for various wavelengths
- ✓ EUV microscopy using FZP
- ✓ Scanning EUV microscopy better incoherent
- ✓ SXR and „water-window“ microscopy
- ✓ EUV, SXR HXR tomography coh., incoh. are OK
- ✓ Contact microscopy

Spatial scales and imaging methods



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